

JAMIL SALEH

**EFEITO DA TERAPIA LASER DE BAIXA POTÊNCIA NA HIPOSSALIVAÇÃO E  
XEROSTOMIA DECORRENTES DA RADIOTERAPIA**

Dissertação apresentada à Faculdade de Odontologia da Pontifícia Universidade Católica do Rio Grande do Sul como parte dos requisitos para obtenção do título de Mestre em Odontologia, área de concentração em Estomatologia Clínica.

Orientadora: Profa. Dra. Fernanda Gonçalves Salum

Porto Alegre

2014

## ***EPÍGRAFE***

---

*“A tarefa não é tanto ver aquilo que ninguém viu, mas pensar o que ninguém ainda pensou sobre aquilo que todo mundo vê.”*

*Arthur Schopenhauer*

## **AGRADECIMENTOS**

---

## **AGRADECIMENTOS**

Posso neste momento agradecer à **minha família constituída** pela compaixão, dedicação e tudo que a cada dia engrandece nosso convívio. Ter aqueles que nos cercam de carinho como um porto seguro é fundamental para o desenvolvimento do mestrado.

À **Aline** em especial, você faz meu dia feliz. Estes treze anos de cumplicidade me permitem compreender o quanto seu amor foi importante para superar todos os desafios que a vida nos impõe (ou eu invento). Nosso filho chegou para encher ainda mais nosso cantinho. Ter o **Benício** junto da gente é uma alegria sem tamanho. A alegria que ele nos demonstra todo dia me acalma no sentido de dizer, bom, estamos (acredito) no caminho certo.

Agradeço à minha **família de origem** por ter me provido de tudo que precisei até o inicio de minha carreira profissional. Estou certo que do que vivemos só me resta o melhor. Às feridas da vida, só o tempo é remédio.

Agradeço aos meus **colegas de consultórios**, pois sem sua prestatividade e proatividade esta etapa seria ainda mais árdua.

Aos meus **colegas de curso** com os quais pude conviver, trocar experiências e, principalmente, viver novas experiências nas rotinas de ambulatório. Que todos se sintam abraçados. Sinto-me honrado de poder desfrutar de sua companhia. Posso

compreender a importância desta especialidade também pelo que nós experimentamos e proporcionamos como um grupo.

Agradeço ao **SERP**, pela contribuição a minha pesquisa. Especialmente ao **Dr. Aroldo Braga Filho**.

Agradeço aos **colaboradores** do Hospital São Lucas ao permitir que eu interferisse em suas rotinas para a condução de minha pesquisa.

Ao **Serviço de Estomatologia**, pela chance de explorar a especialidade em todos os seus aspectos. Também pelo acolhimento.

À Professora **Fernanda G. Salum** pela dedicação, exemplo, cobrança e apoio. Fico agradecido pela paciência e disposição ao compartilhar seu conhecimento na condução de minhas tarefas.

Às professoras **Maria Antonia** e **Karen Cherubini** pelos ensinamentos e apoio.

## ***RESUMO***

---

## RESUMO

A xerostomia e a hipossalivação são importantes sequelas da radioterapia de cabeça e pescoço e podem ter impacto negativo na qualidade de vida dos pacientes. O presente estudo teve como objetivo realizar uma revisão de literatura destas sequelas e avaliar o efeito da terapia laser de baixa potência (TLBP) na hipossalivação, xerostomia e qualidade de vida relacionada à saúde bucal de pacientes submetidos à radioterapia em cabeça e pescoço. Foram selecionados 23 pacientes com histórico de neoplasia maligna em região de cabeça e pescoço, tratados por meio de teleterapia fracionada, cujo portal tenha envolvido ao menos 50% das glândulas salivares maiores. Todos deveriam ter finalizado a radioterapia há pelo menos seis meses e apresentar hipossalivação e xerostomia. A amostra foi aleatoriamente distribuída em grupos laser ( $n=12$ ) e controle ( $n=11$ ). Foi empregado laser de AsGaAl no comprimento de onda de 830 nm (infravermelho), potência de 100 mW e energia de 2 J por ponto. Os pacientes foram submetidos a doze sessões de TLBP, aplicada pontualmente em ambas as glândulas parótidas, submandibulares e sublinguais. No grupo-controle foi adaptado ao aparelho de laser um dispositivo que impedia a emissão de radiação. O fluxo salivar em repouso e sob estímulo foi avaliado em *baseline*, após a 6<sup>a</sup> e a 12<sup>a</sup> sessão de TLBP. A xerostomia foi avaliada por meio de Escala Visual Analógica (EVA) e a qualidade de vida relacionada à saúde bucal, por meio do instrumento *Oral Health Impact Profile* (OHIP-14). Os resultados demonstraram não haver diferença significativa entre os grupos laser e controle quanto à velocidade do fluxo salivar, xerostomia ou qualidade de vida. Por outro lado, em ambos os grupos ao final do tratamento, houve redução significativa da xerostomia e melhora da qualidade de vida.

relacionada à saúde oral. Com base nos resultados, pode-se concluir que a TLBP, nos parâmetros utilizados, não foi capaz de promover elevação clinicamente detectável do fluxo salivar ou redução da xerostomia. Os resultados podem estar associados aos efeitos tardios da radioterapia na estrutura glandular tais como fibrose e atrofia acinar. A melhora dos índices subjetivos xerostomia e qualidade de vida em ambos os grupos ressalta a importância da orientação e acompanhamento dos pacientes irradiados.

Palavras-chave: Saliva. Xerostomia. Radioterapia. Terapia a laser. Qualidade de vida.

***ABSTRACT***

---

## ABSTRACT

Xerostomia and hyposalivation are important sequelae of head and neck radiotherapy and may have a negative impact on quality of life. The present study aimed to evaluate the effects of low level laser therapy (LLLT) on hyposalivation, xerostomia and quality of life related to oral health (QLROH) in irradiated patients. Were selected 23 patients with a history of head and neck malignancy, treated with fractioned teletherapy, whose therapeutic site had involved at least 50% of the major salivary glands. Patients should have completed radiotherapy for at least six months and present hyposalivation and xerostomia. The sample was randomly distributed in laser group (n=12) and control (n=11). A GaAlAs laser, at 830 nm (infrared) wavelength, 100 mW power, 2 J energy per point was used. Patients underwent twelve sessions of LLLT, applied in parotid, submandibular and sublingual glands. In the control group, the laser tool received a plastic tip that blocked radiation emission. Stimulated and unstimulated salivary flow rate was assessed at baseline, after the 6th and 12th session. Xerostomia was assessed by Visual Analogue Scale (VAS) and QLROH, through the Oral Health Impact Profile (OHIP - 14). The results showed no significant difference between the laser and control groups regarding the salivary flow rate, xerostomia or quality of life. However, at the end of the treatment, the xerostomia and the QLROH showed significant improvement in both groups compared to assessments carried out in baseline. Based on the results, we conclude that, in the parameters used, LLLT was not able to increase salivary flow rate or decrease xerostomia. The results may be associated to the late effects of radiotherapy on glandular structure such as fibrosis and acinar atrophy. The

improvement in xerostomia and quality of life highlights the importance of advice given to the irradiated patients and their follow-up.

Keywords: Saliva. Xerostomia. Radiotherapy. Low level laser therapy. Quality of life.

## ***LISTA DE ILUSTRAÇÕES***

---

## **LISTA DE ILUSTRAÇÕES**

Figue 1. Flowchart representing the stages of the study.....78

## ***LISTA DE TABELAS***

---

## **LISTA DE TABELAS**

### **SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS**

Table 1	Diagnostic methods of salivary dysfunctions.....	36
Table 2	Drugs that cause salivary dysfunctions.....	39
Table 3	Systemic disorders associated with salivary dysfunctions.....	42

### **EFFECT OF LOW-LEVEL LASER THERAPY ON RADIOTHERAPY-INDUCED HYPOSALIVATION AND XEROSTOMIA: A PILOT STUDY**

Table 1	Demographic distribution of the patients and characteristics of the treatment within the groups studied .....	81
Table 2	Visual analogic scale (VAS) scores for xerostomia in the laser group and control group in baseline, after the 6 <sup>th</sup> and 12 <sup>th</sup> LLLT sessions.....	82
Table 3	Stimulated and unstimulated salivary flow rate (SFR) (mL/min) in the laser group and control group in baseline, after the 6 <sup>th</sup> and 12 <sup>th</sup> LLLT sessions.....	83
Table 4	Oral health impact profile (OHIP-14) scores for quality of life related to oral health assessment in the laser group and control group in baseline, after the 6 <sup>th</sup> and 12 <sup>th</sup> LLLT sessions.....	83

---

***LISTA DE ABREVIATURAS, SIGLAS E SÍMBOLOS***

## **LISTA DE ABREVIATURAS, SIGLAS E SÍMBOLOS**

<b>ANTT</b>	Anethole Trithione
<b>AsGa</b>	Arseneto de Gálio
<b>AsGaAI</b>	Aluminum-arsenide-gallium
<b>ATP</b>	Adenosina Trifosfato
<b>CMC</b>	Carboxymethylcellulose
<b>CT</b>	Computed Tomography
<b>EVA</b>	Escala Visual Analógica
<b>FDG-PET-CT</b>	Fluorodeoxyglucose-labeled Positron Emission Tomography-computed Tomography
<b>Gy</b>	Gray
<b>hAQP1</b>	Human Aquaporine-1
<b>HIV</b>	Human Immunodeficiency Virus
<b>IMRT</b>	Intensity Modulated Radiotherapy
<b>InGaAIP</b>	Aluminum-gallium-indium-phosphide
<b>KGF</b>	Keratinocyte Growth Factor
<b>LLLT</b>	Low Level Laser Therapy
<b>MnSOD-PL</b>	Manganese Superoxide Dismutase-plasmid/liposomes
<b>MR</b>	Magnetic Resonance
<b>OHIP-14</b>	Oral Health Impact Profile
<b>QoL H&amp;N35</b>	Quality of Life Head and Neck 35
<b>QLROH</b>	Quality of Life Related to Oral Health
<b>TLBP</b>	Terapia Laser de Baixa Potência
<b>TLK1B</b>	Tousled-like kinase 1B

<b>SMA</b>	Smooth Muscle Actin
<b>SFR</b>	Salivary Flow Rate
<b>VFS</b>	Velocidade de Fluxo Salivar
<b>VAS</b>	Visual Analogic Scale

## **SUMÁRIO**

---

## SUMÁRIO

<b>1</b>	<b>INTRODUÇÃO.....</b>	<b>24</b>
<b>2</b>	<b>PROPOSIÇÃO.....</b>	<b>30</b>
<b>2.1</b>	<b>Objetivo Geral.....</b>	<b>30</b>
<b>2.2</b>	<b>Objetivos Específicos.....</b>	<b>30</b>
<b>3</b>	<b>SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS.....</b>	<b>32</b>
	<i>ABSTRACT.....</i>	34
	<i>INTRODUCTION.....</i>	34
	<i>DIAGNOSIS OF SALIVARY DYSFUNCTIONS.....</i>	36
	<i>ETIOLOGY OF SALIVARY DYSFUNCTIONS.....</i>	38
	<i>DRUGS.....</i>	38
	<i>AGING.....</i>	40
	<i>SYSTEMIC DISORDERS.....</i>	41
	<i>Sjögren Syndrome.....</i>	43
	<i>HIV Infection.....</i>	43
	<i>Graft-versus-host disease.....</i>	44
	<i>RADIOTHERAPY AND CHEMOTHERAPY.....</i>	44
	<i>THERAPEUTIC OPTIONS.....</i>	47
	<i>PREVENTIVE THERAPIES.....</i>	47
	<i>DISEASE-MODIFYING AGENTS.....</i>	49
	<i>SYMPOMATIC THERAPIES.....</i>	49
	<i>TOPICAL AND SYSTEMIC STIMULANTS.....</i>	50
	<i>REGENERATIVE THERAPIES.....</i>	52
	<i>DISCUSSION.....</i>	52
	<i>REFERENCES.....</i>	54
<b>4</b>	<b>EFFECT OF LOW-LEVEL LASER THERAPY ON RADIOTHERAPY-INDUCED HYPOSALIVATION AND XEROSTOMIA - A PILOT STUDY.....</b>	<b>73</b>
	<i>ABSTRACT.....</i>	75

<i>INTRODUCTION.....</i>	76
<i>PATIENTS AND METHODS.....</i>	77
<i>Xerostomia and Salivary Flow Rate (SFR).....</i>	79
<i>Quality of Life Related to Oral Health (QLROH).....</i>	79
<i>Low Level Laser Therapy LLLT).....</i>	79
<i>STATISTICAL ANALYSIS.....</i>	80
<i>RESULTS.....</i>	81
<i>Characterization of the sample.....</i>	81
<i>Xerostomia and Salivary Flow Rate (SFR).....</i>	82
<i>Quality of Life Related to Oral Health (QLROH).....</i>	83
<i>DISCUSSION.....</i>	84
<i>CONCLUSIONS.....</i>	87
<i>REFERENCES.....</i>	88
<b>5     DISCUSSÃO GERAL.....</b>	<b>94</b>
<b>6     REFERÊNCIAS.....</b>	<b>99</b>
<b>Anexo A.....</b>	<b>106</b>
<b>Anexo B.....</b>	<b>107</b>
<b>Anexo C.....</b>	<b>110</b>
<b>Anexo D.....</b>	<b>111</b>
<b>Anexo E.....</b>	<b>112</b>
<b>Anexo F.....</b>	<b>113</b>

## ***1 INTRODUÇÃO***

---

## 1 INTRODUÇÃO

O tratamento das neoplasias malignas da região de cabeça e pescoço é multimodal e pode ser realizado por meio de ressecção cirúrgica, radioterapia e/ou quimioterapia. A radioterapia consiste na utilização de doses elevadas de radiação ionizante, que interagem com os tecidos tumorais, atuando sobre o DNA nuclear por meio da produção de radicais livres, o que causa morte ou incapacidade de replicação celular. Sua ação sobre os tecidos não é seletiva, atuando também em células saudáveis, o que a torna tóxica para o organismo (SEGRETO; SEGRETO, 2000). Em neoplasias de cabeça e pescoço, geralmente, a dosimetria de radiação varia de 50 a 70 Gy, fracionada em doses de 2 Gy ao dia (SEIKALY et al., 2004). Atualmente tem sido empregada a técnica de radioterapia de intensidade modulada, de forma que a dose de radiação seja mais intensa na área do tumor, preservando as estruturas adjacentes (BHIDE et al., 2012).

Apesar dos benefícios em neoplasias malignas de cabeça e pescoço, a radioterapia, geralmente, apresenta efeitos adversos tais como alteração da microflora bucal, processos degenerativos e inflamatórios da mucosa, xerostomia, risco aumentado ao desenvolvimento de cáries e de outras doenças infecciosas, osteorradiacionecrose, trismo, dentre outros (JHAM; FREIRE, 2006; SENNHENN-KIRCHNER et al., 2009; CHREANOVIC et al., 2010; WEBER et al., 2010). Os efeitos adversos imediatos ocorrem durante ou logo após a conclusão da radioterapia e regredem com o tempo. Em contraste, os efeitos tardios são, geralmente, considerados irreversíveis e progressivos (JELLEMA et al., 2007; CHOPRA et al., 2011).

As glândulas salivares maiores são frequentemente envolvidas nos portais terapêuticos de radiação por estarem em proximidade com os sítios de tumores primários e cadeias linfáticas da região de cabeça e pescoço. Como consequência, passam por um processo de degeneração, resultando em hipossalivação e xerostomia (EISBRUCH et al., 2001). A gravidade dessas alterações é determinada por fatores como dose de radiação, quantidade de tecido salivar exposto e resposta individual do paciente (DIRIX et al., 2008; LIN et al., 2008). Clinicamente observa-se redução do volume de saliva, que se apresenta espessa, dispersa e localizada na região posterior do assoalho da boca e vestíbulo inferior, além disso, a mucosa torna-se ressecada e atrófica e o dorso da língua despapilado. Os pacientes podem apresentar disgeusia, disfagia e disartria, ulcerações na mucosa bucal (PORTER; SCULLY; HEGARTY, 2004), risco elevado do surgimento de doenças infecciosas tais como cáries, gengivite, periodontite e candidose (PORTER; FEDELE; HABBAB, 2010).

Em tratamentos radioterápicos convencionais, a xerostomia inicia-se a partir da primeira semana. Os danos passam a ser irreversíveis após doses cumulativas de 26 a 39 Gy, muitas vezes com volumes salivares inferiores a 10% do apresentado previamente à radioterapia (PORTER; FEDELE; HABBAB, 2010; VISSINK et al., 2010). As alterações glandulares iniciam-se pelo dano à membrana plasmática, com perda da resposta aos controles autonômicos, há edema, degeneração e necrose das células acinares. Os efeitos tardios são consequência da fibrose e atrofia dos lóbulos (PORTER, 2010). A saliva resultante sofre alterações qualitativas em suas propriedades orgânicas e inorgânicas, com diminuição da atividade das amilases, da capacidade tampão e do pH, com consequente acidificação. Há elevação dos níveis de cálcio, potássio, sódio e redução na

concentração de fosfato (JHAM; FREIRE, 2006). As alterações do fluxo e viscosidade salivares podem persistir por anos e a recuperação dependerá das características da cada paciente (LOPES; MAS; ZÂNGARO, 2006).

Conceitualmente, a xerostomia é a sensação subjetiva de boca seca, enquanto a hipossalivação consiste na redução objetiva do fluxo salivar. Atkinson, Grisius e Massey (2005) classificam os tratamentos da hipossalivação e xerostomia em: 1) preventivos; (2) sintomáticos; (3) estimulantes tópicos e sistêmicos; (4) agentes modificadores de doença e (5) regeneradores. Pode-se destacar o uso de gomas de mascar, de sialagogos sistêmicos, estimulação elétrica, acupuntura, substitutos da saliva, toxina botulínica, transferência de glândula salivar entre outros tratamentos citados na literatura (SEIKALY et al., 2004; JHAM et al., 2007; JELLEMA et al., 2007; MÜNTER, et al., 2007; TEYMOORTASH et al., 2009; JENSEN et al., 2010; RIEGER, 2012).

A terapia laser de baixa potência (TLBP) ou fototerapia laser é um método simples, podendo ser utilizado como adjuvante a tratamentos convencionais ou de forma isolada e eletiva em algumas doenças (BRUGNERA-JUNIOR; PINHEIRO, 1998; LOPES; MAS; ZÂNGARO, 2006; SIMÕES et al., 2008; LONCAR et al., 2011; JENKINS; CARROL, 2011). A TLBP possui baixa energia e não apresenta potencial fototérmico, sendo utilizada por seus efeitos anti-inflamatórios, analgésicos e biomoduladores (LOPES; MAS; ZÂNGARO, 2006; SIMÕES et al., 2008; ANKRI; LUBART; TAITELBAUM, 2010). O efeito resultante da TLBP baseia-se na capacidade de modulação de diversos processos metabólicos, bioquímicos e fotofísicos, que transformam a luz laser em energia útil para a célula. Esta energia provoca reações nas mitocôndrias, com incremento na produção de ATP (Adenosina

Trifosfato), aumento no consumo de glicose pelas células, elevação dos níveis intracelulares de cálcio e do número de mitoses (LOPES; MAS; ZÂNGARO, 2006).

