

# Machine Learning-Based Scoring Functions, Development and Applications with SAnDReS



Gabriela Bitencourt-Ferreira<sup>1,#</sup>, Camila Rizzotto<sup>1,#</sup> and Walter Filgueira de Azevedo Junior<sup>1,2,\*</sup>

<sup>1</sup>Pontifical Catholic University of Rio Grande do Sul - PUCRS, Porto Alegre-RS, Brazil; <sup>2</sup>Specialization Program in Bioinformatics. Pontifical Catholic University of Rio Grande do Sul-(PUCRS). Av. Ipiranga, 6681 Porto Alegre/RS 90619-900, Brazil

**Abstract: Background:** Analysis of atomic coordinates of protein-ligand complexes can provide three-dimensional data to generate computational models to evaluate binding affinity and thermodynamic state functions. Application of machine learning techniques can create models to assess protein-ligand potential energy and binding affinity. These methods show superior predictive performance when compared with classical scoring functions available in docking programs.

**Objective:** Our purpose here is to review the development and application of the program SAnDReS. We describe the creation of machine learning models to assess the binding affinity of protein-ligand complexes.

**Methods:** SAnDReS implements machine learning methods available in the scikit-learn library. This program is available for download at <https://github.com/azevedolab/sandres>. SAnDReS uses crystallographic structures, binding and thermodynamic data to create targeted scoring functions.

**Results:** Recent applications of the program SAnDReS to drug targets such as Coagulation factor Xa, cyclin-dependent kinases and HIV-1 protease were able to create targeted scoring functions to predict inhibition of these proteins. These targeted models outperform classical scoring functions.

**Conclusion:** Here, we reviewed the development of machine learning scoring functions to predict binding affinity through the application of the program SAnDReS. Our studies show the superior predictive performance of the SAnDReS-developed models when compared with classical scoring functions available in the programs such as AutoDock4, Molegro Virtual Docker and AutoDock Vina.

**Keywords:** Machine learning, SAnDReS, cyclin-dependent kinase, protein-ligand interactions, binding affinity, Gibbs free energy.

## 1. INTRODUCTION

Evaluation of protein-ligand interactions based on the atomic coordinates of a binary complex is of fundamental importance to establish the structural basis for the specificity of binders for a receptor [1]. Analy-

sis of protein-ligand interactions identified in complex structures may reveal the critical determinants for binding specificity, which may contribute to drug design and development. Moreover, the availability of structures makes it possible to assess the binding affinity computationally [2-4]. We may evaluate the binding affinity or thermodynamic parameters through quantum mechanics methods [5]. Another methodology to determine this information is the classical molecular dynamics simulation [6].

Quantum mechanics and molecular dynamics simulations have the potential to generate computational

\*Address correspondence to this author at Pontifical Catholic University of Rio Grande do Sul - PUCRS, Porto Alegre-RS, Brazil; Specialization Program in Bioinformatics. Pontifical Catholic University of Rio Grande do Sul (PUCRS). Av. Ipiranga, 6681 Porto Alegre/RS 90619-900 Brazil; Tel/Fax: +55-51-3353-4529; E-mail: [walter@azevedolab.net](mailto:walter@azevedolab.net).

<sup>#</sup>These authors contributed equally to this work and should be regarded as co-first authors.

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models to predict binding affinity. But they present high computational cost when compared with classical scoring functions, being the quantum mechanics approach the most expensive from the computational point of view [5].

