

ESCOLA DE CIÊNCIAS  
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOLOGIA CELULAR E MOLECULAR  
DOUTORADO EM BIOLOGIA CELULAR E MOLECULAR

ANA PAULA AQUISTAPASE DAGNINO

**CARACTERIZAÇÃO DO SISTEMA NOCICEPTINA/ORFANINA FQ-RECEPTOR NOP NA  
MODULAÇÃO DA FIBROMIALGIA EXPERIMENTAL**

Porto Alegre

2019

PÓS-GRADUAÇÃO - *STRICTO SENSU*



Pontifícia Universidade Católica  
do Rio Grande do Sul



Pontifícia Universidade Católica do Rio Grande do Sul  
Escola de Ciências  
Programa de Pós-Graduação em Biologia Celular e Molecular

**ANA PAULA AQUISTAPASE DAGNINO**

**CARACTERIZAÇÃO DO SISTEMA NOCICEPTINA/ORFANINA FQ-  
RECEPTOR NOP NA MODULAÇÃO DA FIBROMIALGIA  
EXPERIMENTAL**

Porto Alegre  
Janeiro de 2019

## **Ficha Catalográfica**

D126c Dagnino, Ana Paula Aquistapase

Caracterização do sistema nociceptina/orfanina FQ receptor NOP na modulação da fibromialgia experimental / Ana Paula Aquistapase Dagnino . – 2019.

148.

Tese (Doutorado) – Programa de Pós-Graduação em Biologia Celular e Molecular, PUCRS.

Orientadora: Profa. Dra. Maria Martha Campos.

1. Fibromialgia. 2. Reserpina. 3. Nociceptina/orfanina FQ. 4. Receptor da nociceptina/orfanina FQ. 5. UFP-101. I. Campos, Maria Martha. II. Título.

Elaborada pelo Sistema de Geração Automática de Ficha Catalográfica da PUCRS  
com os dados fornecidos pelo(a) autor(a).

Bibliotecária responsável: Salete Maria Sartori CRB-10/1363

**ANA PAULA AQUISTAPASE DAGNINO**

**CARACTERIZAÇÃO DO SISTEMA NOCICEPTINA/ORFANINA FQ-  
RECEPTOR NOP NA MODULAÇÃO DA FIBROMIALGIA  
EXPERIMENTAL**

Tese apresentada como requisito parcial para obtenção do grau de Doutor pelo Programa de Pós-Graduação em Biologia Celular e Molecular, Escola de Ciências, Pontifícia Universidade Católica do Rio Grande do Sul.

Orientadora: Prof.a. Dra. Maria Martha Campos

Porto Alegre

Janeiro de 2019

**ANA PAULA AQUISTAPASE DAGNINO**

**CARACTERIZAÇÃO DO SISTEMA NOCICEPTINA/ORFANINA FQ-  
RECEPTOR NOP NA MODULAÇÃO DA FIBROMIALGIA  
EXPERIMENTAL**

Tese apresentada como requisito parcial para obtenção do grau de Doutor pelo Programa de Pós-Graduação em Biologia Celular e Molecular da Pontifícia Universidade Católica do Rio Grande do Sul.

Orientadora: Prof.a. Dra. Maria Martha Campos

**BANCA EXAMINADORA:**

---

Dra. Elaine Cristina Gavioli – UFRN

---

Dra. Elke Bromberg – PUCRS

---

Dr. Pedro Roosevelt Torres Romão – UFCSPA

---

Dra. Mônica Ryff Moreira Roca Vianna (Suplente) – PUCRS

Porto Alegre  
2019

Dedico este trabalho as pessoas mais importantes na minha vida:

Minha mãe, dentre todas as mulheres, a mais guerreira.

Meu pai, dentre todos os homens, o mais generoso.

Meu amor, Luis, o mais amoroso e gentil que conheço.