O efeito da TLBP para o tratamento da xerostomia tem sido investigado em pesquisas pré-clínicas e clínicas (LOPES; MAS; ZÂNGARO, 2006; SIMÕES et al., 2008; LONCAR et al., 2011). Simões et al. (2008) demonstraram elevação no fluxo e na concentração total de proteínas salivares de ratos *Wistar* quando submetidos a TLBP nas glândulas salivares maiores. Neste estudo foi empregado laser de diodo, no comprimento de onda de 808 nm, com dosimetrias de 4 e 8 J/cm<sup>2</sup>. Simões et al. (2010), em um estudo clínico realizado com pacientes irradiados em cabeça e pescoço, empregaram a TLBP para prevenção e tratamento da mucosite oral. Os pacientes submetidos a três aplicações semanais apresentaram redução da xerostomia e elevação do fluxo salivar estimulado. Os autores sugerem que a laserterapia pode ser empregada como adjuvante no tratamento de alterações das glândulas salivares decorrentes da radioterapia. Ao analisarem o efeito da TLBP em pacientes com mucosite, Cowen et al. (1997) também relataram tais achados, os pacientes apresentaram aumento da produção de saliva e da habilidade de deglutição. Loncar et al. (2011) utilizaram o laser de GaAs (Arseneto de Gálio) (904nm), com densidade de energia de 29,5 J/cm<sup>2</sup>, em 34 pacientes com xerostomia durante 10 dias consecutivos. Os autores verificaram que a laserterapia promoveu aumento do fluxo salivar. Ao avaliarem o efeito da TLBP sobre a velocidade do fluxo salivar em pacientes irradiados, Lopes, Mas e Zângaro (2006) observaram que a velocidade do fluxo salivar manteve-se significativamente superiores nos pacientes que receberam a TLBP em comparação aos controles.

A xerostomia e hipossalivação são importantes sequelas da radioterapia e podem ter impacto negativo na qualidade de vida dos pacientes. Há estudos

demonstrando que a TLBP pode ter efeito benéfico no tratamento dessas alterações. O presente estudo teve como objetivo avaliar clinicamente o efeito da TLBP na hipossalivação e xerostomia decorrentes da radioterapia em região de cabeça e pescoço. Além disso, foi avaliada a influência desta modalidade terapêutica na qualidade de vida relacionada à saúde bucal dos pacientes.

---

***2 PROPOSIÇÃO***

## **2 PROPOSIÇÃO**

### **2.1 Objetivo Geral**

Realizar uma revisão da literatura sobre a hipossalivação e xerostomia e avaliar clinicamente o efeito da terapia laser de baixa potência (TLBP) no tratamento destas sequelas em decorrência da radioterapia em região de cabeça e pescoço.

### **2.2 Objetivos Específicos**

Avaliar em pacientes submetidos à radioterapia em região de cabeça e pescoço:

- O efeito da TLBP na hipossalivação, pela avaliação da velocidade do fluxo salivar em repouso e sob estimulação.
- O efeito da TLBP na xerostomia, por meio de Escala Visual Analógica (EVA).
- Se a TLBP, utilizada para o tratamento da hipossalivação e xerostomia, exerce influência na qualidade de vida relacionada à saúde bucal.

---

**3 ARTIGO DE REVISÃO DA LITERATURA**

### **3 ARTIGO DE REVISÃO DA LITERATURA**

#### **SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS**

Artigo submetido para avaliação (Anexo E)

Periódico: Archives of Oral Biology

Qualis Capes Odontologia 2013: A2

Fator de Impacto: 1,549

**SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND  
THERAPEUTICS**

SALEH, Jamil

FIGUEIREDO, Maria Antonia Zancanaro

CHERUBINI, Karen

SALUM, Fernanda Gonçalves

**Oral Medicine Division, Pontifical Catholic University of Rio Grande do Sul-  
PUCRS, Brazil.**

**Corresponding address:**

Fernanda Gonçalves Salum

Pontifícia Universidade Católica do Rio Grande do Sul - PUCRS

Hospital São Lucas

Av. Ipiranga, 6690 – Room 231

CEP: 90610-000 - Porto Alegre – RS – Brazil

Tel/Fax: +55 51 3320-3254

E-mail: [fernanda.salum@pucrs.br](mailto:fernanda.salum@pucrs.br)

## ABSTRACT

Saliva is of paramount importance for the maintenance of oral and general homeostasis. Salivary hypofunction predispose patients to disorders such as dysgeusia, pain and burning mouth, caries and other oral infectious diseases, dysphagia and dysphonia. The aim of this study was to provide an update on the etiology, diagnostic methods and therapeutic strategies for the management of hyposalivation and xerostomia. The present paper describes subjective and objective methods for the diagnosis of salivary dysfunctions; moreover a number of drugs and systemic disorders associated with decreased salivary flow rate are listed. We also focused on the underlying mechanisms to radiotherapy-induced salivary damage. Therapeutics for hyposalivation and xerostomia were discussed and classified as preventive, symptomatic, topical and systemic stimulants, disease-modifying agents, and regenerative. New therapeutic modalities have been studied and involve stem cells transplantation, with special attention to regeneration of damage caused by ionizing radiation to the salivary glands. More studies in this area are needed to provide new perspectives in the treatment of patients with salivary dysfunctions.

**Key words:** salivary glands, xerostomia, radiotherapy, therapeutics, drug therapy.

## INTRODUCTION

Saliva is of paramount importance for the maintenance of oral and general homeostasis. It displays a crucial role in the digestive function, taste, cleaning, hydration of the oral mucosa, and protection of the teeth, due to buffering and remineralization properties.<sup>1,2</sup> Besides, saliva controls the composition of the oral

microflora due to antibacterial, antifungal and antiviral properties, protecting the body from deleterious extrinsic influences.<sup>3-6</sup>

Saliva is composed of more than 99% water along with electrolytes; the protein components include immunoglobulins, digestive enzymes such as amylase and lipase, and antibacterial and antifungal enzymes, as well as mucins.<sup>1,2,7</sup> Salivary secretion is controlled by the autonomous nervous system, mainly by parasympathetic nerve signals.<sup>7,8</sup> About 90% of saliva is produced by the major salivary glands<sup>3,4</sup> and the daily volume varies from 0.5 to 1.0 L.<sup>6,8</sup> When at rest, 65% of saliva is produced by the submandibular glands, which produce saliva rich in mucin, which supplies lubrication for the mucosa.<sup>2,7,9</sup> Under stimulation, the parotids account for 50% of salivary volume.<sup>7-9</sup>

Nederfors<sup>10</sup> suggests that salivary dysfunctions can be divided into three aspects: xerostomia, as subjective alteration; hyposalivation, as objective reduction of salivary flow and alterations in salivary composition. In early stages, hyposalivation is characterized by decreased salivary volume, besides saliva is thick and dispersed. The oral mucosa becomes dry and atrophic, and the patients can gradually show dysgeusia, dysphagia and dysphonia, as well as risk of developing ulcerations<sup>4,6</sup>, caries, gingivitis, periodontitis, candidosis, and bacterial sialadenitis, among others.<sup>11,12</sup> Those changes cause important harm to the oral homeostasis and to the quality of life. Considering the abovementioned, the present study was designed to provide an update on the etiology, diagnostic methods and therapeutic strategies for the management of hyposalivation and xerostomia.

## DIAGNOSIS OF SALIVARY DYSFUNCTIONS

The diagnosis of salivary dysfunctions can be obtained by means of subjective and objective methods. These methods can be classified into questionnaires or interviews, secretion tests, mucosal surface tests, qualitative analyses, functional analyses and glandular morphology analyses<sup>13</sup> (Table 1).

**Table 1.** Diagnostic methods of salivary dysfunctions.

<b>Subjective analysis</b>	Questionnaires or interviews	Questionnaires on xerostomia Visual analogue scale OHIP-14*
	Secretion tests	Sialometry Schirmer oral test
<b>Objective analysis</b>	Mucosal surface tests	Biopsies Salivary Ferning test Mucus®
	Functional analyses	Scintigraphy Wafer test
	Sialochemistry	Salivary composition test Total protein test
	Glandular morphological analyses	Sialography Ultrasonography Magnetic resonance Computed tomography

\* Oral Health Impact Profile -14

\*\* Quality of Life Head and Neck 35

Source: Modified from Löfgren et al.<sup>13</sup>

Subjective methods are used to determine the intensity and cause of xerostomia.<sup>14,15</sup> A number of questionnaires have been utilized, and there is not a consensus on the best form of grading xerostomia, mainly due to the difficulty in obtaining a suitable response from the patient. Pai et al.<sup>16</sup> and Gerdin et al.<sup>17</sup> suggest the application of the visual analogue scale adapted to the evaluation of xerostomia.

Other authors suggest instruments that broaden the analysis of xerostomia, in grading aspects related to chewing, swallowing, speech, sleep and quality of life.<sup>18</sup>

The clinical method most often employed for the diagnosis of salivary dysfunction is sialometry, which consists in the whole saliva collection or the fluid produced by each major gland individually. Sialometry can be done by different methods, under stimulation or at rest.<sup>13</sup> Hyposalivation is considered when salivary flow rate (SFR) is < 0.1 mL/min at rest or < 0.7 mL/min under stimulation.<sup>11,13</sup> Otherwise, there are authors who consider SFR ≤ 0.3 mL/min, at rest, as abnormal.<sup>19</sup> Sialometry provides objective evidence of the reduction in SFR, but it does not help to diagnosis the dysfunction etiology.<sup>4</sup> The Schirmer test is usually employed in the diagnosis of xerophthalmia. Authors suggest its utilization also for quantifying non-stimulated SFR, since it consists in a simple method that provides an adequate diagnosis of glandular function.<sup>20,21</sup>

The assessment methods for the mucosal surface include biopsy of the minor salivary glands, the salivary ferning test and the *mucus*<sup>®</sup> test among others. The biopsy of the lower lip glands is used for the diagnosis of systemic disorders such as amyloidosis, sarcoidosis, Sjögren syndrome and neonatal hemochromatosis.<sup>22</sup> In the ferning test, hyposalivation is detected by electron microscopy, which evaluates the salivary crystals.<sup>23</sup> The *mucus*<sup>®</sup> test is a noninvasive method which analysis mucosal surface by a condenser that measures impedance with sensitive capacitors.<sup>24</sup>

Qualitative analysis of saliva includes assaying inorganic and organic components.<sup>25-27</sup> Diagnostic imaging techniques allow the identification of alterations in anatomic characteristics, as well as the evaluation of glandular function.<sup>28</sup> Salivary scintigraphy provides information about parenchyma and excretion of major salivary glands after endovenous administration of technetium pertechnetate.<sup>29,30</sup> It is a

noninvasive, easy to perform, reproducible technique and well tolerated by patients.<sup>31</sup>

Like scintigraphy, the wafer test is a semiquantitative functional analysis method, employed in the initial diagnosis of salivary gland disorders.<sup>19</sup>

Magnetic resonance (MR) provides excellent image contrast for soft tissues and spatial definition, with the advantage of not using ionizing radiation.<sup>32,33</sup> In sialography by MR, the saliva itself serves as contrast in obtaining the images.<sup>28</sup> Its disadvantages include poor availability, high cost and time-consuming. Computed tomography (CT) is more accessible when compared to MR. It is indicated for the diagnosis of calculi inside the gland or duct and evaluation of bone erosion caused by malignant lesions.<sup>32</sup> The addition of radiodrugs allows the differentiation between benign and malignant lesions. It is the examination of choice in the diagnosis of inflammatory lesions of the salivary glands, but the patient is subjected to ionizing radiation with contrast.

Alternatively, ultrasonography is an easy to perform method with low cost. It enables to differentiate between intra- and extraglandular lesions, as well as between cystic and solid lesions, besides to diagnose calculi and dilations of salivary ducts, and to guide biopsies or drainages.<sup>34</sup>

## **ETIOLOGY OF SALIVARY DYSFUNCTIONS**

### **DRUGS**

More than 500 drugs are able of inducing hyposalivation and xerostomia, but they rarely cause irreversible damage to the salivary glands.<sup>15,35</sup> These drugs cause salivary dysfunction through anticholinergic, sympathomimetic, antimuscarinic,

cytotoxic action or by perturbing the ion transport pathways in the acinar cells<sup>36,37</sup> (Table 2).

**Table 2.** Drugs that cause salivary dysfunctions.

Drug	Subclass	Commercial form
<b>Sympathomimetic drugs</b>		
Antidepressant	-Monoaminoxidase inhibitors -Serotonin uptake inhibitors	Venlafaxine Reboxetine Fluoxetine HCl Maprotiline HCl
Antihypertensive	- Angiotensin converting enzyme Inhibitors - Angiotensin II receptor antagonists - Adrenergic blockers -Central adrenergic stimulants -Beta blockers -Calcium channel blockers	Metoprolol Monoxidine Rilmenedine Captopril Losartan Guanethidine Methyldopa Esmolol Felodipine
Appetite suppressors		Sibutramine Fenfluramine Phentermine
Decongestants		Pseudoephedrine Cetirizine Loratadine
Bronchodilators		Tiotropium
Skeletal muscle relaxants		Tizanidine
Antimigraine agents		Rizatriptain
<b>Anticholinergic drugs</b>		
Tricyclic antidepressants		Amitriptyline Clomipramine Amoxapine Protriptyline Doxepin Imipramine Trimipramine
Diuretics		Furosemide Bumetanide Torsemide

**Table 2.** Drugs that cause salivary dysfunctions.

		Ethacrynic acid
Muscarinic receptor antagonists		
Antipsychotics	-Haloperidol -Phenothiazine derivatives -Antiparkinsonian drugs -Anticonvulsivants	Promazine Triflupromazine Mesoridazine Thioridazine Clozapine Olanzapine Azatadine Brompheniramine Chlorpheniramine Cyproheptadine Dexchlorpheniramine Hydroxyzine Phenindamine
Antihistamines		Azatadine Brompheniramine Chlorpheniramine Cyproheptadine Dexchlorpheniramine Hydroxyzine Phenindamine
<b>Cytotoxic drugs</b>	-Antineoplastics	Fluorouracil Interferon Radioactive iodine
<b>Drugs with unknown mechanism of action</b>	- Proton pump inhibitors -Antimicrobials	Omeprazole Metronidazol Amoxicillin
<b>Drugs with synergistic mechanism of action</b>	-Opioids -Hypnotics	Tramadol Diazepam Lorazepam Alprazolam Lexotan

SOURCE: Modified from Sreebny and Schwartz<sup>37</sup> and Tschoppe et al.<sup>38</sup>

## AGING

Several studies have investigated the effects of aging on the salivary glands, but there is still controversy as to the salivary dysfunctions in the elderly. While some authors have demonstrated impaired glandular function, others have not found salivary dysfunctions in the healthy and non-users of drugs elderly.<sup>39-44</sup> According to

Tylenda et al.<sup>43</sup>, aging leads to the loss of about 30% of acinar cells, with substitution of secretory components by fibrous and adipose tissue.

Ghezzi and Ship<sup>44</sup> found that after the use of an anticholinergic drug, glandular function is more affected in the elderly than in young individuals. Besides, there are changes in salivary levels of sodium, potassium, IgA, proline-rich protein, lactoferrin and lysozyme in elderly.<sup>1,2,45-48</sup> Yeh et al.<sup>49</sup> identified a reduction in the SFR of elderly, even those not using systemic drugs, suggesting a relation between salivary dysfunction and aging.

On the other hand, there are authors that defend the hypothesis that the decrease of SFR in elderly results exclusively from the action of drugs and systemic disorders, more common in this age group than in young individuals.<sup>50-51</sup> According to Locker<sup>50</sup> and Shetty et al.<sup>51</sup> xerostomia is proportional to the number of drugs that the elderly utilize.

## **SYSTEMIC DISORDERS**

Qualitative and quantitative salivary changes can be associated with particular systemic disorders that cause dysfunctions in neurotransmitter receptors, destruction of glandular parenchyma, interference with the secretion process or alterations in fluids and electrolytes.<sup>42</sup> Main systemic disorders related to glandular dysfunctions are listed in Table 3. Since many systemic conditions are associated with hyposalivation and xerostomia, it would not be possible to address them individually, and therefore, we chose to describe Sjögren syndrome, HIV infection and graft-versus-host disease.

**Table 3.** Systemic disorders associated with salivary dysfunctions.

<b>Rheumatological Chronic Inflammatory Disorders</b>	<ul style="list-style-type: none"> <li>- Sjögren Syndrome</li> <li>- Rheumatoid Arthritis</li> <li>- Juvenile Idiopathic Arthritis</li> <li>- Systemic Lupus Erythematosus</li> <li>- Systemic Sclerosis</li> <li>- Primary Biliary Cirrhosis</li> <li>- Mixed Connective Tissue Disease</li> <li>- Sarcoidosis</li> <li>- Amyloidosis</li> <li>- Crohn's Disease</li> <li>- Ulcerative Colitis</li> </ul>
<b>Endocrine Disorders</b>	<ul style="list-style-type: none"> <li>- Diabetes Mellitus</li> <li>- Hyperthyroidism/Hypothyroidism</li> <li>- Cushing Syndrome</li> <li>- Addison's Disease</li> <li>- Depression</li> <li>- Narcolepsy</li> <li>- Parkinson's Disease</li> <li>- Bell's Paralysis</li> <li>- Alzheimer's Disease</li> <li>- Holmes-Adie Syndrome</li> </ul>
<b>Neurologic Disorders</b>	<ul style="list-style-type: none"> <li>- Agenesis of Salivary Glands</li> <li>- Ectodermic Dysplasia</li> <li>- Cystic Fibrosis</li> <li>- Prader-Willi Syndrome</li> <li>- Auto-Immune Thyroiditis</li> <li>- Chronic Pancreatitis</li> <li>- Celiac Disease</li> <li>- Down Syndrome</li> <li>- Familial Amyloidotic Polyneuropathy</li> <li>- Myotonic Dystrophy</li> <li>- Gaucher Disease</li> <li>- Major Thalassemia</li> <li>- Papillon-Lefèvre Syndrome</li> <li>- Dehydration</li> <li>- Eating Disorders</li> <li>- End Stage Renal Disease</li> <li>- Nutritional Deficiencies</li> <li>- Anorexia Nervosa</li> <li>- Bulimia</li> <li>- Anemia</li> <li>- Atrophic Gastritis</li> <li>- Alcohol Abuse</li> </ul>
<b>Genetic, Congenital or Chronic Disorders</b>	<ul style="list-style-type: none"> <li>- HIV/AIDS</li> <li>- Epidemic Parotitis</li> <li>- Epstein-Barr Virus Infection</li> <li>- Bacterial Sialadenitis</li> <li>- Tuberculosis</li> <li>- Hemochromatosis</li> <li>- Wegener's Disease</li> <li>- Hypertension</li> <li>- Fibromyalgia</li> <li>- Chronic Fatigue Syndrome</li> </ul>
<b>Metabolic Disorders</b>	<ul style="list-style-type: none"> <li>- HIV/AIDS</li> <li>- Epidemic Parotitis</li> <li>- Epstein-Barr Virus Infection</li> <li>- Bacterial Sialadenitis</li> <li>- Tuberculosis</li> <li>- Hemochromatosis</li> <li>- Wegener's Disease</li> <li>- Hypertension</li> <li>- Fibromyalgia</li> <li>- Chronic Fatigue Syndrome</li> </ul>
<b>Infectious Disorders</b>	<ul style="list-style-type: none"> <li>- HIV/AIDS</li> <li>- Epidemic Parotitis</li> <li>- Epstein-Barr Virus Infection</li> <li>- Bacterial Sialadenitis</li> <li>- Tuberculosis</li> <li>- Hemochromatosis</li> <li>- Wegener's Disease</li> <li>- Hypertension</li> <li>- Fibromyalgia</li> <li>- Chronic Fatigue Syndrome</li> </ul>
<b>Others</b>	<ul style="list-style-type: none"> <li>- HIV/AIDS</li> <li>- Epidemic Parotitis</li> <li>- Epstein-Barr Virus Infection</li> <li>- Bacterial Sialadenitis</li> <li>- Tuberculosis</li> <li>- Hemochromatosis</li> <li>- Wegener's Disease</li> <li>- Hypertension</li> <li>- Fibromyalgia</li> <li>- Chronic Fatigue Syndrome</li> </ul>

Source: Porter et al.<sup>4</sup>; von Bultzingslowen et al.<sup>42</sup>

## Sjögren Syndrome

One of the disorders most associated with hyposalivation is Sjögren syndrome, an autoimmune disease of unknown etiology that affects the exocrine glands, mainly the salivary and lacrimal glands.<sup>20</sup> The disease may include impaired pulmonary, articular, renal and neurological function in its spectrum.<sup>52,53</sup> The syndrome is classified as primary when limited to the exocrine glands, and secondary if accompanied by other autoimmune diseases.<sup>54,55</sup> Its autoimmune profile is due to the circulating antibodies and glandular lymphocytic infiltrate, composed mainly of TCD4 lymphocytes.<sup>20,56,57</sup> Evidence indicates that the severity of secretory dysfunction is not necessarily related to the degree of glandular infiltration and destruction.<sup>22</sup>