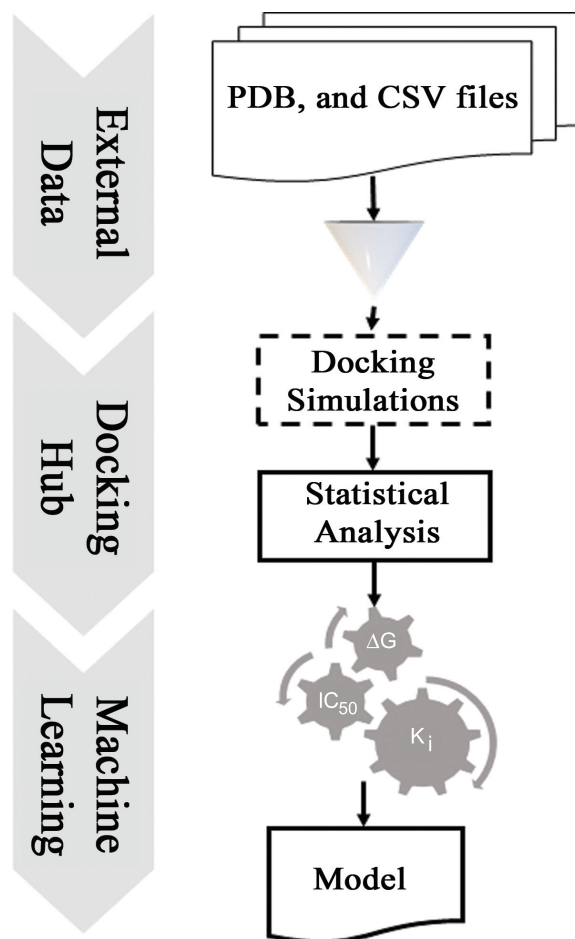
Application of force field and scoring function methods [7] can successfully predict binding affinity for protein-ligand structures. Considering the recent developments in the use of machine learning techniques to generate scoring functions, we may say that they have shown considerable improvement in the predictive performance of these methods [8-16].

Among the recent proposed computational tools to assess binding affinity or thermodynamic data from the atomic coordinates of receptor-ligand complexes, we may highlight the following computational tools: property-encoded shape distributions together with standard support vector machine (PESD-SVM) [17], Random Forest Score (RF-Score series) [18-22], Neural-Network-Based Scoring function (NNScore series) [23-25], Pafnucy [26], Tool to Analyze the Binding Affinity (TABA) [27, 28] and SAnDReS [29, 30]. Our focus here is on the application of the program SAnDReS to estimate protein-ligand binding affinity. SAnDReS is an acronym for Statistical Analysis of Docking Results and Scoring Functions. Several studies reported the successful application of SAnDReS to a wide range of protein systems with different types of binding affinity and thermodynamic data [29-65]. These recent publications highlight the potential of SAnDReS to develop targeted-scoring functions for different protein systems and the superior predictive performance of the targeted-scoring functions developed using SAnDReS when compared against classical scoring functions available in docking programs. Here we give an overview of the methodology employed by SAnDReS with an emphasis on the machine learning techniques used to create targeted-scoring functions. We also discuss the successful application of SAnDReS to create polynomial equations to calculate binding affinity for four protein targets.

## 2. METHODS

SAnDReS integrates different methodologies to carry out docking simulations and for the creation of machine learning models to assess binding affinity. SAnDReS analyzes data from any protein-ligand docking program; the only requisite is to have structures in Protein Data Bank (PDB) [66-68] format, ligands in Structure Data Format (SDF), docking and scoring function data in comma-separated values (CSV) for-

mat. In Fig. (1), we have the main steps used in the SAnDReS. In the first step, we download the protein system composed of PDB and CSV files. In the following, SAnDReS filters the dataset. The filtered data is submitted to docking simulations. In the next step, SAnDReS performs docking; this phase is named docking hub. The docking results are subjected to statistical analysis to evaluate the docking performance of different protocols. Subsequently, SAnDReS generates scoring functions targeted to the protein system of interest. SAnDReS can carry out this last step independently of the other phases.



**Fig. (1).** SAnDReS Schematic Flowchart. SAnDReS downloads structure and CSV files from the PDB and filters and merges them. In the following, SAnDReS carries out docking simulations and analyzes their results. Next, SAnDReS uses the ensemble of structures and binding data to generate machine learning models. The dashed rectangle indicates the non-mandatory step for the generation of targeted-scoring functions. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

In the development of machine learning models, SAnDReS considers as explanatory variables the energy terms and scoring functions calculated by external

programs such as Molegro Virtual Docker (MVD) [69-71], AutoDock4 (AD4) [72, 73] and AutoDock Vina [74]. These explanatory variables will have their relative weights determined by supervised machine learning techniques such as linear regression, least absolute shrinkage and selection operator (Lasso) [75], ridge [76] and elastic net [77]. These last three techniques allow the application of cross-validation methods [29, 30].

The machine learning step of SAnDReS makes use of seven regression classes implemented in Python and accessible in the scikit-learn library [78]. We have one class for each method as follows: Ordinary Linear Regression (*sklearn.linear\_model.LinearRegression*), Lasso (*sklearn.linear\_model.Lasso*), Lasso with cross-validation (*sklearn.linear\_model.LassoCV*), Ridge (*sklearn.linear\_model.Ridge*), Ridge with cross-validation (*sklearn.linear\_model.RidgeCV*), Elastic Net (*sklearn.linear\_model.ElasticNet*) and Elastic Net with cross-validation (*sklearn.linear\_model.ElasticNetCV*).