*O convívio é uma benção. Amo demais vocês!!!*

## **AGRADECIMENTOS**

Aos meus pais, Rodolfo e Graciela Dagnino. Sinto muita gratidão de tê-los como pais, sempre tão amorosos e presentes em minha vida. Muitas vezes estive distante devido ao doutorado, mas sempre com vocês dentro do meu coração. Obrigada pela minha vida.

Ao meu amado Luis Fernando Kanan. Como é possível olhar lá no alto e ter os pés no chão? Nos momentos de risada que escondiam desespero. Homem inteligente, dedicado, bondoso e educado, um *gentleman*. Um suspiro de alento em minha insanidade. Minha alegria e meu bem querer.

À minha orientadora, Maria Martha Campos, muitas vezes amiga, muitas vezes mãe. O que melhor descreve esta mãe científica é o cuidado, o carinho, o capricho e a generosidade com seus alunos. Cientista íntegra, correta, de caráter. Totalmente dedicada à ciência, que não somente é um trabalho, mas uma vida. Bom Prof. Martha, acho que tu és a cereja do bolo do nosso laboratório. Uma vez um colega me desejou “Boa sorte”. E hoje vejo, sorte esta, que tive quando cheguei neste laboratório e aprendi tudo que aprendi com você.

Ao meu amigo e colega Pedro Chagastelles, que traz leveza para o dia-dia do laboratório, que muitas vezes é exaustivo. Cientista astuto, com humor a flor da pele. Descontraído, sem mesmices, criativo. As melhores pessoas na minha vida chegaram e se foram com muita facilidade. Espero encontrar esta alma em outras vidas, se possível ainda nesta, na Itália...

Ao meu amigo e colega Rodrigo Braccini, defensor de suas crenças e ideologias. Me acompanhou desde o começo da vida de doutorado, quando não sabia nem como pegar um camundongo. Organizado, regrado, entusiasta e cientista obstinado.

As minhas queridas amigas e colegas de laboratório, Priscilla Pail, Raquel Dal Sasso e Renata Medeiros, pela colaboração em experimentos, pelas risadas e por compartilhar dos mesmos sentimentos inerentes à busca pela Pós-graduação, tão almejada. Em nenhum momento esta caminhada foi individual.

Aos professores e colaboradores deste projeto, Dr. Samuel Greggio, Dra. Gianina Teribele, Dr. Jaderson Costa da Costa, Dr. Maurício Bogo e Dra. Talita Pereira, sem os quais este trabalho não teria sido tão completo e elegante. Sem dúvida a contribuição de vocês foi imensa para que este trabalho fosse promissor.

Aos técnicos de laboratório Rafaela Rubim, Juliano Soares, Moema Queiroz Vieira e Janaína Pasetti Nunes, extremamente solícitos e empenhados em auxiliar no que fosse preciso no laboratório, sempre nos passando seus conhecimentos nas técnicas utilizadas.

À equipe do CeMBE, em especial as queridas Priscila Alves da Silveira e Andressa Vargas Ribeiro, pelo bom humor e profissionalismo, sempre muito criteriosas no manejo com os animais.

“Às vezes todos os nossos pensamentos são inquietantes”