The treatment of extraglandular manifestations of the disease includes drugs such as hydroxychloroquine, methotrexate and systemic corticosteroids, which appear to foster increased SFR.<sup>54,58</sup> The monoclonal antibodies rituximab and epratuzumab have been shown to be effective in reducing glandular inflammation, alleviating xerostomia and increasing SFR.<sup>59,60</sup>

## HIV Infection

Between 2 and 10% of patients infected by HIV present with xerostomia and up to 37% reduction in SFR.<sup>61,62</sup> The principal causes of dysfunction are disease of the salivary glands associated with HIV, Kaposi sarcoma, non-Hodgkin lymphoma, intraglandular lymphadenopathy and acute suppurative sialadenitis.<sup>4</sup> Glandular involvement is more common in children, who also show more favorable therapeutic

results than do adults. The outcome of anti-retroviral treatments has led to a decrease in the prevalence of these alterations.<sup>63</sup> Xerostomia in HIV-positive patients can also be the consequence of drugs such as didanosine, reverse transcriptase inhibitors or proteases.<sup>4</sup>

### **Graft-versus-host disease**

Graft-versus-host disease is an association of clinical alterations that appear after bone marrow transplantation.<sup>64,65</sup> The disease is mediated by autoreactive T lymphocytes which infiltrate several tissues and organs.<sup>66</sup> The disease shows different degrees of morbidity, with cutaneous, oral, ophthalmologic, pulmonary, articular and genitourinary manifestations. In the oral mucosa, there are lichenoid lesions, erythema, ulcerations, and areas of hyperkeratosis and atrophy.<sup>67</sup> Xerostomia is the result of fibrosis of the parotid glands and alterations in the chemical composition of saliva, with decreased levels of sodium and increased potassium. In the chronic course of the disease, the muscarinic receptors are harmed.<sup>68</sup>

### **RADIOTHERAPY AND CHEMOTHERAPY**

The major salivary glands are often involved in the radiation portals because they are in the proximity of the primary tumor sites and lymphatic chains of the head and neck region. As a consequence of radiotherapy, they undergo a process of degeneration, resulting in hyposalivation and xerostomia.<sup>69</sup> The severity of dysfunction is determined by factors such as dose, radiation portals and individual

response of the patient.<sup>70,71</sup> In malignant tumors located in the posterior portion of the mouth and in the oropharynx, radiotherapy includes parallel and opposing lateral fields on the upper region of the neck and side of the face. In these cases, both parotids are irradiated, leading to greater levels of xerostomia.<sup>72,73</sup>

Radiotherapy-induced hyposalivation ranges from a small degree of dry mouth to total lack of saliva and oral mucosa atrophy. The saliva undergoes qualitative alterations, there are decreased activity of amylases, buffering capacity and pH, with consequent acidification.<sup>74</sup> There are also increased levels of calcium, chloride, magnesium and proteins and reduction in bicarbonate.<sup>75</sup> These alterations indicate that the parotids function is more affected than that of the other glands.<sup>76</sup> Eisbruch et al.<sup>69</sup> suggested that on the radiotherapy the decrease in SFR can be related to development of mucositis, which occurs due to reduction in mucins, and epidermal and fibroblast growth factors.

Damage to the acinar cells becomes irreversible after cumulative doses of 26 to 39 Gy, causing the patients to have a SFR less than 10% of that prior to radiotherapy.<sup>77,78</sup> Xerostomia is perceived in the initial phases of radiotherapy, with reductions of 50 to 60% in SFR in the first week, and up to about 80% at the end of the seventh week.<sup>79,80</sup>

Despite being stable, because they do not have a high mitotic rate, acinar cells respond readily to radiation.<sup>81</sup> The mechanisms that lead to tissue destruction and greater sensitivity of salivary glands are still inconclusive.<sup>81,82</sup> The glandular alterations begin with damage to the plasma membrane, loss of response to autonomic control, edema, degeneration and necrosis of acinar cells. The acute effects begin 24 hours after the start of therapy and stabilize in 72 hours. The late effects are a consequence of fibrosis and acinar atrophy<sup>12,83</sup>, which occur because of

mesenchymal alterations, including changes in the extracellular matrix, especially laminin and collagen IV.<sup>84</sup>

Hakim et al.<sup>85</sup> evaluated, in the parotids of rabbits, functional changes and Ki67, smooth muscle actin (SMA) and tenascin-C immunodetection after irradiation with 15 Gy. There was a significant change in the absorption of <sup>99m</sup>Tc-pertechnetate, decreased Ki67 detection and marked redistributions of SMA and tenascin-C. According to the authors, the increased tenascin-C detection, caused by the damage to the basal membrane of acinar cells, and the reduction in SMA expression can be responsible for the functional glandular changes.

Avila et al.<sup>86</sup> demonstrated that ionizing radiation induced apoptosis in the acinar cells of the parotids of rodents. Apoptosis induced by radiation was dose-dependent and was correlated with salivary dysfunctions. In addition, the apoptotic response seen after irradiation was dependent on p53 expression. Cannon et al.<sup>87</sup> demonstrated radiotherapy toxicity to the parotid gland by means of fluorodeoxyglucose-labeled positron emission tomography-computed tomography (FDG-PET-CT).

The alterations in SFR and viscosity can persist for years and recovery depends on the characteristics of each patient.<sup>74</sup> Currently, with the use of intensity-modulated radiotherapy (IMRT), the tissues located in the proximity of the tumor are more preserved in comparison with conventional radiotherapy.<sup>88</sup> IMRT is a technique that allows radiation to be dosed and distributed in the tumor more precisely, thereby sparing surrounding tissues.<sup>89</sup>

Exclusive chemotherapy treatment causes transient xerostomia, with recovery of salivary levels as prior to treatment one year after the end of treatment cycles.<sup>90</sup> According to Jensen et al.<sup>90</sup>, chemotherapeutic drugs appear to affect the function of

acinar and ductal cells, by influencing cell division. Chemotherapeutic agents can induce dilation of the excretory duct, acinar degeneration and inflammation of glandular tissue.<sup>91</sup>

## **THERAPEUTIC OPTIONS**

The therapeutic approach of salivary dysfunctions depends basically on residual glandular function and is aimed at the alleviation of symptoms and prevention and correction of eventual sequelae, as well as at the treatment of associated systemic diseases. The treatment of hyposalivation and xerostomia can be classified as (1) preventive, (2) symptomatic, (3) topical and systemic stimulants, (4) disease-modifying agents, and (5) regenerative.<sup>42,92</sup>

### ***PREVENTIVE THERAPIES***

Cytoprotective drugs, used to minimizing the effects of radiotherapy, have been studied. Amifostine is an organic thiophosphate utilized in patients subjected to high doses of ionizing radiation, whose portal includes a large part of the parotids.<sup>93</sup> Its protective effect prevents the formation of free radicals and provides DNA repair, reducing the intensity and duration of xerostomia, without interfering with the control of the tumor and patients survival.<sup>94</sup> Despite these benefits, amifostine has adverse effects such as nausea, vomiting, hypotension, transient hypocalcemia and allergic reactions.<sup>95</sup> Another cytoprotector described in the literature is tempol, a stable nitroxide that still needs clinical studies to support its use in humans.<sup>77</sup> Cotrim et al.<sup>96</sup> demonstrated that tempol maintained normal SFR in irradiated animals.

The insulin-like growth factor and keratinocyte growth factor (KGF) in animals suppress apoptosis and favor the survival and proliferation of acinar cells after radiotherapy.<sup>97,98</sup> Zheng et al.<sup>99</sup> demonstrated that KGF prevented hyposalivation in mice, since the transgenic animals showed a higher number of acinar and endothelial cells.

Teymoortash et al.<sup>100</sup> suggested the utilization of botulinum toxin as an alternative preventive against salivary damage caused by ionizing radiation. The intraglandular application of the toxin, prior to radiotherapy, significantly prevented functional and histological changes in rats.

In irradiated patients, the transfer of the submandibular gland to the submental space is also indicated as a preventive method. The transfer allows the structure to receive a lower quantity of radiation, thereby maintaining its excretory function.<sup>101</sup>

Some preclinical studies of genetic transfer for glandular protection have been conducted. Baum et al.<sup>102</sup> and Delporte et al.<sup>103</sup> in studying human aquaporin-1 (hAQP1) in animals, observed that its action on the salivary glands causes an increase in aqueous secretion in response to an osmotic gradient. Currently, a phase I study was done in patients to evaluate its safety in humans.<sup>104</sup> Another study indicated positive results with use of manganese superoxide dismutase-plasmid/liposomes (MnSOD-PL) in the prevention of the noxious effects of ionizing radiation on the salivary glands.<sup>105</sup> Palaniyandi et al.<sup>106</sup> identified a splice variant of a cellular kinase, Tousled-like kinase 1B (TLK1B), which when overexpressed protected normal epithelial cells against ionizing radiation-induced cell death. The results demonstrated a reduction in acinar atrophy, glandular fibrosis and inflammatory infiltrate.

## **DISEASE-MODIFYING AGENTS**

Disease-modifying agents especially include immunomodulatory and immunosuppressive drugs, utilized in patients with Sjögren syndrome, with the objective of reestablishing the altered immunologic mechanisms.<sup>20</sup> Interferons are proteins that regulate cell proliferation and differentiation, cellular expression of surface antigens and induce enzymes.<sup>22</sup> The results with respect to the use of interferon alpha for xerostomia are controversial. Ferraccioli et al.<sup>107</sup> found that parenteral administration of interferon alpha-2 three times a week produced an increase in salivary and lachrymal secretions. Ship et al.<sup>108</sup> observed that this drug enhanced SFR under stimulation but did not alter flow at rest. On the other hand, Cummins et al.<sup>109</sup> found considerable elevation in SFR at rest and small alteration in stimulated SFR.

Another disease-modifying agent is rituximab, a monoclonal antibody that crossreacts with the antigen CD20, present in more than 90% of B cells. Its benefit with regard to xerostomia is related to reducing glandular lymphocytic infiltrate occurring in Sjögren syndrome.<sup>60</sup>

## **SYMPTOMATIC THERAPIES**

Drinking water frequently is an alternative more commonly used by patients with xerostomia, but saliva substitutes can provide higher viscosity and protection to the oral mucosa.<sup>77</sup> The ideal agent should provide long-lasting and intense hydration of the oral mucosa, requiring a minimal number of applications, without adverse

effects.<sup>6</sup> Saliva substitutes differ in chemical composition and viscosity.<sup>110</sup> They are mostly composed of carboxymethylcellulose (CMC), mucins, xanthan gum, hydroxyethylcellulose, linseed oil or polyethylene oxide.<sup>77,110</sup> When lubricants containing CMC are compared to those with mucins and xanthan gum, their rheologic and moisturizing properties are inferior.<sup>111</sup> Gelatinous substitutes of saliva, containing polyglycerylmethacrylate has also been suggested and are indicated for periods of decreased SFR.<sup>112</sup> According to Dost and Farah<sup>113</sup>, the length of salivary substitutes tends to be limited and there is a need for frequent reapplication.

Other symptomatic strategies include the slow release and continuous oral lubrication mechanisms.<sup>114</sup> Tsibouklis et al.<sup>115</sup> cite hydrogel films that allow the continuous release of substances for treatment of xerostomia. However, these can interfere with speech in patients.<sup>116</sup>

### ***TOPICAL AND SYSTEMIC STIMULANTS***

Pilocarpine is a parasympathomimetic, non-selective muscarinic agonist, which has been indicated for the treatment of xerostomia. Its recommended initial dose is 5 mg/day up to a maximum of 30 mg/day.<sup>117</sup> In patients subjected to radiotherapy of the head and neck, the maximal effect of pilocarpine is obtained between two and three months after the start of treatment.<sup>118</sup> Its adverse effects include hyperhidrosis, nausea, rhinitis, dizziness, intestinal colic and polakuria. Its use is contraindicated in heart patients, individuals with chronic obstructive pulmonary disease, asthma and glaucoma.<sup>119</sup> Epstein and Schubert<sup>120</sup> proposed the association of pilocarpine and anethole trithione (ANTT) to potentiate the effect of both on the salivary function. ANTT does not have a cholinergic action, but increases

the availability of muscarinic receptors in the post-synaptic membrane.<sup>121,122</sup> However, this association did not demonstrate positive results in the treatment of xerostomia in patients with primary Sjögren syndrome.<sup>123</sup>

Cevimeline is a selective muscarinic agonist for M1 and M3 receptors, found in the salivary and lacrimal glands. Since it has no effect on M2 receptors, it shows fewer adverse effects when compared to pilocarpine, and besides, it has a long-lasting action. The most common associated side effect is dyspepsia.<sup>124</sup> The recommended dose is 30 mg administered three times a day.<sup>125</sup>

Another systemic sialogogue is bethanecol, a carbamic ester of β-methylcholine resistant to cholinesterase, whose action is concentrated in the M3 receptors. Jham et al.<sup>126</sup>, in a randomized phase III trial, observed a significant increase in SFR at rest and a decrease in xerostomia in patients treated with head and neck radiotherapy. The dose indicated is 25 mg, three times a day. Its adverse effects, despite being infrequent, include nausea and diarrhea.

To stimulate salivary secretion, other drugs such as bromhexine<sup>127,128</sup> and nizatidine<sup>129,130</sup> are described in the literature. Bromhexine is a drug with mucolytic properties, used in respiratory infections, which also enhances salivary and lachrymal secretion. Nizatidine is an H2 receptor antagonist which inhibits acetylcholinesterase, allowing a greater availability of acetylcholine. Both have shown favorable results in SFR and have been used in patients with Sjögren syndrome.<sup>127-130</sup>

Sugar-free chewing gum and jellybeans can be utilized as topical salivary stimulants.<sup>92</sup> They usually contain xylitol (low calorie sugar), which inhibits the growth of cariogenic bacteria and reduces the incidence of caries.<sup>131</sup> Kleinegger<sup>132</sup> describes SalivaSure® as a topical stimulant that does not irritate soft tissues or cause tooth decay. Its composition includes citric acid and xylitol.

Domingo<sup>133</sup> utilized electrostimulation in patients with salivary dysfunction, and obtained an increased SFR from parotids. Meanwhile, the results with this technique are still inconclusive.<sup>134</sup> Acupuncture is also cited as an alternative for xerostomia. Subjective and objective aspects of salivary function are improved with those technique.<sup>135,136</sup>

In addition to this, hyperbaric oxygen in irradiated patients demonstrates increased salivary function<sup>137</sup> and reduction in the number of cariogenic bacteria and *Candida albicans*. Teguh et al.<sup>138</sup> pointed out that hyperbaric oxygen favors neoangiogenesis and recruitment of bone marrow stem cells.

### **REGENERATIVE THERAPIES**

Stem cell transplantation is emerging as an alternative for the reestablishment of glandular function, since these cells have the capacity to self-renew and differentiate into any type cell constituent of the salivary glands.<sup>139-141</sup> Bone marrow stem cells and adipose tissue-derived stromal cells have been tested.<sup>142-145</sup> Yamamura et al.<sup>146</sup> suggested the utilization of dental pulp cells as a form of treatment for hyposalivation after radiotherapy.

### **DISCUSSION**

Salivary dysfunctions are common, have a negative impact on the quality of life, and can be caused by a number of local and systemic conditions. Clinicians must be aware of the signs and symptoms of salivary disorders and be able to diagnose and treat them. In this review, we described subjective methods, as well as objective

methods for determining alterations in salivary secretion. In addition to this, we addressed the possible etiologic factors and established treatments in the literature, as well as new therapeutic strategies still under investigation.

The use of medications is the most frequent cause of xerostomia<sup>4,37,38</sup>, because a number of drugs are associated with salivary dysfunctions. Tricyclic antidepressants, sedatives, tranquilizers, antihistamines, antihypertensives and anticonvulsants are examples of often used drugs that are associated with salivary hypofunction.<sup>4,37</sup> Besides drugs, changes related to aging<sup>43-49</sup> and a number of systemic disorders<sup>4,42,56,61,68</sup>, such as Sjögren syndrome, are associated with decreased SFR.

Another important factor associated with salivary dysfunction is radiotherapy. Approximately 70% of patients who receive radiotherapy of the head and neck develop hyposialia, due to a progressive decrease in salivary gland function.<sup>69-72</sup> There is macroscopically detectable loss of glandular structure as a consequence of radiotherapy, besides microscopic alterations indicative of cell death, hypovascularization, formation of fibrous tissue and edema.<sup>81-86</sup> In these cases, preventive therapies such as submandibular gland transfer and the use of cytoprotective agents, growth factors and botulinum toxin have been tested for minimizing the effects of ionizing radiation on the salivary glands.<sup>93,96-101</sup>

The treatment of salivary dysfunctions is mainly devoted to alleviate symptoms and to stimulate residual glandular function. The symptomatic treatments consist mainly in salivary substitutes, which provide continuous lubrication of the oral mucosa<sup>6,77,11-113</sup>. Currently, devices for slow and prolonged release of these oral lubricants, mimicking glandular function, have been tested.<sup>114-116</sup> In relation to salivary stimulation methods, current interest is focused on the development of drugs

that increase SFR without adverse effects. Such drugs have not yet been found, although the use of pilocarpine, cevimeline and bethanechol has been established.

119,120,124,126

Clinicians, together with the patient, should select a therapeutic modality most suited for each case. The patient must be informed about the etiologic factors and adverse effects of hyposalivation, instructed about oral hygiene, drinking water frequently (to maintain the mucosa hydrated) and to avoid spicy and containing sugar foods, caffeine or alcohol.<sup>36</sup> New therapeutic modalities have been studied and involve stem cells transplantation<sup>140-146</sup>, with special attention to regeneration of damage caused by ionizing radiation to the salivary glands. More studies in this area are needed to provide new perspectives in the treatment of patients with salivary dysfunctions.

## REFERENCES

1. Schenkels LC, Veerman EC, Nieuw Amerongen AV. Biochemical composition of human saliva in relation to other mucosal fluids. *Crit Rev Oral Biol Med* 1995;6(2):161-175.
2. Amerongen AV, Veerman EC. Saliva--the defender of the oral cavity. *Oral Dis* 2002;8(1):12-22.
3. Ghezzi EM, Lange LA, Ship JA. Determination of variation of stimulated salivary flow rates. *J Dent Res* 2000;79(11):1874-1878.
4. Porter SR, Scully C, Hegarty AM. An update of the etiology and management of xerostomia. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2004;97(1):28-46.
5. Farsi NM. Signs of oral dryness in relation to salivary flow rate, pH, buffering capacity and dry mouth complaints. *BMC Oral Health* 2007; 7:15.

6. Thelin WR, Brennan MT, Lockhart PB, Singh ML, Fox PC, Papas AS, et al. The oral mucosa as a therapeutic target for xerostomia. *Oral Dis* 2008;14(8):683-689.
7. Dodds MW, Johnson DA, Yeh CK. Health benefits of saliva: a review. *J Dent* 2005;33(3):223-233.
8. Mese H, Matsuo R. Salivary secretion, taste and hyposalivation. *J Oral Rehabil* 2007;34(10):711-723.
9. Dijkema T, Raaijmakers CP, Braam PM, Roesink JM, Monninkhof EM, Terhaard CH. Xerostomia: a day and night difference. *Radiother Oncol* 2012;104(2):219-223.
10. Nederfors T. Xerostomia and hyposalivation. *Adv Dent Res* 2000;14:48-56.
11. Scully C, Felix DH. Oral medicine -- update for the dental practitioner: dry mouth and disorders of salivation. *Br Dent J* 2005;199(7):423-427.
12. Porter SR. Xerostomia: prevalence, assessment, differential diagnosis and implications for quality of life. *Oral Dis* 2010;16(6):501–502.
13. Löfgren CD, Wickström C, Sonesson M, Lagunas PT, Christersson C. A systematic review of methods to diagnose oral dryness and salivary gland function. *BMC Oral Health* 2012;12:29.
14. Eisbruch A, Rhodus N, Rosenthal D, Murphy B, Rasch C, Sonis S. The prevention and treatment of radiotherapy-induced xerostomia. *Semin Radiat Oncol* 2003;13(3):302-308.
15. Osailan SM, Pramanik R, Shirlaw P, Proctor GB, Challacombe SJ. Clinical assessment of oral dryness: development of a scoring system related to salivary flow and mucosal wetness. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2012;114(5):597-603.