SAnDReS makes use of a polynomial equation [79-82] to evaluate the binding affinity or Gibbs free energy of binding for protein-ligand complexes. Let us consider that we estimated the energy of the protein-ligand interactions through three terms named here  $x_1$ ,  $x_2$  and  $x_3$ . These terms are the explanatory variables of the equation below,

$$y = \lambda_0 + \lambda_1 x_1 + \lambda_2 x_2 + \lambda_3 x_3 + \lambda_4 x_1 \cdot x_2 + \lambda_5 x_1 \cdot x_3 + \lambda_6 x_2 \cdot x_3 + \lambda_7 x_1^2 + \lambda_8 x_2^2 + \lambda_9 x_3^2$$

where  $\lambda_0$  is the regression constant and  $\lambda$ 's are the relative weights of each explanatory variable. The response variable is  $y$ , which could be the Gibbs free energy of binding ( $\Delta G$ ) or the logarithm of the binding affinity (*i.e.*,  $\log(K_i)$ ). We take binding affinity and thermodynamic information from three databases: BindingDB [83, 84], MOAD [42, 85, 86] and PDBbind [87]. Considering that we have mixed and squared energy terms, we ended up with nine weights related to these explanatory variables. We have a total of 511 possible polynomial equations [29, 30].

SAnDReS generates predictive models, also testing different supervised machine learning techniques. Taking together, we verify a total of 3577 models for each dataset. We consider, as a dataset, a protein system formed with crystallographic structures of protein-ligand complexes with experimental data for binding affinity or Gibbs free energy of binding. SAnDReS assesses the predictive performance of the machine learning models and classical scoring function through the evaluation of Spearman's rank and Pearson correlation coefficients [88].

### 3. RESULTS AND DISCUSSION

As we previously highlighted, there are studies with SAnDReS applied to a variety of protein systems. Table 1 summarizes recently published protein systems related to the development of machine learning models to predict binding affinity for a specific protein system. Table 2 shows the predictive performance of SAnDReS polynomial scoring functions and classical scoring functions [29, 30, 35-37]. All these studies bring predictive performance comparisons of classical scoring functions against the targeted-scoring functions generated with SAnDReS for systems involving specific protein families and based on crystallographic structural data and experimental binding affinity information.

Analysis of Table 1 indicates that SAnDReS can generate machine learning models taking energy terms calculated with different classical scoring functions, such as the ones calculated with MVD and AD4. We also see from the protein systems for which SAnDReS was tested so far, that its performance is not restricted to a specific enzymatic class or type of binding affinity. We have models for CDK [36], HIV-1 protease [35], 3-dehydroquinase dehydratase [37] and coagulation factor Xa [29]. SAnDReS analyzed protein systems with experimental data such as  $K_i$  [29, 35, 37],  $IC_{50}$  [36] and  $\Delta G$  [15]. SAnDReS can handle any binding affinity data or thermodynamic parameters in the development of machine learning models.

It is worth mentioning that the majority of the polynomial equations built with SAnDReS are for a specific protein, with one exception of a predictive model made to estimate the Gibbs free energy of binding trained with a dataset composed with 48 high-resolution crystallographic structures [15]. This dataset took different types of enzymes and protein classes, expecting to generate an all-purpose predictive model to evaluate  $\Delta G$  based on the atomic coordinates of protein-ligand complexes. We did not include an analysis of the predictive performance of this  $\Delta G$  dataset in this review since it takes a wide range of enzymatic classes.

Analysis of the predictive performance of a machine learning model compared with classical scoring functions strongly indicate that the former shows a higher correlation with experimental data. Spearman's rank correlation coefficients ( $\rho$ ) for test sets (Table 2) indicated values ranging from 0.08 (HIV-1 protease) [35] to 0.771 (3-dehydroquinase dehydratase) [37] for classical scoring functions. On the other hand, taking these two systems, machine learning models overperformed classical scores, with  $\rho$  of 0.368 and 0.943, respec-