16. Pai S, Ghezzi EM, Ship JA. Development of a Visual Analogue Scale questionnaire for subjective assessment of salivary dysfunction. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2001;91(3):311-316.
17. Gerdin EW, Einarson S, Jonsson M, Aronsson K, Johansson I. Impact of dry mouth conditions on oral health-related quality of life in older people. *Gerodontology* 2005;22(4):219-226.
18. Dirix P, Nuyts S, Vander Poorten V, Delaere P, Van den Bogaert W. The influence of xerostomia after radiotherapy on quality of life: results of a questionnaire in head and neck cancer. *Support Care Cancer* 2008;16(2):171-179.
19. Sánchez-Guerrero J, Aguirre-García E, Pérez-Dosal MR, Kraus A, Cardiel MH, Soto-Rojas AE. The wafer test: a semi-quantitative test to screen for xerostomia. *Rheumatology* 2002;41(4):381-389.
20. Tincani A, Andreoli L, Cavazzana I, Doria A, Favero M, Fenini MG, et al. Novel aspects of Sjögren's syndrome in 2012. *BMC Med* 2013;11:93.
21. López-Jornet P, Camacho-Alonso F, Bermejo-Fenoll A. A simple test for salivary gland hypofunction using Oral Schirmer's test. *J Oral Pathol Med* 2006;35(4):244-248.
22. Varela-Centelles P, Sanchez-Sanchez M, Seoane J. Lip biopsy for the diagnosis of Sjögren's syndrome: beware of the punch. *Int J Oral Maxillofac Surg* 2014;43(1):127-130.
23. Maragou M, Vaikousis E, Ntre A, Koronis N, Georgiou P, Hatzidimitriou E, et al. Tear and saliva ferning tests in Sjögren's syndrome (SS). *Clin Rheumatol* 1996;15(2):125-132.

24. Fujimaki Y, Tsunoda K, Ishimoto SI, Okada K, Kinoshita M, Igaki H, et al. Non-invasive objective evaluation of radiotherapy-induced dry mouth. *J Oral Pathol Med* 2014;43(2):97-102.
25. Thorn JJ, Prause JU, Oxholm P. Sialochemistry in Sjögren's syndrome: a review. *J Oral Pathol Med* 1989;18(8):457-468.
26. Kalk WW, Vissink A, Spijkervet FK, Bootsma H, Kallenberg CG, Nieuw Amerongen AV. Sialometry and sialochemistry: diagnostic tools for Sjögren's syndrome. *Ann Rheum Dis* 2001;60(12):1110-1116.
27. Almståhl A, Wikström M, Groenink J. Lactoferrin, amylase and mucin MUC5B and their relation to the oral microflora in hyposalivation of different origins. *Oral Microbiol Immunol* 2001;16(6):345-352.
28. Murdoch-Kinch CA. Salivary gland imaging. *J Calif Dent Assoc* 2011;39(9):649-654.
29. Vinagre F, Santos MJ, Prata A, da Silva JC, Santos AI. Assessment of salivary gland function in Sjögren's syndrome: the role of salivary gland scintigraphy. *Autoimmun Rev* 2009;8(8):672-676.
30. Zou Q, Jiao J, Zou MH, Xu JH, Pan YF, Chen JN, et al. Semi-quantitative evaluation of salivary gland function in Sjögren's syndrome using salivary gland scintigraphy. *Clin Rheumatol* 2012;31(12):1699-1705.
31. Klutmann S, Bohuslavizki KH, Kröger S, Bleckmann C, Brenner W, Mester J, et al. Quantitative salivary gland scintigraphy. *J Nucl Med Technol* 1999;27(1):20-26.
32. Burke CJ, Thomas RH, Howlett D. Imaging the major salivary glands. *Br J Oral Maxillofac Surg* 2011;49(4):261-269.

33. Kato H, Kanematsu M, Toida M, Kawaguchi T, Shibata T, Kajita K, et al. Salivary gland function evaluated by diffusion-weighted MR imaging with gustatory stimulation: preliminary results. *J Magn Reson Imaging* 2011;34(4):904-909.
34. Orlandi MA, Pistorio V, Guerra PA. Ultrasound in sialadenitis. *J Ultrasound* 2013;16(1):3-9.
35. Turner MD, Ship JA. Dry mouth and its effects on the oral health of elderly people. *J Am Dent Assoc* 2007;138:15S-20S.
36. Napeñas JJ, Brennan MT, Fox PC. Diagnosis and treatment of xerostomia (dry mouth). *Odontology* 2009;97(2):76-83.
37. Sreebny LM, Schwartz SS. A reference guide to drugs and dry mouth--2nd edition. *Gerodontics* 1997;14(1):33-47.
38. Tschoppe P, Wolgin M, Pischon N, Kielbassa AM. Etiologic factors of hyposalivation and consequences for oral health. *Quintessence Int* 2010;41(4):321-333.
39. Pedersen W, Schubert M, Izutsu K, Mersai T, Truelove E. Age-dependent decreases in human submandibular gland flow rates as measured under resting and post-stimulation conditions. *J Dent Res* 1985;64(5):822-825.
40. Navazesh M, Christensen C, Brightman V. Clinical criteria for the diagnosis of salivary gland hypofunction. *J Dent Res* 1992;71(7):1363-1369.
41. Percival RS, Challacombe SJ, Marsh PD. Flow rates of resting whole and stimulated parotid saliva in relation to age and gender. *J Dent Res* 1994;73(8):1416-1420.
42. von Bültzingslöwen I, Sollecito TP, Fox PC, Daniels T, Jonsson R, Lockhart PB, et al. Salivary dysfunction associated with systemic diseases: systematic review and

- clinical management recommendations. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2007;103(S57):1-15.
43. Tylenda CA, Ship JA, Fox PC, Baum BJ. Evaluation of submandibular salivary flow rate in different age groups. *J Dent Res* 1988;67(9):1225-1228.
44. Ghezzi EM, Ship JA. Aging and secretory reserve capacity of major salivary glands. *J Dent Res* 2003;82(10):844-848.
45. Baum BJ, Kousvelari EE, Oppenheim FG. Exocrine protein secretion from human parotid glands during aging: stable release of the acidic proline-rich proteins. *J Gerontol* 1982;37(4):392-395.
46. Baum BJ, Costa PT Jr, Izutsu KT. Sodium handling by aging human parotid glands is inconsistent with a two-stage secretion model. *Am J Physiol* 1984;246(1 Pt 2):R35-39.
47. Finkelstein MS, Tanner M, Freedman ML. Salivary and serum IgA levels in a geriatric outpatient population. *J Clin Immunol* 1984;4(2):85-91.
48. Zhang A, Sun H, Wang P, Wang X. Salivary proteomics in biomedical research. *Clin Chim Acta* 2013;415:261-265.
49. Yeh CK, Johnson DA, Dodds MW. Impact of aging on human salivary gland function: a community-based study. *Aging* 1998;10(5):421-428.
50. Locker D. Subjective reports of oral dryness in an older adult population. *Community Dent Oral Epidemiol* 1993;21(3):165-168.
51. Shetty SR, Bhowmick S, Castelino R, Babu S. Drug induced xerostomia in elderly individuals: An institutional study. *Contemp Clin Dent* 2012;3(2):173-175.
52. al-Hashimi I. The management of Sjögren's syndrome in dental practice. *J Am Dent Assoc* 2001;132(10):1409-1417.

53. Stojan G, Baer AN, Danoff SK. Pulmonary manifestations of Sjögren's syndrome. Curr Allergy Asthma Rep 2013;13(4):354-360.
54. Nazmul-Hossain AN, Morarasu GM, Schmidt SK, Walker AJ, Myers SL, Rhodus NL. A current perspective on Sjögren's syndrome. J Calif Dent Assoc 2011;39(9):631-637.
55. Vitali C, Bootsma H, Bowman SJ, Dorner T, Gottenberg JE, Mariette X, et al. Classification criteria for Sjögren's syndrome: we actually need to definitively resolve the long debate on the issue. Ann Rheum Dis 2013;72(4):476-478.
56. Berglová I, Krejsek J, Kolácková M, Slezák R. B cell toll-like receptors with respect to the pathogenesis of Sjögren's syndrome. Acta Medica 2011;54(2):51-57.
57. González S, Sung H, Sepúlveda D, González M, Molina C. Oral manifestations and their treatment in Sjögren's syndrome. Oral Dis 2013 Mar 18 doi: 10.1111/odi.12105.
58. Miyawaki S, Nishiyama S, Matoba K. Efficacy of low-dose prednisolone maintenance for saliva production and serological abnormalities in patients with primary Sjögren's syndrome. Intern Med 1999;38(12):938-943.
59. Steinfeld SD, Tant L, Burmester GR, Teoh NK, Wegener WA, Goldenberg DM, et al. Epratuzumab (humanised anti-CD22 antibody) in primary Sjögren's syndrome: an open-label phase I/II study. Arthritis Res Ther 2006;8(4):R129.
60. Pijpe J, Meijer JM, Bootsma H, van der Wal JE, Spijkervet FK, Kallenberg CG, et al. Clinical and histologic evidence of salivary gland restoration supports the efficacy of rituximab treatment in Sjögren's syndrome. Arthritis Rheum 2009;60(11):3251-3256.
61. Schiødt M. Less common oral lesions associated with HIV infection: prevalence and classification. Oral Dis 1997;3 (Suppl 1):S208-213.

62. Jankittivong A, Lin AL, Johnson DA, Langlais RP, Yeh CK. Salivary secretion, mucin concentrations and Candida carriage in HIV-infected patients. *Oral Dis* 2009;15(3):229-234.
63. Cavasin Filho JC, Giovani EM. Xerostomy, dental caries and periodontal disease in HIV+ patients. *Braz J Infect Dis* 2009;13(1):13-17.
64. Nagler RM, Nagler A. Salivary gland involvement in graft-versus-host disease: the underlying mechanism and implicated treatment. *Isr Med Assoc J* 2004;6(3):167-172.
65. Blazar BR, Murphy WJ, Abedi M. Advances in graft-versus-host disease biology and therapy. *Nat Rev Immunol* 2012;12(6):443-458.
66. Nagler RM, Sherman Y, Nagler A. Histopathological study of the human submandibular gland in graft versus host disease. *J Clin Pathol* 1999;52(5):395-397.
67. Mays JW, Sarmadi M, Moutsopoulos NM. Oral manifestations of systemic autoimmune and inflammatory diseases: diagnosis and clinical management. *J Evid Based Dent Pract* 2012;12(3 Suppl):265-282.
68. Imanguli MM, Atkinson JC, Mitchell SA, Avila DN, Bishop RJ, Cowen EW, et al. Salivary gland involvement in chronic graft-versus-host disease: prevalence, clinical significance, and recommendations for evaluation. *Biol Blood Marrow Transplant* 2010;16(10):1362-1369.
69. Eisbruch A, Kim HM, Terrell JE, Marsh LH, Dawson LA, Ship JA. Xerostomia and its predictors following parotid-sparing irradiation of head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 2001;50(3):695-704.
70. Lin SC, Jen YM, Chang YC, Lin CC. Assessment of xerostomia and its impact on quality of life in head and neck cancer patients undergoing radiotherapy, and

- validation of the Taiwanese version of the xerostomia questionnaire. *J Pain Symptom Manage* 2008;36(2):141-148.
71. Dirix P, De Keyzer F, Vandecaveye V, Stroobants S, Hermans R, Nuyts S. Diffusion-weighted magnetic resonance imaging to evaluate major salivary gland function before and after radiotherapy. *Int J Radiat Oncol Biol Phys* 2008;71(5):1365-1371.
72. Mira JG, Wescott WB, Starcke EN, Shannon IL. Some factors influencing salivary function when treating with radiotherapy. *Int J Radiat Oncol Biol Phys* 1981;7(4):535-541.
73. Joyston-Bechal S. Management of oral complications following radiotherapy. *Dent Update* 1992;19(6):232-234, 236-238.
74. de Barros Pontes C, Polizello AC, Spadaro AC. Clinical and biochemical evaluation of the saliva of patients with xerostomia induced by radiotherapy. *Braz Oral Res* 2004;18(1):69-74.
75. Dreizen S, Brown LR, Handler S, Levy BM. Radiation-induced xerostomia in cancer patients. Effect on salivary and serum electrolytes. *Cancer* 1976;38(1):273-278.
76. Almståhl A, Wikström M. Electrolytes in stimulated whole saliva in individuals with hyposalivation of different origins. *Arch Oral Biol* 2003;48(5):337-344.
77. Vissink A, Mitchell JB, Baum BJ, Limesand KH, Jensen SB, Fox PC, et al. Clinical management of salivary gland hypofunction and xerostomia in head-and-neck cancer patients: successes and barriers. *Int J Radiat Oncol Biol Phys* 2010;78(4):983-991.
78. Porter SR, Fedele S, Habbab KM. Xerostomia in head and neck malignancy. *Oral Oncol* 2010;46(6):460-463.

79. Burlage FR, Coppes RP, Meertens H, Stokman MA, Vissink A. Parotid and submandibular/sublingual salivary flow during high dose radiotherapy. *Radiother Oncol* 2001;61(3):271-274.
80. Dirix P, Nuyts S, Van den Bogaert W. Radiation-induced xerostomia in patients with head and neck cancer: a literature review. *Cancer* 2006;107(11):2525-2534.
81. Konings AW, Coppes RP, Vissink A. On the mechanism of salivary gland radiosensitivity. *Int J Radiat Oncol Biol Phys* 2005;62(4):1187-1194.
82. Grundmann O, Mitchell GC, Limesand KH. Sensitivity of salivary glands to radiation: from animal models to therapies. *J Dent Res* 2009;88(10):894-903.
83. Stephens LC, Ang KK, Schultheiss TE, King GK, Brock WA, Peters LJ. Target cell and mode of radiation injury in rhesus salivary glands. *Radiother Oncol* 1986;7(2):165-174.
84. Gustafsson H, Aalto Y, Franzén L, Thornell LE, Henriksson R. Effects of fractionated irradiation on the cytoskeleton and basal lamina in parotid glands--an immunohistochemical study. *Acta Oncol* 1998;37(1):33-40.
85. Hakim SG, Jacobsen HCh, Hermes D, Kosmehl H, Lauer I, Nadrowitz R, et al. Early immunohistochemical and functional markers indicating radiation damage of the parotid gland. *Clin Oral Investig* 2004;8(1):30-35.
86. Avila JL, Grundmann O, Burd R, Limesand KH. Radiation-induced salivary gland dysfunction results from p53-dependent apoptosis. *Int J Radiat Oncol Biol Phys* 2009;73(2):523-539.
87. Cannon B, Schwartz DL, Dong L. Metabolic imaging biomarkers of postradiotherapy xerostomia. *Int J Radiat Oncol Biol Phys* 2012;83(5):1609-1616.
88. Dirix P, Nuyts S. Parotid gland function after radiotherapy. *Lancet Oncol* 2010;11(6):515-516.

89. Murdoch-Kinch CA, Kim HM, Vineberg KA, Ship JA, Eisbruch A. Dose-effect relationships for the submandibular salivary glands and implications for their sparing by intensity modulated radiotherapy. *Int J Radiat Oncol Biol Phys* 2008;72(2):373-382.
90. Jensen SB, Mouridsen HT, Reibel J, Brünner N, Nauntofte B. Adjuvant chemotherapy in breast cancer patients induces temporary salivary gland hypofunction. *Oral Oncol* 2008;44(2):162-173.
91. Lockhart PB, Sonis ST. Alterations in the oral mucosa caused by chemotherapeutic agents. A histologic study. *J Dermatol Surg Oncol* 1981;7(12):1019-1025.
92. Atkinson JC, Grisius M, Massey W. Salivary hypofunction and xerostomia: diagnosis and treatment. *Dent Clin North Am* 2005;49(2):309-326.
93. De Souza CA, Santini G, Marino G, Nati S, Congiu AM, Vigorito AC, et al. Amifostine (WR-2721), a cytoprotective agent during high-dose cyclophosphamide treatment of non-Hodgkin's lymphomas: a phase II study. *Braz J Med Biol Res* 2000;33(7):791-798.
94. Sasse AD, Clark LG, Sasse EC, Clark OA. Amifostine reduces side effects and improves complete response rate during radiotherapy: results of a meta-analysis. *Int J Radiat Oncol Biol Phys* 2006;64(3):784-791.
95. Wasserman TH, Brizel DM, Henke M, Monnier A, Eschwege F, Sauer R, et al. Influence of intravenous amifostine on xerostomia, tumor control, and survival after radiotherapy for head-and-neck cancer: 2-year follow-up of a prospective, randomized, phase III trial. *Int J Radiat Oncol Biol Phys* 2005;63(4):985-990.
96. Cotrim AP, Hyodo F, Matsumoto K, Sowers AL, Cook JA, Baum BJ, et al. Differential radiation protection of salivary glands versus tumor by Tempol with

- accompanying tissue assessment of Tempol by magnetic resonance imaging. Clin Cancer Res 2007;13(16):4928-4933.
97. Lombaert IM, Brunsting JF, Wierenga PK, Kampinga HH, de Haan G, Coppes RP. Keratinocyte growth factor prevents radiation damage to salivary glands by expansion of the stem/progenitor pool. Stem Cells 2008;26(10):2595-2601.
98. Limesand KH, Said S, Anderson SM. Suppression of radiation-induced salivary gland dysfunction by IGF-1. PLoS One 2009;4(3):e4663.
99. Zheng C, Cotrim AP, Rowzee A, Swaim W, Sowers A, Mitchell JB, et al. Prevention of radiation-induced salivary hypofunction following hKGF gene delivery to murine submandibular glands. Clin Cancer Res 2011;17(9):2842-2851.
100. Teymoortash A, Müller F, Juricko J, Bieker M, Mandic R, Librizzi D, et al. Botulinum toxin prevents radiotherapy-induced salivary gland damage. Oral Oncol 2009;45(8):737-739.
101. Liu XK, Su Y, Jha N, Hong MH, Mai HQ, Fan W, et al. Submandibular salivary gland transfer for the prevention of radiation-induced xerostomia in patients with nasopharyngeal carcinoma: 5-Year outcomes. Head Neck 2011;33(3):389-395.
102. Baum BJ, Zheng C, Cotrim AP, McCullagh L, Goldsmith CM, Brahim JS, et al. Aquaporin-1 gene transfer to correct radiation-induced salivary hypofunction. Handb Exp Pharmacol 2009;(190):403-418.
103. Delporte C, O'Connell BC, He X, Lancaster HE, O'Connell AC, Agre P, et al. Increased fluid secretion after adenoviral-mediated transfer of the aquaporin-1 cDNA to irradiated rat salivary glands. Proc Natl Acad Sci U S A 1997;94(7):3268-3273.

104. Alevizos LG. National Institute of Dental and Craniofacial Research (NIDCR); 2006 – [cited 2013 Nov 8]. Available from: <http://www.clinicaltrials.gov/ct/show/NCT00372320>.
105. Epperly MW, Carpenter M, Agarwal A, Mitra P, Nie S, Greenberger JS. Intraoral manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) radioprotective gene therapy decreases ionizing irradiation-induced murine mucosal cell cycling and apoptosis. *In Vivo* 2004;18(4):401-410.
106. Palaniyandi S, Odaka Y, Green W, Abreo F, Caldito G, De Benedetti A, et al. Adenoviral delivery of Tousled kinase for the protection of salivary glands against ionizing radiation damage. *Gene Ther* 2011;18(3):275-282.
107. Ferraccioli GF, Salaffi F, De Vita S, Casatta L, Avellini C, Carotti M, et al. Interferon alpha-2 (IFN alpha 2) increases lacrimal and salivary function in Sjögren's syndrome patients. Preliminary results of an open pilot trial versus OH-chloroquine. *Clin Exp Rheumatol* 1996;14(4):367-371.
108. Ship JA, Fox PC, Michalek JE, Cummins MJ, Richards AB. Treatment of primary Sjögren's syndrome with low-dose natural human interferon-alpha administered by the oral mucosal route: a phase II clinical trial. IFN Protocol Study Group. *J Interferon Cytokine Res* 1999;19(8):943-951.
109. Cummins MJ, Papas A, Kammer GM, Fox PC. Treatment of primary Sjögren's syndrome with low-dose human interferon alfa administered by the oromucosal route: combined phase III results. *Arthritis Rheum* 2003;49(4):585-593.
110. Hahnel S, Behr M, Handel G, Bürgers R. Saliva substitutes for the treatment of radiation-induced xerostomia--a review. *Support Care Cancer* 2009;17(11):1331-1143.

111. Vissink A, s-Gravenmade EJ, Panders AK, Vermey A, Petersen JK, Visch LL, et al. A clinical comparison between commercially available mucin- and CMC-containing saliva substitutes. *Int J Oral Surg* 1983;12(4):232-238.
112. Regelink G, Vissink A, Reintsema H, Nauta JM. Efficacy of a synthetic polymer saliva substitute in reducing oral complaints of patients suffering from irradiation-induced xerostomia. *Quintessence Int* 1998;29(6):383-388.
113. Dost F, Farah CS. Stimulating the discussion on saliva substitutes: a clinical perspective. *Aust Dent J* 2013;58(1):11-17.
114. Kam AY, McMillan AS, Pow EH, Leung KC, Luk HW. A preliminary report on patient acceptance of a novel intra-oral lubricating device for the management of radiotherapy-related xerostomia. *Clin Oral Investig* 2005;9(3):148-153.
115. Tsibouklis J, Middleton AM, Patel N, Pratten J. Toward mucoadhesive hydrogel formulations for the management of xerostomia: the physicochemical, biological, and pharmacological considerations. *J Biomed Mater Res A* 2013;101(11):3327-3338.
116. McMillan AS, Tsang CS, Wong MC, Kam AY. Efficacy of a novel lubricating system in the management of radiotherapy-related xerostomia. *Oral Oncol* 2006;42(8):842-848.
117. Radvansky LJ, Pace MB, Siddiqui A. Prevention and management of radiation-induced dermatitis, mucositis, and xerostomia. *Am J Health Syst Pharm* 2013;70(12):1025-1032.
118. Nieuw Amerongen AV, Veerman EC. Current therapies for xerostomia and salivary gland hypofunction associated with cancer therapies. *Support Care Cancer* 2003;11(4):226-231.

119. Johnson JT, Ferretti GA, Nethery WJ, Valdez IH, Fox PC, Ng D, et al. Oral pilocarpine for post-irradiation xerostomia in patients with head and neck cancer. *N Engl J Med* 1993;329(6):390-395.
120. Epstein JB, Schubert MM. Synergistic effect of sialagogues in management of xerostomia after radiation therapy. *Oral Surg Oral Med Oral Pathol* 1987;64(2):179-182.
121. Ukai Y, Taniguchi N, Takeshita K, Kimura K, Enomoto H. Chronic anethole trithione treatment enhances the salivary secretion and increases the muscarinic acetylcholine receptors in the rat submaxillary gland. *Arch Int Pharmacodyn Ther* 1984;271(2):206-212.
122. Hamada T, Nakane T, Kimura T, Arisawa K, Yoneda K, Yamamoto T, et al. Treatment of xerostomia with the bile secretion-stimulating drug anethole trithione: a clinical trial. *Am J Med Sci* 1999;318(3):146-151.
123. Schiødt M, Oxholm P, Jacobsen A. Treatment of xerostomia in patients with primary Sjögren's syndrome with sulfarlem. *Scand J Rheumatol Suppl* 1986;61:250-252.
124. Chambers MS, Posner M, Jones CU, Biel MA, Hodge KM, Vitti R, et al. Cevimeline for the treatment of postirradiation xerostomia in patients with head and neck cancer. *Int J Radiat Oncol Biol Phys* 2007;68(4):1102-1109.
125. Chambers MS. Sjögren's syndrome. *ORL Head Neck Nurs* 2004;22(4):22-30.
126. Jham BC, Teixeira IV, Aboud CG, Carvalho AL, Coelho M, Freire AR. A randomized phase III prospective trial of bethanechol to prevent radiotherapy-induced salivary gland damage in patients with head and neck cancer. *Oral Oncol* 2007;43(2):137-142.

127. Avisar R, Savir H, Machtey I, Ovknin L, Shaked P, Menache R, et al. Clinical trial of bromhexine in Sjögren's syndrome. *Ann Ophthalmol* 1981;13(8):971-973.
128. Nanni JM, Nguyen KH, Alford CE, Robinson CP, Stewart CM, Maeda N, et al. Assessment of bromhexine as a treatment regimen in Sjögren's syndrome-like disease in the NOD (non-obese diabetic) mouse. *Clin Exp Rheumatol* 1997;15(5):515-521.
129. Adachi K, Ono M, Kawamura A, Yuki M, Fujishiro H, Kinoshita Y. Nizatidine and cisapride enhance salivary secretion in humans. *Aliment Pharmacol Ther* 2002;16(2):297-301.
130. Kasama T, Shiozawa F, Isozaki T, Matsunawa M, Wakabayashi K, Odai T, et al. Effect of the H<sub>2</sub> receptor antagonist nizatidine on xerostomia in patients with primary Sjögren's syndrome. *Mod Rheumatol* 2008;18(5):455-459.
131. Van Loveren C. Sugar alcohols: what is the evidence for caries-preventive and caries-therapeutic effects? *Caries Res* 2004;38(3):286-293.
132. Kleinegger CL. Dental management of xerostomia--opportunity, expertise, obligation. *J Calif Dent Assoc* 2007;35(6):417-424.
133. Domingo DL. The effects of electrostimulation on saliva production in postradiation head and neck cancer patients. *Oral Surg Oral Med Oral Pathol* 2004;97(4):464-465.
134. Fedele S, Wolff A, Strietzel F, López RM, Porter SR, Konttinen YT. Neuroelectrostimulation in treatment of hyposalivation and xerostomia in Sjögren's syndrome: a salivary pacemaker. *J Rheumatol* 2008;35(8):1489-1494.
135. Braga FP, Sugaya NN, Hirota SK, Weinfeld I, Magalhães MH, Migliari DA. The effect of acupuncture on salivary flow rates in patients with radiation-induced xerostomia. *Minerva Stomatol* 2008;57(7-8):343-348.

136. Garcia MK, Chiang JS, Cohen L, Liu M, Palmer JL, Rosenthal DI, et al. Acupuncture for radiation-induced xerostomia in patients with cancer: a pilot study. Head Neck 2009;31(10):1360-1368.
137. Cankar K, Finderle Z, Jan J. The effect of hyperbaric oxygenation on postradiation xerostomia and saliva in patients with head and neck tumours. Caries Res 2011;45(2):136-141.
138. Teguh DN, Levendag PC, Noever I, Voet P, van der Est H, van Rooij P, et al. Early hyperbaric oxygen therapy for reducing radiotherapy side effects: early results of a randomized trial in oropharyngeal and nasopharyngeal cancer. Int J Radiat Oncol Biol Phys 2009;75(3):711-716.
139. Lombaert IM, Brunsting JF, Wierenga PK, Kampinga HH, de Haan G, Coppes RP. Cytokine treatment improves parenchymal and vascular damage of salivary glands after irradiation. Clin Cancer Res 2008;14(23):7741-7750.
140. Feng J, van der Zwaag M, Stokman MA, van Os R, Coppes RP. Isolation and characterization of human salivary gland cells for stem cell transplantation to reduce radiation-induced hyposalivation. Radiother Oncol 2009;92(3):466-471.
141. Pringle S, Van Os R, Coppes RP. Concise review: Adult salivary gland stem cells and a potential therapy for xerostomia. Stem Cells 2013;31(4):613-619.
142. Lin CY, Chang FH, Chen CY, Huang CY, Hu FC, Huang WK, et al. Cell therapy for salivary gland regeneration. J Dent Res 2011;90(3):341-346.
143. Sumita Y, Liu Y, Khalili S, Maria OM, Xia D, Key S, et al. Bone marrow-derived cells rescue salivary gland function in mice with head and neck irradiation. Int J Biochem Cell Biol 2011;43(1):80-87.

144. Kojima T, Kanemaru S, Hirano S, Tateya I, Ohno S, Nakamura T, et al. Regeneration of radiation damaged salivary glands with adipose-derived stromal cells. *Laryngoscope* 2011;121(9):1864-1869.
145. Lim JY, Yi T, Choi JS, Jang YH, Lee S, Kim HJ, et al. Intraglandular transplantation of bone marrow-derived clonal mesenchymal stem cells for amelioration of post-irradiation salivary gland damage. *Oral Oncol* 2013;49(2):136-143.
146. Yamamura Y, Yamada H, Sakurai T, Ide F, Inoue H, Muramatsu T, et al. Treatment of salivary gland hypofunction by transplantation with dental pulp cells. *Arch Oral Biol* 2013;58(8):935-942.

---

**4 ARTIGO DE PESQUISA**

**4 ARTIGO DE PESQUISA****EFFECT OF LOW-LEVEL LASER THERAPY ON RADIOTHERAPY-INDUCED HYPOSALIVATION AND XEROSTOMIA: A PILOT STUDY**

Artigo submetido para avaliação (Anexo F)

Periódico: Photomedicine and Laser Surgery

Qualis Capes Odontologia 2013: B1

Fator de Impacto: 1,634

**EFFECT OF LOW-LEVEL LASER THERAPY ON RADIOTHERAPY-  
INDUCED HYPOSALIVATION AND XEROSTOMIA: A PILOT STUDY**

***LLLT on radiotherapy-induced salivary dysfunction***

SALEH, Jamil\*

FIGUEIREDO, Maria Antonia Zancanaro\*

CHERUBINI, Karen\*

BRAGA-FILHO, Aroldo\*\*

SALUM, Fernanda Gonçalves\*

**\*Oral Medicine Division, Pontifical Catholic University of Rio Grande do Sul-  
PUCRS, Brazil.**

**\*\*Radiotherapy Service, São Lucas Hospital, Pontifical Catholic University of  
Rio Grande do Sul- PUCRS, Brazil.**

**Corresponding address:**

Fernanda Gonçalves Salum

Pontifícia Universidade Católica do Rio Grande do Sul - PUCRS

Hospital São Lucas

Av. Ipiranga, 6690 – Room 231

CEP: 90610-000 - Porto Alegre – RS – Brazil

Tel/Fax: +55 51 3320-3254

E-mail: [fernanda.salum@pucrs.br](mailto:fernanda.salum@pucrs.br)

## ABSTRACT

**Objective:** The present pilot study aimed to assess the effect of low level laser therapy (LLLT) on hyposalivation and xerostomia due to head and neck radiotherapy. **Background:** Major salivary glands are commonly involved at the radiation sites as they are close to the primary tumor and lymph chain in the head and neck region. As a consequence of the radiotherapy, they go through a degenerative process which results in salivary hypofunction. **Methods:** Twenty three patients with a history of head and neck malignancy and treated by fractionated teletherapy (dosimetry ranging from 45 to 70 Gy), whose therapeutic site had involved at least 50% of the major salivary glands were selected. Patients were randomly distributed into laser group (n=12) and control group (n=11). A AsAlGa laser was used punctually in the major salivary glands, twice a week for six weeks, with a 12-session total. Stimulated and unstimulated salivary flow rate (SFR) was assessed, as well as the xerostomia and quality of life related to oral health (QLROH). **Results:** Analysis has not shown any significant difference between the groups with regards to the SFR and xerostomia, and the QLROH. However, at the end of the treatment, the xerostomia and the QLROH showed significant improvement in both groups compared to assessments carried out in baseline, highlighting the importance of advice given to the irradiated patients and their follow-up. **Conclusion:** In the parameters used, LLLT was not able to increase SFR or decrease xerostomia. The results may be associated to the late effects of radiotherapy on glandular structure such as fibrosis and acinar atrophy.

**Key Words:** low-level laser therapy, head and neck cancer, radiotherapy, phototherapy.

## INTRODUCTION

The major salivary glands are commonly involved at the radiation sites as they are close to primary tumor and lymph chains of the head and neck region. As a consequence of radiotherapy they go through a degenerative process resulting in hyposalivation and xerostomia.<sup>1</sup> Approximately 70% of the irradiated patients have developed such alterations<sup>2</sup>, with several complications, such as total or partial loss of taste, mouth burning and pain, susceptibility to oral ulcerations, cavities and other infections, dysphagia and dysphonia and even psychological alterations that negatively influence their quality of life.<sup>3</sup>

The dose of ionizing radiation, amount of salivary tissue exposed and patient's individual response are the main factors influencing glandular alterations.<sup>4-6</sup> Damage becomes irreversible after cumulative doses ranging from 26 to 39 Gy, and the salivary flow rate (SFR) can get under 10% from the one presented before radiation.<sup>7,8</sup> Despite being stable, as they do not have high mitotic rates, acinar cells respond quickly to radiation.<sup>9,10</sup> The mechanisms that lead to tissue destruction and salivary glands radiosensitivity have not been totally understood so far. Salivary gland alterations start with the damage to the plasmatic membrane, with loss of response to the autonomic controls, and progression to edema, degeneration and acinar cell apoptosis. Acute effects start 24 hours after therapy starts and stabilize within 72 hours. Late effects are the consequence of fibrosis and acinar atrophy<sup>11,12</sup>, which occur due to mesenchymal alterations, including changes in the extra cellular matrix, specifically in the laminin and in collagen IV.<sup>13,14</sup>

Low level laser therapy (LLLT) is a simple low cost tool that can be used as an adjuvant to conventional treatments, or alone and electively in some diseases.<sup>15-18</sup> Its

effects are based on the modulation of several metabolic, biochemical and photophysical processes that transform laser light into useful energy for the cell. The LLLT effect on xerostomia has been investigated in clinical and pre-clinical studies.<sup>16,17,19,20</sup> Simões et al.<sup>16</sup> demonstrated that LLLT increased the SFR and the levels of salivary proteins in Wistar rats. In patients irradiated in the head and neck region, Lopes et al.<sup>19</sup> and Simões et al.<sup>20</sup> showed that LLLT decreased xerostomia and increased stimulated SFR.

The objective of the present pilot study was to clinically assess LLLT effect on radiotherapy-induced hyposalivation and xerostomia. Besides, the influence of this therapeutic modality on the quality of life related to oral health was also assessed.

## PATIENTS AND METHODS

Fifty-one patients treated with radiotherapy in head and neck region were assessed consecutively; these patients came from Rio Grande do Sul State Radiotherapy Services, Brazil (São Lucas Hospital - Pontifical Catholic University of Rio Grande do Sul, Santa Casa Hospital Complex, Bruno Born Hospital, Tacchini Hospital and Charity Hospital). From these, 23 individuals between 37 and 69 years old were selected; all with a history of head and neck malignancy, treated with ionizing radiation, through fractioned teletherapy, with dosimetry ranging from 45 to 70 Gy, whose therapeutic site had involved at least 50% of the major salivary glands (uni or bi-laterally). Patients should have been followed up for at least six months after radiotherapy (with no relapses or metastasis), present Karnofsky<sup>21</sup> performance scale higher or equal to 60, present xerostomia and hyposalivation (unstimulated salivary

flow rate below 0.1 mL/min and, under stimulation, below 0.7 mL/min). All the selected patients signed the Informed Consent Form.

The individuals were randomly distributed into laser group (n=12) and control group (n=11). All the patients received instructions regarding oral hygiene, mucosal hydration and were advised to avoid spicy and citric foods consumption, as well as alcoholic beverages and tobacco. The study flowchart is shown in figure 1.

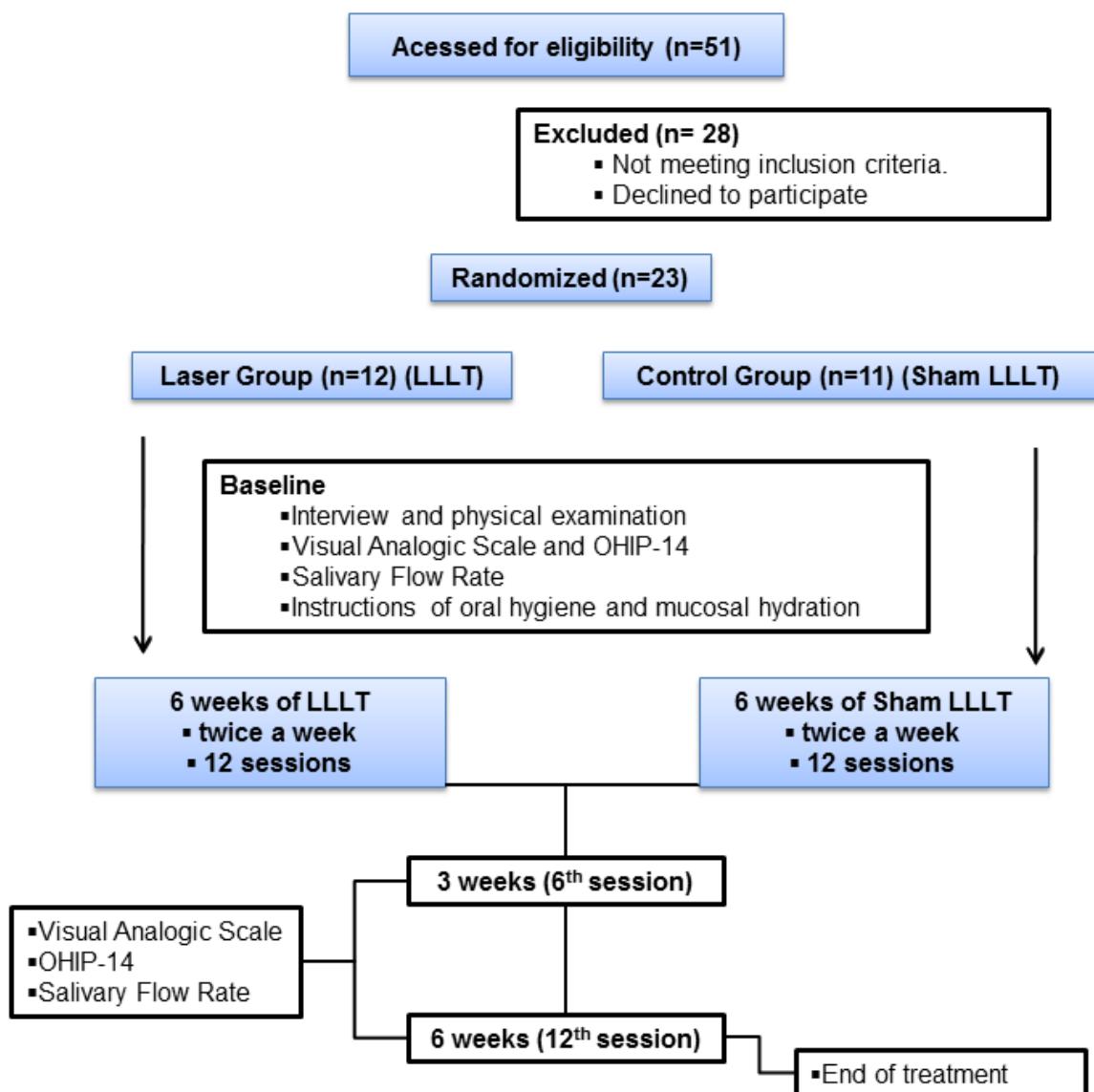


Figure 1. Flowchart representing the stages of the study.