**Table 1. Protein systems studied with SAnDReS.**

S.No.	Protein System	Size	Machine Learning Model
1	Coagulation factor Xa (EC 3.4.21.6) with $K_i$ data [29]	57 <sup>a</sup> 25 <sup>b</sup>	Score <sub>110</sub> <sup>c</sup> =1.603905-0.006305.x -0.005256.y.z-0.000028.x.z +0.002801.y <sup>2</sup> +0.002439.z <sup>2</sup>
2	HIV-1 protease (EC 3.4.23.16) with $K_i$ data [35]	51 <sup>a</sup> 20 <sup>b</sup>	Score <sub>504</sub> <sup>d</sup> =-5.685144+0.01199.x +0.004743.y+0.001676.z -0.000024.x.y+0.000106.x.z -0.00004.y.z
3	Cyclin-dependent kinase (EC 2.7.11.22) with $IC_{50}$ data [36]	122 <sup>a</sup> 54 <sup>b</sup>	Score <sub>482</sub> <sup>e</sup> =-7.074331-0.001829.x +0.001529.y-0.001136.z +0.000003.x.y
4	3-dehydroquinate dehydratase (EC 4.2.1.10) with $K_i$ data [37]	18 <sup>a</sup> 4 <sup>b</sup>	Score <sub>369</sub> <sup>f</sup> =-7.268556-0.545897.y -1.288947.z-0.019562.x -0.396378.y.z+0.438998.z <sup>2</sup>

<sup>a</sup>Training set, <sup>b</sup>Test set. <sup>c</sup>Calculated with MVD, where x=Electro Score, y=MolDock Score and z=Interaction Score. <sup>d</sup>Calculated with MVD, where x=PLANTS, y=Interaction Score and z=Ligand efficiency 3 Score.

<sup>e</sup>Calculated with MVD, where x=Re-rank Score, y=Internal Score and z=Electro Long Score. <sup>f</sup>Calculated with AD4, where x=vdW+Hbond+desolv Energy, y=Electrostatic Energy and z=Final intermolecular Energy.

**Table 2. Predictive performance of machine learning (M) models generated with SAnDReS.**

S.No.	$\rho^a$	p-value <sup>a</sup>	$\rho^b$	p-value <sup>b</sup>
1	0.560 (M) 0.190 (C)	5.920.10 <sup>-6</sup> (M) 1.574.10 <sup>-1</sup> (C)	0.435 (M) 0.174 (C)	2.975.10 <sup>-2</sup> (M) 4.055.10 <sup>-1</sup> (C)
2	0.525 (M) 0.479 (C)	7.707.10 <sup>-5</sup> (M) 3.795.10 <sup>-4</sup> (C)	0.368 (M) 0.080 (C)	1.106.10 <sup>-1</sup> (M) 7.383.10 <sup>-1</sup> (C)
3	0.390 (M) 0.211 (C)	9.065.10 <sup>-6</sup> (M) 1.943.10 <sup>-2</sup> (C)	0.346 (M) -0.298 (C)	1.044.10 <sup>-2</sup> (M) 2.874.10 <sup>-2</sup> (C)
4	0.675 (M) 0.427 (C)	4.16.10 <sup>-3</sup> (M) 9.90.10 <sup>-2</sup> (C)	0.943 (M) 0.771 (C)	4.81.10 <sup>-3</sup> (M) 7.24.10 <sup>-2</sup> (C)

<sup>a</sup>Training set, <sup>b</sup>Test set

C: The highest-correlation classical scoring function (calculated with MVD). M: The highest-correlation machine learning model.

tively. The machine learning model generated to predict inhibition constant of 3-dehydroquinate dehydratase showed the highest  $\rho$  amongst the models produced with SAnDReS so far. This performance seems to be directly related to the higher correlation of the energy terms from the classical scoring functions used to create the targeted scoring function model, which also shows a significant correlation for the same test set.

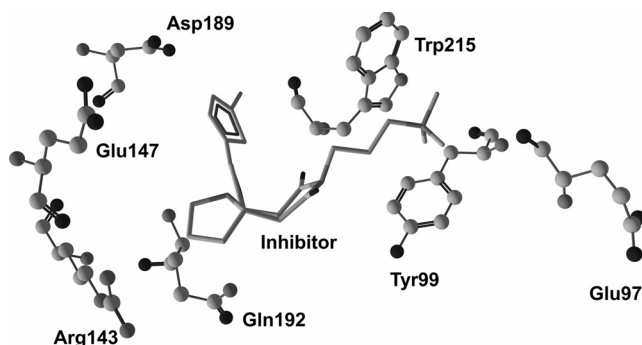
Furthermore, the use of cross and square terms of variables in the polynomial equation used by SAnDReS confers additional flexibility during the machine-learning modeling allowing the model to adapt the scoring function to the protein system of interest. New explanatory variables, including cross and square variables let us explore additional regions of the scoring function space [31], not explored with a linear poly-

mial equation. Such a deeper polynomial equation showed superior predictive performance when compared with classical scoring functions.