### **Xerostomia and Salivary Flow Rate (SFR)**

Visual Analogic Scale (VAS) developed by Pai et al.<sup>22</sup>, consisting of eight items related to xerostomia and quantified by 10cm-long horizontal lines, was used for xerostomia assessment.

Stimulated and unstimulated whole SFR was determined. Samplings were collected in the morning, between 8:00 and 11:00 a.m., in previously weighed polypropylene vials. Patients were advised not to ingest food or drink, not to use cosmetics or drugs on the lips, and not to smoke or undergo physical stress one hour before the procedure. For unstimulated saliva sampling, patients were advised to keep their lips slightly open allowing saliva to run passively into a vial for 15 minutes. For stimulated salivary sampling a 1.0 cm long and 0.5 cm diameter latex cylinder held by dental floss was used. Stimulation was carried out for 5 minutes, and at each minute the content collected was deposited into the vial. The content of both samplings was weighed on a precision scale and transformed into mL/min.<sup>23,24</sup>

Xerostomia and SFR were assessed in baseline, after the sixth session and at the end of the 12<sup>th</sup> treatment session.

### **Quality of Life Related to Oral Health (QLROH)**

The QLROH was assessed through the Oral Health Impact Profile (OHIP-14) questionnaire, Portuguese language version.<sup>25</sup> This instrument was also applied in baseline, after the sixth session and after the 12<sup>th</sup> treatment session.

### **Low Level Laser Therapy (LLLT)**

A AsAlGa diode laser was used (Thera Lase® DMC Equipamentos Ltda., São Carlos, SP, Brazil) according to the following parameters: 830 nm (infrared)

wavelength, 100 mW power, continuous emissions, 2 J energy per point, application time 20 seconds per point. The area of the spot tip of this tool is 0.03 cm<sup>2</sup>. Patients underwent two LLLT weekly sessions for six weeks, total of 12 sessions. Before each application the power of the tool was calibrated and then, checked by means of a power meter.

LLLT was carried out in the major salivary glands punctually. Three points were applied on each parotid gland, two on each submandibular and two on each sublingual gland<sup>19</sup>. The tip of the tool remained in contact with the patients' skin in the applications on the parotid and submandibular glands. On the points meant for the sublingual glands, the tip was placed on the floor of the mouth.

The same number of sessions and application protocol were carried out with the control group as with the laser group; however, the tool received a plastic tip with rubber interior that blocked radiation emission, which was confirmed by means of a power meter before the applications in the control group.

## **STATISTICAL ANALYSIS**

Stimulated and unstimulated SFR, as well as the scores from the quality of life related to oral health (OHIP-14) were compared between the groups by the Mann-Whitney test. In each group, in order to compare the SFR and the quality of life scores obtained in baseline, after the LLLT 6<sup>th</sup> and 12<sup>th</sup> session, Friedman test was used complemented by its multiple comparison test. The VAS scores were assessed by using the repeated-measures analysis of variance (ANOVA) complemented by the Tukey multiple comparison test. Values P≤0.05 were considered significant.

## RESULTS

### Characterization of the sample

The demographic characteristics of the sample, the treatments for the malignancies, systemic alterations and drugs used during the study are shown in Table 1. The time between radiotherapy and the beginning of the study was about 46 months in the control group and 40 months in the laser group. Tobacco and alcohol had been used by 73.91% and 52.17% of the patients, respectively, up till the moment malignant neoplasia was diagnosed. At the time they were included in the study, all patients had already interrupted the use of those substances.

Table 1. Demographic distribution of the patients and characteristics of the treatment within the groups studied.

Characteristic	Laser Group n=12	Control Group n=11
<b>Sex</b>		
Male	6 (50%)	9 (81.81%)
Female	6 (50%)	2 (18.18%)
<b>Age</b>		
Range	44-69	37-66
Mean(±SD)	58.66 ±9.07	55.63 ±8.65
<b>Treatment</b>		
Radiotherapy + Chemotherapy	3 (25%)	3 (27.27%)
Radiotherapy + Surgery	4 (33.33%)	6 (54.54%)
Radiotherapy + Surgery + Chemotherapy	5 (41.66%)	2 (18.18%)
<b>Submandibular Excision</b>		
No	4 (33.33%)	3 (27.27%)
Both	2 (16.66%)	-
Right	4 (33.33%)	4 (36.36%)
Left	2 (16.66%)	4 (36.36%)
<b>Systemic Disease</b>		
No	4 (33.33%)	7 (63.63%)
Hypertension	5 (41.66%)	3 (27.27%)
Diabetes	2 (16.66%)	1 (9.09%)
Hypothyroidism	3 (25%)	1 (9.09%)
Other	2 (16.66%)	1 (9.09%)
<b>Systemic Medications</b>		
No	3 (25%)	6 (54.54%)
Thyroid Hormone	3 (25%)	1 (9.09%)
Analgesic	2 (16.66%)	-
Antihypertensive	1 (8.33%)	1 (9.09%)
Antilipidemic	2 (16.66%)	1 (9.09%)
Anti-diabetic	1 (8.33%)	1 (9.09%)
Antidepressants	1 (8.33%)	1 (9.09%)

### **Xerostomia and Salivary Flow Rate (SFR)**

Xerostomia scores, which were assessed by VAS, presented no significant difference between the groups in both baseline, after the 6<sup>th</sup> session, or at the end of the treatment (12<sup>th</sup> session). However, both in the laser group and in the control group, there was a significant decrease in xerostomia at the end of the 12<sup>th</sup> session of the treatment if compared with the assessment carried out in baseline (Table 2).

The stimulated and unstimulated SFR did not differ significantly between the groups in baseline, after the 6<sup>th</sup> session, or at the end of the treatment (12<sup>th</sup> session). When comparing the SFR of each group at different experimental times, a significant increase in unstimulated SFR was observed at the end of the treatment in the control group (Table 3).

Table 2. Visual analogic scale (VAS) scores for xerostomia in the laser group and control group in baseline, after the 6<sup>th</sup> and 12<sup>th</sup> LLLT sessions.

<b>Assessment</b>	<b>Control Group n=11</b>		<b>Laser Group n=12</b>	
	Mean	DP	Mean	DP
Baseline	6.51 <sup>A</sup>	1.43	6.30 <sup>A</sup>	1.67
6 <sup>th</sup> session	6.40 <sup>A</sup>	1.33	4.74 <sup>A</sup>	1.79
12 <sup>th</sup> session	4.65 <sup>B</sup>	1.96	4.15 <sup>B</sup>	2.63

Means followed by different letters in the column differ significantly through ANOVA test, using repeated measurements design, complemented by Tukey multiple comparison test, with 5% significance level.

Table 3. Stimulated and unstimulated salivary flow rate (SFR) (mL/min) in the laser group and control group in baseline, after the 6<sup>th</sup> and 12<sup>th</sup> LLLT sessions.

SFR (mL/min)	Control-group		Laser-group		P*
	n=11	Median (P25 – P75)	Rank**	Median (P25 – P75)	
Unstimulated	Baseline	0.025 (0.007 – 0.036)	1.64 <sup>B</sup>	0.013 (0.006 – 0.025)	1.75 <sup>A</sup> 0.347
	6 <sup>th</sup> session	0.017 (0.014 – 0.043)	1.73 <sup>AB</sup>	0.016 (0.004 – 0.043)	2.08 <sup>A</sup> 0.608
	12 <sup>th</sup> session	0.028 (0.013 – 0.055)	2.64 <sup>A</sup>	0.019 (0.009 – 0.046)	2.17 <sup>A</sup> 0.379
Stimulated	Baseline	0.111 (0.051 – 0.189)	2.09 <sup>A</sup>	0.063 (0.022 - 0.118)	1.75 <sup>A</sup> 0.288
	6 <sup>th</sup> session	0.129 (0.078 – 0.217)	2.09 <sup>A</sup>	0.073 (0.023 – 0.169)	2.17 <sup>A</sup> 0.379
	12 <sup>th</sup> session	0.110 (0.049 – 0.133)	1.82 <sup>A</sup>	0.103 (0.021 – 0.157)	2.08 <sup>A</sup> 0.695

\* Comparison between groups: Mann-Whitney's test

\*\* Comparison between times of the study: mean ranks followed by different letters in the column, differ significantly in the Friedman non-parametric test, complemented by its multiple comparison test, at a 5%significance level.

### Quality of Life Related to Oral Health (QLROH)

The QLROH, assessed through OHIP-14, did not differ significantly between the groups in baseline, after the 6<sup>th</sup> session, or at the end of the treatment (12<sup>th</sup> session). Both in the laser group and the control group, there was a significant decrease in the OHIP-14 scores at the end of the treatment when compared to the assessment carried out in baseline (Table 4).

Table 4. Oral health impact profile (OHIP-14) scores for quality of life related to oral health assessment in the laser group and control group in baseline, after the 6<sup>th</sup> and 12<sup>th</sup> LLLT sessions.

OHIP-14	Control Group		P
	n=11	Median (P25 – P75)	
	Median (P25 – P75)		
Baseline	10.23 <sup>A</sup> (6.39 – 12.82)	10.48 <sup>A</sup> (6.82 – 14.00)	0.786
6 <sup>th</sup> session	5.17 <sup>B</sup> (2.28 – 10.69)	7.55 <sup>AB</sup> (5.65 – 11.19)	0.413
12 <sup>th</sup> session	3.53 <sup>B</sup> (0.66 – 10.44)	2.52 <sup>B</sup> (1.69 – 9.84)	0.976

Mann-Whitney's test significant at p≤0.05

## DISCUSSION

The present pilot study has investigated the LLLT effect on hyposalivation and xerostomia in patients with head and neck malignancies treated with radiotherapy. All these selected patients presented important salivary dysfunction as a consequence of radiotherapy. Although the literature show, in pre-clinical and clinical studies, the benefits of LLLT in salivary flow increase<sup>16,19,20,26-30</sup>, there is not any study investigating its effects on patients that have already ended radiotherapy and present hyposalivation and xerostomia as a sequela.

Low level laser radiation is a non-ionizing and non-invasive form of radiation, well tolerated by the tissues and with no mutagenic effects. LLLT effects on salivary glands have not been completely understood so far. Studies have shown an increase in the number of duct epithelial cell mitosis, and stimulation to protein synthesis in submandibular glands of rats.<sup>26,31</sup>. As a result of mitochondria stimulation, increase in ATP levels, increase in glucose consumption by the cells, and the intracellular calcium level, LLLT can promote cell proliferation, increase in the anti-apoptotic protein expression as well as in blood micro-circulation in the salivary glands.<sup>19,29,32,33</sup> However, the present study has not confirmed the hypothesis that LLLT could stimulate residual gland function in patients submitted to head and neck radiotherapy. No significant difference was observed between laser and control groups regarding the SFR and OHIP-14 scores and VAS. When using LLLT in head and neck radiated patients, Cowen et al.<sup>34</sup>, Lopes et al.<sup>19</sup>, Simões et al.<sup>20</sup> and Oton-leite et al.<sup>30</sup> observed decrease in xerostomia and increase in the SFR, suggesting lasertherapy as an adjuvant in the treatment of salivary alterations due to radiotherapy. However, in those studies, the LLLT was applied concurrently to

radiotherapy, i.e., when irreversible morphological alterations such as, acinar atrophy and fibrosis had not been produced in the major salivary glands yet. In contrast, in the present study pilot, patients had ended radiotherapy for at least six months thus, the negative results obtained with regards to LLLT, can be attributed to late alterations due to ionizing radiation in the glandular structure. All patients had received ionizing radiation doses ranging from 45 to 70 Gy, which several authors<sup>35-37</sup> consider irreversible for glandular function restoration.

At the end of the treatment, however, there was a decrease in xerostomia and a improvement in the QLROH in both groups. It is important to highlight that all patients were advised, early in the experiment, about the importance of oral mucosa hydration, dental hygiene care and frequent stimulus of the salivary glands. Those recommendations were reinforced in each of the 12 sessions regardless of the group the patient was in. The assistance to the individuals, carried out during the six weeks of the study, was the determining factor for the improvement of xerostomia and QLROH, showing the need of a follow-up for the head and neck irradiated patients, in order to manage the sequelae caused by radiotherapy. Decrease in xerostomia was not followed by a clinically significant increase in the amount of saliva. Dawes<sup>38</sup> and Jensen et al.<sup>39</sup> suggest that xerostomia does not necessarily present correlation with the SFR. Bhide et al.<sup>40</sup> mention that the difference between the patient perception and the salivary flow could suffer from the influence of external factors.

Although there was no difference in the SFR between the laser and control groups, a significant increase in the unstimulated SFR was observed in the control group at the end of the experiment, in relation to the baseline assessment. With regards to the stimulated whole saliva, the values obtained at the end of the study did not differ from the baseline values in both groups. The result of the unstimulated SFR

in the control group, in spite of being statistically significant, did not have any clinical significance once the values were much below what is considered normal. The median of the unstimulated SFR in that group was 0.028 mL/min at the end of the study, while the normal value is 0.1 mL/min. During randomization of the sample, the laser group presented two patients with bilateral neck dissection and excision of both submandibular glands, while the control group did not have patients in such condition. Taking into consideration that the submandibular glands supply two thirds of unstimulated whole saliva, that factor might have justified the difference in unstimulated SFR in the control group.

The lack of an LLLT therapeutic protocol made these study methodological definitions difficult. In the literature, the power, power density, wave length, and all other parameters differ considerably in the studies that used LLLT for xerostomia treatment.<sup>19,20</sup> We have opted for using the infrared wave length due to the depth of the glandular parenchyma to be irradiated<sup>17</sup>. The frequency of the sessions in the other studies ranged from once a week<sup>20</sup> to daily applications<sup>19</sup>. In our study the sessions were carried out twice a week for six weeks with the objective of keeping a steady cell response. Furthermore, as some authors mention that LLLT can stimulate not only the healthy cells but also the tumor cells.<sup>41-43</sup> Hence, during LLLT applications, the areas close to the already treated tumor lesions were avoided.

As xerostomia has serious consequences on the patients quality of life, we have opted for assessing the QLROH, using the OHIP-14. Nowadays, several tools are available for that purpose, not a single one considered gold-standard<sup>44</sup>, though. Taking into account the decrease in the OHIP-14 scores along the six-week-study, the relation between xerostomia relief perception and improvement in the quality of life of the individuals in the research is evident.

Saliva plays an important role in the oral and general homeostasis, once salivary dysfunctions predispose individuals to several complications. Radiotherapy carried out in the head and neck region is one of the main causes of salivary disorders, interfering negatively in the patients' quality of life. Some studies have evidenced the LLLT benefits in the gland function during the radiotherapy treatment. However, the results of the present pilot study show that in patients submitted to a 45 Gy minimum dosimetry in head and neck, 12 LLLT sessions applied for six weeks, were unable to promote salivary flow increase. The degree of cell destruction due to teletherapy, especially on the parotids, might have been one of the main factors associated with the results obtained. Nevertheless, xerostomia and quality of life showed improvement, thus highlighting the importance of advice and follow-up to the irradiated patients. New therapeutical modalities, especially in tissue engineering, must be further investigated in order to restore the gland function and, therefore, improve the quality of life of the patients with radiotherapy sequelae.

## **CONCLUSIONS**

In the parameters used, LLLT was not able to increase SFR, or decrease xerostomia, hence the hypothesis that LLLT could stimulate residual gland function in patients treated with head and neck radiotherapy has not been confirmed. The results may be associated to the late effects of radiotherapy on glandular structure such as fibrosis and acinar atrophy.

## ACKNOWLEDGMENTS

The authors thank National Council for Scientific and Technological Development (CNPQ) and Radiotherapy Service of São Lucas Hospital (PUCRS, Brazil) for their contributions to the development of research.

## AUTHOR DISCLOSURE STATEMENT

No competing financial interests exist.

## REFERENCES

1. Eisbruch, A., Kim, H.M., Terrell, J.E., Marsh, L.H., Dawson, L.A., and Ship, J.A. (2001). Xerostomia and its predictors following parotid-sparing irradiation of head-and-neck cancer. *Int. J. Radiat. Oncol. Biol. Phys.* 50, 695-704.
2. Mira, J.G., Wescott, W.B., Starcke, E.N., and Shannon, I.L. (1981). Some factors influencing salivary function when treating with radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* 7, 535-541.
3. Dirix, P., Nuyts, S., Vander Poorten, V., Delaere, P., and Van den Bogaert, W. (2008). The influence of xerostomia after radiotherapy on quality of life: results of a questionnaire in head and neck cancer. *Support Care Cancer.* 16, 171-179.
4. Deasy, J.O., Moiseenko, V., Marks, L., Chao, K.S., Nam, J., and Eisbruch, A. (2010). Radiotherapy dose-volume effects on salivary gland function. *Int. J. Radiat. Oncol. Biol. Phys.* 76, S58-63.
5. Li, Y., Taylor, J.M., Ten Haken, R.K., and Eisbruch, A. (2007). The impact of dose on parotid salivary recovery in head and neck cancer patients treated with radiation therapy. *Int. J. Radiat. Oncol. Biol. Phys.* 67, 660-669.

6. Murdoch-Kinch, C.A., Kim, H.M., Vineberg, K.A., Ship, J.A., and Eisbruch, A. (2008). Dose-effect relationships for the submandibular salivary glands and implications for their sparing by intensity modulated radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* 72, 373-382.
7. Vissink, A., Mitchell, J.B., Baum, B.J., et al (2010). Clinical management of salivary gland hypofunction and xerostomia in head-and-neck cancer patients: successes and barriers. *Int. J. Radiat. Oncol. Biol. Phys.* 78, 983–991.
8. Porter, S.R., Fedele, S., and Habbab, K.M. (2010). Xerostomia in head and neck malignancy. *Oral Oncol.* 46, 460–463.
9. Baum, B.J., Kousvelari, E.E., and Oppenheim, F.G. (1982). Exocrine protein secretion from human parotid glands during aging: stable release of the acidic proline-rich proteins. *J. Gerontol.* 37, 392-395.
10. Grundmann, O., Mitchell, G.C., and Limesand, K.H. (2009). Sensitivity of salivary glands to radiation: from animal models to therapies. *J. Dent. Res.* 88, 894-903.
11. Porter, S.R. (2010). Xerostomia: prevalence, assessment, differential diagnosis and implications for quality of life. *Oral Dis.* 16, 501–502.
12. Gustafsson, H., Aalto, Y., Franzén, L., Thornell, L.E., and Henriksson, R. (1998). Effects of fractionated irradiation on the cytoskeleton and basal lamina in parotid glands--an immunohistochemical study. *Acta Oncol.* 37, 33-40.
13. Hakim, S.G., Jacobsen, H.Ch., Hermes, D., et al. (2004). Early immunohistochemical and functional markers indicating radiation damage of the parotid gland. *Clin. Oral Investig.* 8, 30-35.
14. Avila, J.L., Grundmann, O., Burd, R., and Limesand, K.H. (2009). Radiation-induced salivary gland dysfunction results from p53-dependent apoptosis. *Int. J. Radiat. Oncol. Biol. Phys.* 73, 523-529.

15. Lucas, C., Criens-Poublon, L.J., Cockrell, C.T., and de Haan, R.J. (2002). Wound healing in cell studies and animal model experiments by Low Level Laser Therapy; were clinical studies justified? a systematic review. *Lasers Med. Sci.* 17, 110-134.
16. Simões, A., Nicolau, J., de Souza, D.N., et al. (2008). Effect of defocused infrared diode laser on salivary flow rate and some salivary parameters of rats. *Clin. Oral Investig.* 12, 25-30.
17. Lončar, B., Stipetić, M.M., Baričević, M., and Risović, D. (2011). The effect of low-level laser therapy on salivary glands in patients with xerostomia. *Photomed. Laser Surg.* 29, 171-175.
18. Jenkins, P.A., and Carroll, J.D. (2011). How to report low-level laser therapy (LLLT)/photomedicine dose and beam parameters in clinical and laboratory studies. *Photomed. Laser Surg.* 29, 785-787.
19. Lopes, C.O., Mas, J.R., and Zângaro, R.A. (2006). Low-level laser therapy in the prevention of radiotherapy-induced xerostomia and oral mucositis. *Radiol. Bras.* 39, 131-136.
20. Simões, A., de Campos, L., de Souza, D.N., de Matos, J.A., Freitas, P.M., and Nicolau, J. (2010). Laser phototherapy as topical prophylaxis against radiation-induced xerostomia. *Photomed. Laser Surg.* 28, 357-363.
21. Su, C.C., Lee, K.D., Yeh, C.H., Kao, C.C., and Lin, C.C. (2013). Measurement of physical activity in cancer survivors: a validity study. *J. Cancer Surviv.* 7.
22. Pai, S., Ghezzi, E.M., and Ship, J.A. (2001). Development of a visual analogue scale questionnaire for subjective assessment of salivary dysfunction. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 91, 311-316.

23. Cherubini, K., Lorandi, C.S., Krapf, S.M., et al. (2006). Association between recurrent aphthous stomatitis and salivary thiocyanate levels. *J. Oral Sci.* 48, 153-156.
24. Navazesh, M., and Kumar, S.K. (2008). Measuring salivary flow: challenges and opportunities. *J. Am. Dent. Assoc.* 139, Suppl:35S-40S.
25. Oliveira, B.H., and Nadanovsky, P. (2005). Psychometric properties of the Brazilian version of the Oral Health Impact Profile-short form. *Community Dent. Oral Epidemiol.* 33, 307-314.
26. Takeda, Y. (1988). Irradiation effect of low-energy laser on rat submandibular salivary gland. *J. Oral Pathol.* 17, 91–94.
27. Campos, L., Simões, A., Sá, P.H., and Eduardo, C.P. (2009). Improvement in quality of life of an oncological patient by laser phototherapy. *Photomed. Laser Surg.* 27, 371-374.
28. Vidović Juras, D., Lukac, J., Cekić-Arambasin, A., et al. (2010). Effects of low-level laser treatment on mouth dryness. *Coll. Antropol.* 34, 1039-1043.
29. Onizawa, K., Muramatsu, T., Matsuki, M., et al. (2009). Low-level (gallium-aluminum-arsenide) laser irradiation of Par-C10 cells and acinar cells of rat parotid gland. *Lasers Med. Sci.* 24, 155-161.
30. Oton-Leite, A.F., Elias, L.S., Morais, M.O., et al. (2013). Effect of low level laser therapy in the reduction of oral complications in patients with cancer of the head and neck submitted to radiotherapy. *Spec. Care Dentist.* 33, 294-300.
31. Plavnik, L.M., De Crosa, M.E., and Malberti, A.I. (2003). Effect of low-power radiation (helium/neon) upon submandibular glands. *J. Clin. Laser Med. Surg.* 21, 219–225.

32. Karu, T. (1989). Laser biostimulation: a photobiological phenomenon. *J. Photochem. Photobiol. B.* 3, 638-640.
33. Maegawa, Y., Itoh, T., Hosokawa, T., Yaegashi, K., and Nishi, M. (2000). Effects of near-infrared low-level laser irradiation on micro-circulation. *Lasers Surg. Med.* 27, 427–437.
34. Cowen, D., Tardieu, C., Schubert, M. et al. (1997). Low energy helium-neon laser in the prevention of oral mucositis in patients undergoing bone marrow transplant: results of a double blind randomized trial. *Int. J. Radiat. Oncol. Biol. Phys.* 38, 697-703.
35. Franzén, L., Funegård, U., Ericson, T., and Henriksson, R. (1992). Parotid gland function during and following radiotherapy of malignancies in the head and neck. A consecutive study of salivary flow and patient discomfort. *Eur. J. Cancer.* 28, 457-462.
36. Funegård, U., Franzén, L., Ericson, T., and Henriksson, R. (1994). Parotid saliva composition during and after irradiation of head and neck cancer. *Eur. J. Cancer B. Oral Oncol.* 30B, 230-233.
37. Someya, M., Sakata, K., Nagakura, H., Nakata, K., Ouchi, A., and Hareyama, M. (2003). The changes in irradiated salivary gland function of patients with head and neck tumors treated with radiotherapy. *Jpn. J. Clin. Oncol.* 33, 336-340.
38. Dawes, C. (2008). Salivary flow patterns and the health of hard and soft oral tissues. *J. Am. Dent. Assoc.* 139, 18S-24S.
39. Jensen, S.B., Pedersen, A.M., Reibel, J., and Nauntofte, B. (2003). Xerostomia and hypofunction of the salivary glands in cancer therapy. *Support. Care Cancer.* 11, 207-225.

40. Bhide, S.A., Miah, A.B., Harrington, K.J., Newbold, K.L., and Nutting, C.M. (2009). Radiation-induced xerostomia: pathophysiology, prevention and treatment. *Clin. Oncol. (R. Coll. Radiol.)* 21, 737-744.
41. Pinheiro, A.L., Carneiro, N.S., Vieira, A.L., et al. (2002). Effects of low-level laser therapy on malignant cells: in vitro study. *J. Clin. Laser Med. Surg.* 20, 23-26.
42. Kreisler, M., Christoffers, A.B., Willershausen, B., and d'Hoedt, B. (2003). Low-level 809 nm GaAlAs laser irradiation increases the proliferation rate of human laryngeal carcinoma cells in vitro. *Lasers Med. Sci.* 18, 100-103.
43. Monteiro, J.S., Pinheiro, A.N., de Oliveira, S.C., et al. (2011). Influence of laser phototherapy ( $\lambda$ 660 nm) on the outcome of oral chemical carcinogenesis on the hamster cheek pouch model: histological study. *Photomed. Laser Surg.* 29, 741-745.
44. Vartanian J.G., Carvalho A.L., Furia, C.L. et al. (2007). Questionnaires validated in the Brazilian population for evaluation of the quality of life in patients with head and neck cancer. *Rev. Bras. Cir. Cabeça PESCOÇO.* 36, 108 -115.

## 5 DISCUSSÃO GERAL

A saliva é de suma importância para a manutenção da homeostase oral e geral, desempenhando papel crucial nas funções digestiva e gustativa, limpeza e hidratação da mucosa bucal, proteção dos dentes e controle da microflora oral devido às suas propriedades antibacteriana, antifúngica e antiviral (AMERONGEN; VEERMAN, 2002; PORTER; SCULLY; HEGARTY, 2004; FARSI, 2007; THELIN, et al., 2008; FUJIMAKI, et al., 2014). Diversos fatores estão envolvidos nas disfunções salivares tais como fármacos (SREEBNY; SCHWARTZ, 1997; SCULLY, 2003; TURNER; JAHANGIRI; SHIP, 2008), alterações relacionadas ao envelhecimento (PERCIVAL; CHALLACOMBE; MARSH, 1994, TYLENDAL; SHIP; FOX, 1988), doenças sistêmicas (GUGGENHEIMER; MOORE, 2003), radioterapia direcionada à região de cabeça e pescoço (EISBRUCH, et al., 2001; JENSEN, et al., 2010; KONINGS; COPPES; VISSINK, 2005), dentre outros.

Os mecanismos pelos quais a radiação ionizante causa danos às glândulas salivares permanecem pobemente compreendidos. Konings, Copes e Vissink (2005) sugerem que a injúria esteja relacionada ao dano a membrana plasmática das células glandulares. Os radicais livres formados pela ação da radiação ionizante atuam sobre os canais lipídicos da membrana, alterando a ação dos receptores muscarínicos. Em uma fase tardia o dano à função glandular é marcado por redução na quantidade de células acinares funcionais, causado pela morte de células progenitoras e células-tronco (KONINGS; COPPES; VISSINK, 2005).

Apesar dos esforços na tentativa de estabelecer-se um protocolo de tratamento para a xerostomia e hipossalivação decorrentes da radioterapia, não há uma terapêutica-padrão. O presente estudo propôs o emprego da TLBP para o

manejo de disfunções salivares decorrentes da radioterapia. Este estudo diferencia-se dos demais que empregaram a TLBP em pacientes irradiados pela utilização dessa modalidade terapêutica em indivíduos que haviam finalizado a radioterapia há pelo menos seis meses. Foram selecionados 23 pacientes submetidos a regime radioterápico semelhante para tratamento de neoplasias malignas na região de cabeça e pescoço. Quanto à composição dos grupos, apesar da randomização, no grupo-controle houve predomínio de pacientes do sexo masculino, enquanto no grupo-laser o número de pacientes dos sexos masculino e feminino foi semelhante. Dois pacientes do grupo-laser sofreram excisão bilateral das glândulas submandibulares e seis foram submetidos à excisão unilateral desta glândula durante o tratamento cirúrgico da neoplasia maligna. No grupo-controle nenhum dos 11 pacientes sofreu excisão de ambas as submandibulares e oito indivíduos foram submetidos à excisão unilateral. Nos demais itens tais como idade, alterações sistêmicas e uso de medicamentos a distribuição dos grupos demonstrou homogeneidade.

Os efeitos da teleterapia sobre as glândulas salivares são observados desde o início do tratamento e os danos passam a ser irreversíveis após doses cumulativas de 26 a 39 Gy (VISSINK et al., 2010; PORTER; FEDELE; HABBAB, 2010). Dos 23 pacientes da amostra deste estudo, 15 receberam dose de radiação de 70 Gy, além de portais terapêuticos bilaterais (com irradiação de ambas as parótidas), acarretando níveis severos de hipossalivação, muitas vezes próximos à completa ausência de saliva (DE BARROS PONTES; POLIZELLO; SPADARO, 2004).

Conforme discutido no artigo de pesquisa, a hipótese de que a TLBP pudesse estimular a função glandular residual nos pacientes irradiados não foi confirmada no estudo. Não foi observada diferença significativa entre os grupos laser e controle

quanto à velocidade do fluxo salivar, em repouso e sob estimulação, ou quanto aos escores de xerostomia e qualidade de vida. Em estudos prévios, a TLBP foi aplicada concomitantemente à radioterapia (COWEN et al., 1997; LOPES; MAS; ZÂNGARO, 2006; SIMÕES et al., 2010; OTON-LEITE et al., 2013), promovendo redução da xerostomia e elevação do fluxo salivar. Na presente investigação, a TLBP foi empregada após o término da radioterapia, quando os pacientes já apresentavam importante disfunção salivar. Podem-se atribuir os resultados negativos da TLBP sobre o fluxo salivar às alterações morfológicas promovidas pela radiação ionizante na estrutura glandular tais como atrofia acinar e fibrose.

Entretanto, em ambos os grupos, houve redução da xerostomia e melhora da qualidade de vida relacionada à saúde oral. Durante o estudo todos os pacientes foram esclarecidos sobre hipossalivação e xerostomia, receberam instruções de higiene oral e foram orientados a manter a mucosa bucal hidratada e a evitar o uso de alimentos condimentados, de bebidas com teor ácido e de fumo ou álcool. O acompanhamento dos pacientes durante o estudo foi o fator preponderante à melhora das variáveis subjetivas xerostomia e qualidade de vida.

Para avaliação da xerostomia empregamos a EVA, um instrumento bem estabelecido na mensuração da dor (AITKEN, 1969; NGAN; KESS; WILSON, et al., 1989; LIM, et al., 1995). Pai et al. (2001) e Jham et al. (2007) sugeriram este como um método seguro, de fácil aplicação e importante na mensuração da xerostomia. A xerostomia, mesmo que variando em sua intensidade, é um sintoma que inevitavelmente interfere no bem-estar dos pacientes irradiados (GUCHELAAR; VERMES; MEERWALDT, 1997), portanto, optamos por avaliar também a qualidade de vida relacionada à saúde oral. Levando-se em consideração a redução nos escores do OHIP-14 ao longo das seis semanas de estudo, fica evidente a relação

entre a percepção de alívio da xerostomia e a melhora na qualidade de vida dos pesquisados.

A TLBP ainda carece de padronização em pesquisas, uma vez que muitos estudos não descrevem adequadamente todos os parâmetros metodológicos, além da grande variação em relação aos aparelhos utilizados. Deve-se considerar também que não há estudos prévios empregando a TLBP em tecido glandular já irradiado e apresentando as alterações morfológicas descritas previamente. Esses fatores dificultaram a determinação da metodologia deste estudo, principalmente quanto à energia a ser empregada por ponto, número e frequência de sessões de TLBP. Como o objetivo desta terapia é entregar uma quantidade específica de energia ao tecido-alvo, o parâmetro energia pode ser considerado o principal a ser determinado nos estudos (GARCEZ; RIBEIRO; NÚÑEZ, 2012). As propriedades ópticas de um tecido em situação patológica diferem daquelas de um tecido sadio (GARCEZ; RIBEIRO; NÚÑEZ, 2012). No presente estudo a TLBP foi aplicada em um tecido glandular que apresentava uma série de alterações decorrentes da radiação ionizante. Em função destas alterações como fibrose e redução da celularidade, que poderiam acarretar em menor distribuição da energia aplicada, optou-se por utilizar 2J de energia por ponto, valor superior ao dos demais estudos em que a TLBP foi utilizada concomitante à radioterapia. Para obtenção de 2 J de energia utilizando-se potência de 100 mW e um aparelho com spot de 0,03 cm<sup>2</sup>, foi necessário aplicar-se dosimetria de 70 J/cm<sup>2</sup>. Em pesquisas prévias em ratos e em humanos submetidos à radioterapia, foram aplicadas de quatro a trinta sessões de TLBP (LOPES, MAS, ZÂNGARO, 2006; SIMÕES, et al., 2008; SIMÕES, et al., 2010; LONCAR, et al., 2011). No presente estudo determinamos a frequência de duas sessões semanais, em vez de sessões diárias a fim garantir a adesão dos pacientes

da amostra à terapia, uma vez que muitos necessitavam deslocar-se de outras cidades da região. Por tratar-se de um estudo-piloto optamos por não estender demasiadamente o período experimental, estipulado em seis semanas, totalizando 12 sessões de TLBP.

A radioterapia realizada em região de cabeça e pescoço é uma das principais causas de disfunções salivares. Visando prevenir tais alterações, alguns estudos têm evidenciado os benefícios da TLBP na função glandular quando empregada de forma concomitante à radioterapia. Na presente investigação, realizada em pacientes submetidos à dosimetria mínima de 45 Gy de radiação ionizante, os resultados demonstraram que 12 sessões de TLBP aplicadas durante seis semanas não promoveram elevação do fluxo salivar. As alterações tardias promovidas pela teleterapia nas glândulas salivares maiores tais como atrofia acinar e fibrose foi provavelmente o fator que determinou os resultados deste estudo. Por outro lado, a melhora dos índices subjetivos xerostomia e qualidade de vida ressalta a importância da orientação e acompanhamento dos pacientes irradiados. Novas modalidades terapêuticas, principalmente na área de engenharia tecidual, devem ser investigadas para promover o restabelecimento da função glandular e consequente melhora na qualidade de vida nos pacientes portadores de sequelas decorrentes da radioterapia.

O efeito da TLBP utilizada de forma preventiva, ou seja, concomitante à radioterapia também deve ser investigado em outras pesquisas pré-clínicas e clínicas no intuito de determinar as alterações histológicas e moleculares promovidas no tecido glandular, além de estabelecer um protocolo-padrão de laserterapia a ser utilizado nesses pacientes.

## REFERÊNCIAS

- AITKEN, R. C. Measurement of feelings using visual analogue scales. **Proc R Soc Med**, London, v. 62, n. 10, p. 989-93, Oct. 1969.
- AMERONGEN, A. V.; VEERMAN, E. C. Saliva--the defender of the oral cavity. **Oral Dis**, Hounds Mills, v. 8, n. 1, p. 12-22, Jan. 2002.
- ANKRI, R.; LUBART, R.; TAITELBAUM, H. Estimation of the optimal wavelengths for laser-induced wound healing. **Lasers Surg Med**, New York, v. 42, n. 8, p. 760-4, Oct. 2010.
- ATKINSON, J. C.; GRISIUS, M.; MASSEY, W. Salivary hypofunction and xerostomia: diagnosis and treatment. **Dent Clin North Am**, Philadelphia, v. 49, n. 2, p. 309-26, Apr. 2005.
- BHIDE, A. S. et al. The role of intensity modulated radiotherapy in advanced oral cavity carcinoma. **J Cancer Res Ther**, Mumbai, v. 8 suppl 1, p. 67-71, Jan. 2012.
- BRUGNERA-JUNIOR, A.; PINHEIRO, A. L. **Lasers na Odontologia Moderna**. São Paulo: Editora Pancast, 1998. 356p.
- CHOPRA, S. et al. Factors predictive of severity of osteoradionecrosis of the mandible. **Head Neck**, New York, v. 33, n. 11, p. 1600-5, Nov. 2011.
- CHREANOVIC, B. R. et al. Osteoradionecrosis of the jaws- a current overview- part 2: dental management and therapeutic options for treatment. **Oral Maxillofac Surg**, Berlin, v. 14, p. 81-95, June 2010.

COWEN, D. et al. Low energy helium-neon laser in the prevention of oral mucositis in patients undergoing bone marrow transplant: results of a double blind randomized trial. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 38, n. 4, p. 697-703, July 1997.

DE BARROS PONTES, C.; POLIZELLO, A. C.; SPADARO, A. C. Clinical and biochemical evaluation of the saliva of patients with xerostomia induced by radiotherapy. **Braz Oral Res.**, São Paulo, v. 18, n. 1, p. 69-74, Jan-Mar. 2004.

DIRIX, P. et al. The influence of xerostomia after radiotherapy on quality of life. **Support Care Cancer**, Berlin, v. 16, p. 171–9, Feb. 2008.

EISBRUCH, A. et al. Xerostomia and its predictors following parotid-sparing irradiation of head-and-neck cancer. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 50, n. 3, p. 695-704, July 2001.

FARSI, N. M. Signs of oral dryness in relation to salivary flow rate, pH, buffering capacity and dry mouth complaints. **BMC Oral Health**, London, v. 9, n. 7, p. 15, Nov. 2007.

FUJIMAKI, Y. et al. **Non-invasive objective evaluation of radiotherapy-induced dry mouth**. **J Oral Pathol Med**, Copenhagen, v. 43, n. 2, p. 97-102, Feb. 2014.

GARCEZ, A. S.; RIBEIRO, M. S.; NÚÑEZ, S. C. **Laser de baixa potência: princípios básicos e aplicações clínicas na odontologia**. Rio de Janeiro: Elsevier, 2012. 259 p.

GUCHELAAR, H. J.; VERMES, A.; MEERWALDT, J. H. Radiation-induced xerostomia: pathophysiology, clinical course and supportive treatment. **Support Care Cancer**, Berlin, v. 5, p. 281–8, July 1997.

GUGGENHEIMER, J.; MOORE, P. A. Xerostomia: etiology, recognition and treatment. **J Am Dent Assoc**, Chicago, v. 134, n. 1, p. 61-9, Jan. 2003.

JELLEMA, A. P. et al. Impact of radiation-induced xerostomia on quality of life after primary radiotherapy among patients with head and neck cancer. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 69, n. 3, p. 751-760, Nov. 2007.

JENKINS, P. A.; CARROLL, J. D. How to report low-level laser therapy (LLLT)/photomedicine dose and beam parameters in clinical and laboratory studies. **Photomed Laser Surg**, Larchmont, v. 29, n. 12, p. 785-7, Dec. 2011.

JENSEN, S. B. et al. A systematic review of salivary gland hypofunction and xerostomia induced by cancer therapies: management strategies and economic impact. **Support Care Cancer**, Berlin, v. 18, n. 8, p. 1061-79, Aug. 2010.

JHAM, B. C.; FREIRE, A. R. Oral complications of radiotherapy in the head and neck. **Rev Bras Otorrinolaringol**, São Paulo, v. 72, n. 5, p. 704-8, Sept.-Oct. 2006.

JHAM, B. C. et al. A randomized phase III prospective trial of bethanechol to prevent radiotherapy-induced salivary gland damage in patients with head and neck cancer. **Oral Oncol**, Oxford, v. 43, n. 2, 137-42, Feb. 2007.