Another noteworthy result related to the machine learning models created with SAnDReS is the poor performance of the predictive model built to estimate  $IC_{50}$  of CDKs [36]. We consider two possible causes for this weak predictive power of the machine learning model. Firstly, so far, this is the largest dataset used to generate targeted scoring functions with SAnDReS, over 170 crystallographic structures. Secondly, the source of the binding affinity information ( $IC_{50}$ ) generally is not as consistent as inhibition constant ( $K_i$ ) or dissociation constant ( $K_d$ ) data [26]. Therefore, we expect datasets with  $IC_{50}$  to present mediocre predictive performance when compared with models created to estimate other types of binding affinity.

To overcome the limitation of  $IC_{50}$  predictive performance, we could have applied the Cheng–Prusoff [89] equation or similar approach to convert  $IC_{50}$  to  $K_i$ . Such an approach can generate a converted- $K_i$  dataset. We did not follow this approach, since we see as adequate to focus on modeling direct experimental data, not derived experimental information such as the one obtained using the Cheng–Prusoff equation.

Analysis of the machine learning models to predict inhibition of the protein systems built so far with SAnDReS clearly showed that the regression equation could capture essential structural features related to protein-ligand interactions specific for the system under study. For the coagulation factor Xa [29], the machine learning model indicated the prevalence of electrostatic interactions in the polynomial equation developed with SAnDReS. In Fig. (2), we have the residues involved in most of the electrostatic interactions for the crystallographic structures in the coagulation factor Xa dataset.

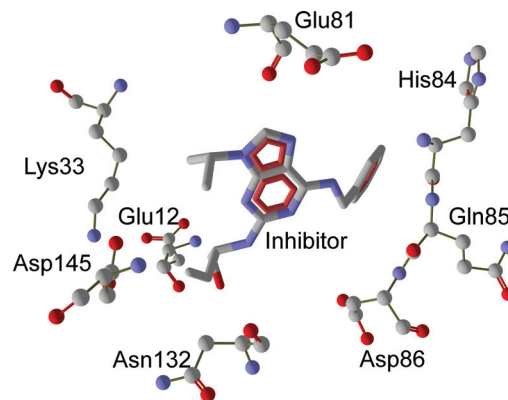


**Fig. (2).** Main residues involved in intermolecular electrostatic interactions for factor Xa. We used the program MVD [69] and the crystallographic structure of coagulation factor Xa in complex with an inhibitor (PDB access code: 2JKH) [90] to generate this figure. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

Analysis of the structures in the dataset suggests that the prevalence of electrostatic intermolecular interactions in the polynomial equation may be due to the presence of charged residues in the binding pocket (Arg 143, Gln 192 and Asp 189) of the coagulation factor Xa. Moreover, analysis of strong coagulation factor Xa inhibitors shows that amine moieties fill the protein binding pocket, which is involved in cation- $\pi$  interactions with residues Tyr 99, Phe 174 and Trp 215.

The development of a machine learning model built to predict inhibition of CDKs [36] also showed the prevalence of electrostatic interactions in the polynomial equation (Table 1), as we can see for the presence

of the Electro Long Score in the score<sub>482</sub>. In Fig. (3), we have the residues participating in most of the intermolecular contacts for the crystallographic structures in the CDK dataset. We see the preponderance of the electrostatic intermolecular interactions with the participation of charged residues Glu 12, Lys 33, Glu 81, His 84, Gln 85, Asp 86, Asn 132 and Asp 145 [91-96].



**Fig. (3).** Intermolecular interactions in the ATP-binding pocket of CDK2. We used the program MVD [69] and the crystallographic structure of CDK2 in complex with the inhibitor roscovitine (PDB access code: 2A4L) [90] to generate this figure. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

Analysis of the machine learning model to predict binding affinity for 3-dehydroquinone dehydratase [37] indicates the participation of electrostatic interactions, as observed for factor Xa and CDK2. Nevertheless, we also identified energy terms involving intermolecular hydrogen bond and van der Waals interactions in the machine learning model, which indicates that the SAnDReS method can identify a wide range of intermolecular interactions.