KONINGS, A. W.; COPPES, R. P.; VISSINK, A. On the mechanism of salivary gland radiosensitivity. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 62, n. 4, p. 1187-94, July 2005.

LIM, H. M.; LEW, K. K.; TAY, D. K. A clinical investigation of the efficacy of low level laser therapy in reducing orthodontic postadjustment pain. **Am J Orthod Dentofacial Orthop**, St. Louis, v. 108, n. 6, p. 614-22, Dec. 1995.

LIN, S. C. et al. Assessment of xerostomia and its impact on quality of life in head and neck cancer patients undergoing radiotherapy, and validation of the Taiwanese version of the xerostomia questionnaire. **J Pain Symptom Manage**, Madison, v. 36, n. 2, p. 141-8, Aug. 2008.

LONCAR, B. et al. The effect of low-level laser therapy on salivary glands in patients with xerostomia. **Photomed Laser Surg**, Larchmont, v. 29, n. 3, p. 171-5, Mar. 2011.

LOPES, C. O.; MAS, J. R. I.; ZÂNGARO, R. A. Low-level laser therapy in the prevention of radiotherapy-induced xerostomia and oral mucositis. **Radiol Bras**, São Paulo, v. 39, n. 2, p. 131-6, 2006.

MÜNTER, M. W. et al. Changes in salivary gland function after radiotherapy of head and neck tumors measured by quantitative pertechnetate scintigraphy: comparison of intensity-modulated radiotherapy and conventional radiation therapy with and without amifostine. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 67, n. 3, p. 651-9, Mar. 2007.

NGAN, P.; KESS, B.; WILSON, S. Perception of discomfort by patients undergoing orthodontic treatment. **Am J Orthod Dentofacial Orthop**, St. Louis, v. 96, n. 1, p. 47-53, July 1989.

OTON-LEITE, A. F. et al. Effect of low level laser therapy in the reduction of oral complications in patients with cancer of the head and neck submitted to radiotherapy. **Spec Care Dentist**, Chicago, v. 33, n. 6, p. 294-300, Nov.-Dec. 2013.

PAI, S. et al. Development of a visual analogue scale questionnaire for subjective assessment of salivary dysfunction. **Oral Surg Oral Med Oral Pathol Oral Radiol Endod**, St Louis, v. 91, n. 3, p. 311-6, Mar. 2001.

PERCIVAL, R. S.; CHALLACOMBE, S. J.; MARSH, P. D. Flow rates of resting whole and stimulated parotid saliva in relation to age and gender. **J Dent Res**, Chicago, v. 73, n. 8, p. 1416-20, Aug. 1994.

PORTER, S. R. Xerostomia: prevalence, assessment, differential diagnosis and implications for quality of life. **Oral Dis**, v. 16, n. 6, p. 501–2, 2010.

PORTER, S. R.; FEDELE, S.; HABBAB, K. M. Xerostomia in head and neck malignancy. **Oral Oncol**, Oxford, v. 46, n. 6, p. 460–3, June 2010.

PORTER, S. R.; SCULLY, C.; HEGARTY, A. M. An update of the etiology and management of xerostomia. **Oral Surg Oral Med Oral Pathol Oral Radiol Endod**, St Louis, v. 97, n.1, p. 28-46, Jan. 2004.

RIEGER, J. M. et al. Functional outcomes related to the prevention of radiation-induced xerostomia: oral pilocarpine versus submandibular salivary gland transfer. **Head Neck**, New York, v. 34, n. 2, p. 168–74, Feb. 2012.

SCULLY, C. Drug effects on salivary glands: Dry mouth. **Oral Dis**, Hounds mills, v. 9, n. 4, p. 165–176, July 2003.

SEGRETO, H. R., SEGRETO, R. A. Radiobiology: review and update. Cellular, molecular and clinical aspects. **Folha Med**, v. 119, p. 9-27, 2000.

SEIKALY, H. et al. Long-term outcomes of submandibular gland transfer for prevention of postradiation xerostomia. **Arch Otolaryngol Head Neck Surg**, Chicago, v. 130, n. 8, p. 956-96, Aug. 2004.

SENNHENN-KIRCHNER, S. et al. Dental therapy before and after radiotherapy - an evaluation on patients with head and neck malignancies. **Clin Oral Invest**, Berlin, v. 13, n. 2, p. 157-64, June 2009.

SIMÕES, A. et al. Effect of defocused infrared diode laser on salivary flow rate and some salivary parameters of rats. **Clin Oral Invest**, Berlin, v. 12, n.1, p. 25–30, Mar. 2008.

SIMÕES, A. et al. Laser phototherapy as topical prophylaxis against radiation-induced xerostomia. **Photomed and Laser Surg**, Larchmont, v. 28, n.3, p. 357-63, June 2010.

SREEBNY, L. M.; SCHWARTZ, S. S. A reference guide to drugs and dry mouth--2nd edition. **Gerodontology**, Mount Desert, v. 14, n. 1, p.33-47, July 1997.

TEYMOORTASH, A. et al. Botulinum toxin prevents radiotherapy-induced salivary gland damage. **Oral Oncol**, Oxford, v. 45, n. 8, p. 737–9, Aug. 2009.

THELIN, W. R. et al. The oral mucosa as a therapeutic target for xerostomia. **Oral Dis**, Hounds Mills, v. 14, n.8, p. 683-9, Nov. 2008.

TURNER, M.; JAHANGIRI, L.; SHIP, J. A. Hyposalivation, xerostomia and the complete denture: A systematic review. **J Am Dent Assoc**, Chicago, v. 139, n. 2, p.146–150, Feb. 2008.

TYLEND, C. A. et al. Evaluation of submandibular salivary flow rate in different age groups. **J Dent Res.**, Chicago, v. 67, n. 9, p. 1225-8, Sept. 1988.

VISSINK, A. et al. Clinical management of salivary gland hypofunction and xerostomia in head-and-neck cancer patients: successes and barriers. **Int J Radiat Oncol Biol Phys**, Elmsford, v. 78, n. 4, p. 983-91, Nov. 2010.

WEBER, C. et al. Limited mouth opening after primary therapy of head and neck cancer. **Oral Maxillofac Surg**, Berlin, v. 14, n. 3, p. 169-73, Sept. 2010.

## ANEXO A

## APROVAÇÃO DA COMISSÃO CIENTÍFICA E DE ÉTICA DA FACULDADE DE ODONTOLOGIA DA PUCRS



*Comissão Científica e de Ética  
Faculdade da Odontologia da PUCRS*

---

*Porto Alegre 31 de outubro de 2012*

**O Projeto de: Dissertação**

**Protocolado sob nº:** 0052/12

**Intitulado:** Efeito clínico da laserterapia na hipossalivação e xerostomia decorrentes da radioterapia em região de cabeça e pescoço.

**Pesquisador Responsável:** Profa. Dra. Fernanda Gonçalves Salum

**Pesquisadores Associados:** Jamil Saleh

**Nível:** Dissertação / Mestrado

Foi **aprovado** pela Comissão Científica e de Ética da Faculdade de Odontologia da PUCRS em **31 de outubro de 2012**.

*Este projeto deverá ser imediatamente encaminhado ao CEP/PUCRS.*

**Profa. Dra. Ana Maria Spohr**

Coordenadora da Comissão Científica e de Ética da  
Faculdade de Odontologia da PUCRS

## ANEXO B

### APROVAÇÃO DO COMITÊ DE ÉTICA EM PESQUISA DA PUCRS

PONTIFÍCIA UNIVERSIDADE  
CATÓLICA DO RIO GRANDE  
DO SUL - PUC/RS



#### PARECER CONSUBSTANCIADO DO CEP

##### DADOS DO PROJETO DE PESQUISA

**Título da Pesquisa:** EFEITO CLÍNICO DA LASERTERAPIA NA HIPOSSALIVAÇÃO E XEROSTOMIA DECORRENTES DA RADIOTERAPIA EM REGIÃO DE CABEÇA E PESCOÇO

**Pesquisador:** Fernanda Gonçalves Salum

**Área Temática:**

**Versão:** 1

**CAAE:** 10513712.7.0000.5336

**Instituição Proponente:** Pontifícia Universidade Católica do Rio Grande do Sul - PUC/RS

##### DADOS DO PARECER

**Número do Parecer:** 154.280

**Data da Relatoria:** 23/11/2012

##### Apresentação do Projeto:

Projeto de mestrado vinculado a FO. Bem estruturado. Objetivos e metodologia bem explicitadas. Critérios de inclusão e exclusão bem definidos. Estudo clínico, prospectivo, randomizado, cego, placebo-controlado. No presente estudo é necessário que utilizar o placebo do laser, para que se verifique o real efeito da laserterapia sobre a hipossalivação e xerostomia. A laserterapia não é empregada como protocolo padrão ao tratamento da xerostomia e hipossalivação. Pode ser empregada de modo coadjuvante aos tratamentos convencionais ou de forma isolada. Caso, por meio desta pesquisa, haja confirmação da eficácia da laserterapia no tratamento da xerostomia e hipossalivação, os indivíduos que fizeram parte do grupo placebo serão chamados para nova avaliação e laserterapia.

##### Objetivo da Pesquisa:

**Objetivo Primário:**

Avaliar o efeito clínico da laserterapia sobre a xerostomia e hipossalivação de pacientes submetidos à radioterapia para tratamento de neoplasias de cabeça e pescoço.

**Objetivo Secundário:**

Avaliar em pacientes submetidos à radioterapia em região de cabeça e pescoço:-o efeito da LLLT na hipossalivação, por meio de avaliação da velocidade do fluxo salivar;-o efeito da LLLT na xerostomia, por meio de escala analógica visual;-se a laserterapia, utilizada para o tratamento da hipossalivação e xerostomia, exerce influência na qualidade de vida relacionada à saúde bucal.

**Avaliação dos Riscos e Benefícios:**

Riscos:

Não há riscos referentes ao uso do laser terapêutico de baixa potência desde que paciente e profissional utilizem proteção ocular, conforme descrito no projeto de pesquisa.

Benefícios:

Elevação do fluxo salivar, diminuição da sensação de boca seca e melhoria na qualidade de vida dos pacientes.

**Comentários e Considerações sobre a Pesquisa:**

Serão selecionados para participar deste estudo 60 pacientes de ambos os sexos, submetidos à radioterapia para tratamento de neoplasias malignas de orofaringe há pelo menos seis meses. Os pacientes serão distribuídos aleatoriamente em dois grupos: Grupo-laser: n=30 e Grupocontrole: n=30. O pacientes serão selecionados no Serviço de Radioterapia do Hospital São Lucas (SERP) da PUCRS. Neste Serviço o regime radioterápico é realizado com o aparelho Theratronix, modelo Phoenix, unidade de teleterapia rotacional por Cobalto 60 (fôtons), com energia de 1,25 MeV. Durante a anamnese serão registrados em ficha individual os dados de identificação do paciente, história médica, uso de medicamentos, antecedentes amiliares e hábitos de tabagismo e etilismo .

Será realizado o exame físico loco-regional e intrabucal. A xerostomia será avaliada por meio de escala visual analógica (EVA), desenvolvida por Pai et al. (2001) e utilizada por Jham (2006) para avaliação deste sintoma. Esta escala consiste de oito itens relacionados à xerostomia e quantificados por meio de linhas horizontais medindo 10 cm de extensão. Os valores estão compreendidos entre o zero (que significa ausência do sintoma) e dez

(sintomatologia máxima). Os pacientes serão orientados a marcar um traço vertical sobre a linha relacionada a cada um dos itens. Para a mensuração do fluxo salivar, serão obtidas amostras de saliva total. Para coleta da saliva total em repouso, o paciente deverá evitar a movimentação..incluindo,pequenos,movimentos,da língua,bochechas,mandíbula ou lábios. Os lábios devem ser mantidos ligeiramente abertos permitindo que a saliva escorre passivamente dentro de frasco de polipropileno graduado. O tempo de execução do processo deverá ser de 15 minutos. Para a coleta sob estimulação será empregado estímulo mecânico. Os pacientes serão orientados a mastigar um cilindro de látex medindo 1,0 cm de comprimento por 0,5 cm de diâmetro preso a um pedaço de fio dental para evitar deglutição. O paciente deverá mastigar a peça durante um minuto e depois remover toda a saliva por expectoração ou deglutição. A mastigação deverá prosseguir por um período de 5 minutos com o mesmo dispositivo de látex. A cada minuto o paciente deve expelir o conteúdo acumulado no frasco. A qualidade de vida dos pacientes será avaliada por meio do instrumento OHIP-14 (Oral Health Impact Profile), versão em português, traduzida e validada por Oliveira e Nadanovsky (2005). Os itens do OHIP-14 são agrupados em sete subescalas: limitação

funcional, dor física, desconforto psicológico, limitação física, limitação psicológica, limitação social e incapacidade. As aplicações do laser serão realizadas no ambulatório do Serviço de Estomatologia e Prevenção do Câncer Bucomaxilofacial do Hospital São Lucas-PUCRS. Os pacientes do grupo laser serão submetidos a duas aplicações semanais de radiação laser de diodo durante seis semanas, totalizando 12 sessões. As aplicações serão realizadas nas glândulas salivares maiores, de forma pontual e contínua. Serão aplicados

três pontos sobre cada glândula parótida, dois pontos sobre cada submandibular e dois pontos sobre cada sublingual na dosimetria de 199 J/cm<sup>2</sup> totalizando energia de 6 J por ponto. No grupo controle os procedimentos serão os mesmos, no entanto com o aparelho desligado, onde um timer emitirá um sinal sonoro mimetizando a aplicação do grupo laser. A escala visual analógica, o instrumento OHIP-14 e a coleta de saliva serão realizadas nos momentos baseline, sexta sessão e décima segunda sessão.

**Considerações sobre os Termos de apresentação obrigatória:**

- TCLE , ok;
- folha de rosto, ok;
- Autorização para uso das dependências do serviço de estomatologia e prevenção do câncer bucomaxilofacial, ok;
- Carta da CCFO, ok;
- Termo de confidencialidade para uso de dados, ok;
- Autorização para acessar dados dos prontuários de radioterapia, ok;
- Lattes, ok;
- Orçamento, ok.

**Recomendações:**

Não há recomendações a fazer.

**Conclusões ou Pendências e Lista de Inadequações:**

Não há pendências e inadequações no presente protocolo.

**Situação do Parecer:**

Aprovado

**Necessita Apreciação da CONEP:**

Não

**Considerações Finais a critério do CEP:**

Parecer de acordo com a reunião do colegiado.

PORTO ALEGRE, 26 de Novembro de 2012

---

**Assinador por:**  
**Rodolfo Heriberto Schneider**  
**(Coordenador)**

**ANEXO C****Escala visual analógica**

1. Marque a dificuldade que você tem para falar devido à secura

Não é difícil 0-----10 Muito difícil

2. Marque a dificuldade que você tem para engolir devido à secura

Não é difícil 0-----10 Muito difícil

3. Marque a quantidade de saliva na sua boca

Muita saliva 0-----10 Nenhuma saliva

4. Marque a secura da sua boca

Não é seca 0-----10 Muito seca

5. Marque a secura da sua garganta

Não é seca 0-----10 Muito seca

6. Marque a secura dos seus lábios

Não são secos 0-----10 Muito secos

7. Marque a secura da sua língua

Não é seca 0 -----10 Muito seca

8. Marque o grau da sua sede

Não sinto sede 0-----10 Sinto muita sede

## ANEXO D

### OHIP-14 (ORAL HEALTH IMPACT PROFILE)

*REPRODUÇÃO DO "PERFIL DE IMPACTO NA SAÚDE ORAL" (OHIP14)<sup>26</sup>*

Nos últimos seis meses, por causa de problemas com seus dentes ou sua boca:	Nunca	Raramente	Às vezes	Repetidamente	Sempre
1. Você teve problemas para falar alguma palavra?					
2. Você sentiu que o sabor dos alimentos tem piorado?					
3. Você sentiu dores em sua boca ou nos seus dentes?					
4. Você se sentiu incomodado(a) ao comer algum alimento?					
5. Você ficou preocupado(a)?					
6. Você se sentiu estressado(a)?					
7. Sua alimentação ficou prejudicada?					
8. Você teve que parar suas refeições?					
9. Você encontrou dificuldade para relaxar?					
10. Você se sentiu envergonhado(a)?					
11. Você ficou irritado(a) com outras pessoas?					
12. Você teve dificuldade para realizar suas atividades diárias?					
13. Você sentiu que a vida, em geral, ficou pior?					
14. Você ficou totalmente incapaz de fazer suas atividades diárias?					

## ANEXO E

### SUMISSÃO DO ARTIGO DE REVISÃO DA LITERATURA NO PERIÓDICO *ARCHIVES OF ORAL BIOLOGY*

#### **Submission Confirmation for SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS**

ees.aob.0.27aea2.5d179d91@eesmail.elsevier.com em nome de Archives of Oral Biology [AOB@elsevier.com]

Enviado quarta-feira, 5 de março de 2014 10:14

Para: Fernanda Gonçalves Salum; fernanda\_salum@hotmail.com

Archives of Oral Biology

Title: SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS

Authors: jamil saleh; Maria A Figueiredo, PhD; Karen Cherubini, PhD; Fernanda G Salum, PhD

Article Type: Review Article

Dear Fernanda,

Your submission entitled "SALIVARY HYPOFUNCTION: AN UPDATE ON ETIOLOGY, DIAGNOSIS AND THERAPEUTICS" has been received by Archives of Oral Biology.

You may check on the progress of your paper by logging on to the Elsevier Editorial System as an author. The URL is <http://ees.elsevier.com/aob/>.

Your manuscript will be given a reference number once an Editor has been assigned.

Thank you for submitting your work to this journal. Please do not hesitate to contact me if you have any queries.

Kind regards,

(On behalf of the Editors)

Archives of Oral Biology

\*\*\*\*\*  
For any technical queries about using EES, please contact Elsevier Author Support at [authorsupport@elsevier.com](mailto:authorsupport@elsevier.com)

## ANEXO F

### SUMISSÃO DO ARTIGO DE PESQUISA NO PERIÓDICO *PHOTOMEDICINE AND LASER SURGERY*

#### **Photomedicine and Laser Surgery - Manuscript ID PHO-2014-3741**

onbehalfof+photomedicine.editorial+gmail.com@manuscriptcentral.com em nome de photomedicine.editorial@gmail.com  
Enviado segunda-feira, 17 de março de 2014 14:18  
Para: Fernanda Goncalves Salum

17-Mar-2014

Dear Dr. Salum:

Your manuscript entitled "EFFECT OF LOW-LEVEL LASER THERAPY ON RADIOTHERAPY-INDUCED HYPOSALIVATION AND XEROSTOMIA: A PILOT STUDY" has been successfully submitted online and is presently being given full consideration for publication in Photomedicine and Laser Surgery.

However, we would like to inform you that if your manuscript, which includes text, abstract, references and tables or figures, is not formatted according to the author instructions, we will not be able to process your submission. We will notify you of the changes to be made and unsubmit your paper, enabling you to implement the formatting corrections and re-submit once they are complete.

To help defray the publication costs as we increase the number of articles we publish in each issue, for manuscripts submitted after January 1, 2010 the Journal is implementing page charges of \$60 per printed page. Please note that payment of page charges can be waived under certain circumstances and is not a prerequisite for publication.

Your manuscript ID is PHO-2014-3741.

Please mention the above manuscript ID in all future correspondence or when calling the office for questions. If there are any changes in your street address or e-mail address, please log in to Manuscript Central at <http://mc.manuscriptcentral.com/photomedicine> and edit your user information as appropriate.

You can also view the status of your manuscript at any time by checking your Author Center after logging in to <http://mc.manuscriptcentral.com/photomedicine>.

Thank you for submitting your manuscript to Photomedicine and Laser Surgery.

Sincerely,  
Photomedicine and Laser Surgery Editorial Office

Register to receive email notifications from the Journal(s) that are critical to advancing your work: [www.liebertpub.com/liebertconnect](http://www.liebertpub.com/liebertconnect)