Molecular docking is the most used computational approach in the early stages of drug design [71-74] and the application of machine-learning techniques has shown to positively contribute to speed up drug discovery [81]. Considering the prospect for drug design and development, the results obtained so far using the program SAnDReS strongly indicate that we have a synergism between the computational environment applied to carry out docking simulations and the machine-learning methods used to generate targeted-scoring functions to predict protein-ligand binding affinity based on the atomic coordinates of receptor-drug complexes. Both main computational approaches available in the program SAnDReS have the potential to speed up drug discovery and design. Firstly, by a user-friendly computational environment, mostly based on open-source software, able to carry out docking simula-

tions that immensely contribute to the identification of potential new inhibitors of druggable protein targets. Secondly, by a computational integrated environment that uses state-of-the-art machine learning approaches that fully integrate all tasks related to the development of targeted-scoring functions, from the automatic downloading of structural and binding experimental data to the generation of novel scoring functions to predict binding affinity taking the atomic coordinates of target-drug complexes.

In summary, in the early stages of drug design and development, the availability of the atomic coordinates of an enzyme target makes it possible to apply the program SAnDReS to identify new potential inhibitors for a druggable target with predictive power superior to classical methods. Furthermore, the innovative theoretical approach of using SAnDReS to explore the scoring function space [31] to find an adequate computational model to predict the binding affinity or thermodynamic data brings an integrated computational environment that not only is able to perform docking and machine-learning modeling but also to provide a theoretical framework that makes it possible to medicinal chemists to explore key structural features responsible for the binding affinity of drugs. These structure features are unique to the protein system being studied since SAnDReS can generate a targeted-scoring specific for this druggable protein target. As highlighted for the electrostatic interactions found in the machine learning models for coagulation factor Xa [29] and CDK [36], identification of the most relevant intermolecular interactions responsible for binding affinity provides the information of chemical environment that allows us to refine structural parameters of a potential drug making it more specific for a given protein target. Medicinal chemists can carry out the modification of the potential drug to maximize the major intermolecular interactions identified through machine-learning approaches available in the program SAnDReS.

## CONCLUSION

The main idea behind the development of the SAnDReS is to have a computational tool to explore the scoring function space through fine-tuning energy terms generated by other programs and calibrating a scoring function to a protein system of interest. We focus on experimental data for the structure and binding information, to generate machine learning models based strictly on experimental information. Moreover, with SAnDReS, we can carry out protein-ligand docking simulations in an integrated computational environment

with SAnDReS. This program focuses on the execution of docking simulations of protein-ligand systems using the programs MVD, AD4 and Vina. SAnDReS analyzes the performance of molecular docking simulations and generates machine learning models built to predict binding affinity using as explanatory variables the energy terms available in scoring functions of any docking programs. In this review, we described the SAnDReS application to generate machine learning models to calculate binding affinity. SAnDReS seeks to create a model considering a dataset of crystallographic structures for which binding affinity or thermodynamic data is available. With this approach, SAnDReS is adequate for protein systems with at least 20 crystallographic structures. So far, the models generated with SAnDReS to predict binding affinity showed superior predictive performance when compared with classical scoring functions. Furthermore, SAnDReS was able to capture in the machine learning models, essential structural features responsible for binding affinities, such as the electrostatic interactions in the polynomial equations for the Coagulation factor Xa and CDKs.

## LIST OF ABBREVIATIONS

AD4	= AutoDock4
ATP	= Adenosine triphosphate
CDK	= Cyclin-dependent kinase
CSV	= Comma-separated values
CV	= Cross validation
DG	= Variation of Gibbs free energy of binding
EC	= Enzyme classification number
IC <sub>50</sub>	= Half-maximal inhibitory concentration
K <sub>d</sub>	= Dissociation constant
K <sub>i</sub>	= Inhibition constant
Lasso	= Least absolute shrinkage and selection operator
MOAD	= Mother of all databases
MVD	= Molegro virtual docker
NNScore	= Neural-network-based scoring function
PDB	= Protein data bank
PESD-SVM	= Property-encoded shape distributions together with standard support vector machine
RF-Score	= Random forest score



SAnDReS = Statistical analysis of docking results and scoring functions  
 SDF = Structure data format  
 TABA = Tool to analyze the binding affinity

### CONSENT FOR PUBLICATION

Not applicable.

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### CONFLICT OF INTEREST

Walter Filgueira de Azevedo Junior is an acting Section Editor for the journal *Current Medicinal Chemistry*.

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