*Led Zeppelin*

## **RESUMO**

A fibromialgia é caracterizada por dor generalizada, sendo acompanhada por distúrbios funcionais e afetivos. Este estudo avaliou a implicação do receptor do peptídeo nociceptina/orfanina FQ (NOPr) em um modelo murino de fibromialgia. Os protocolos experimentais foram aprovados pela Comissão de Ética no Uso de Animais (CEUA/PUCRS 15/00487). A fibromialgia foi induzida em camundongos fêmeas CF-1 (20-24 g, 4 semanas de idade) pela administração de reserpina (0,25 mg/kg; via subcutânea), uma vez ao dia, durante 3 dias consecutivos. Grupos controle receberam veículo. No quarto dia, os camundongos foram tratados com uma dose única de nociceptina (N/OFQ) ou, do antagonista peptídico seletivo, UFP-101, administrados por via intraperitoneal (i.p., 0,3-5 nmol/kg), intracerebroventricular (i.c.v., 0,3 -1 nmol/sítio) ou intratecal (i.t., 0,3-5 nmol/sítio), 30 min antes das sessões experimentais. Em outra série de experimentos, os animais foram tratados com o antagonista UFP-101 (1 nmol/kg) ou, com o antagonista não peptídico, SB-612111 (6,6 µmol/kg), administrados por via intraperitoneal, durante três dias consecutivos, 30 min após a injeção diária de reserpina. No 4º dia, os animais também receberam um dos antagonistas, 30 min antes das avaliações comportamentais. Os animais foram submetidos aos testes de Von Frey, placa quente, nado forçado, labirinto em cruz elevado, rotarod e de preensão palmar. A expressão da pré-pró-nociceptina ppN/OFQ e do NOPr foi determinada por RT-qPCR e imunoistoquímica. O microPET [<sup>18</sup>F]-FDG foi utilizado para avaliar os padrões de ativação cerebral em camundongos tratados com reserpina. A distribuição do tamanho das fibras musculares do masseter e do gastrocnêmio foi avaliada através de análise histológica. A área e densidade das mitocôndrias no músculo esquelético foram analisadas por microscopia eletrônica de transmissão. Nos protocolos de tratamentos agudos, a administração i.t. ou i.p. de N/OFQ (1 nmol/sítio ou 1 nmol/kg, respectivamente) reduziu significativamente a alodínia mecânica induzida pela reserpina. Contudo, a administração i.p. de N/OFQ, na dose de 5

nmol/kg, teve efeito oposto, induzindo hipernocicepção. Em relação aos efeitos agudos do UFP-101, este antagonista peptídico, administrado pelas vias i.c.v. (1 nmol/sítio), i.t. (3 e 5 nmol/sítio) ou i.p. (1, 3 e 5 nmol/kg), reduziu significativamente a hipersensibilidade mecânica em camundongos tratados com reserpina. O tratamento agudo com N/OFQ ou UFP-101 não alterou significativamente a hipersensibilidade térmica, pelas vias i.c.v. ou i.p. A administração i.t. de N/OFQ (3 nmol/sítio) ou de UFP-101 (5 nmol/sítio) teve um efeito inibitório significativo na nocicepção térmica. No teste da natação forçada, a reserpina elevou o tempo de imobilidade, e este foi inibido de forma significativa pela N/OFQ, administrada pelas vias i.c.v (1 nmol/sítio) ou i.t. (3 nmol/sítio). N/OFQ e UFP-101 não modificaram nenhum parâmetro relacionado à ansiedade. Os tratamentos repetidos com UFP-101 e com SB-612111 reduziram a alodínia mecânica ( $37 \pm 8\%$  e  $43 \pm 15,2\%$ ), a hipernocicepção térmica ( $32,2 \pm 5\%$  e  $45 \pm 17,5\%$ ), melhoraram a coordenação motora no rotarod (aumento de 7 e 2 vezes no tempo de permanência) e a força de preensão palmar ( $15 \pm 16\%$  e  $9 \pm 5,5\%$ ), respectivamente. A administração de ambos antagonistas não foi capaz de alterar parâmetros de ansiedade ou depressão. A fibromialgia induzida por reserpina foi associada ao aumento na expressão de RNAm para a ppN/OFQ na medula espinhal lombar (dia 3) e no masseter (dias 1 e 2), enquanto a expressão do RNAm do NOPr foi aumentada no músculo masseter (dia 1). Alternativamente, a expressão de RNAm do NOPr foi reduzida no tálamo/hipotálamo (dia 3). A análise por imunoistoquímica revelou expressão aumentada do NOPr no gânglio da raiz dorsal (dia 4). O UFP-101 causou uma diminuição no metabolismo de [<sup>18</sup>F]-FDG no giro do cingulado, no colículo superior, no mesencéfalo esquerdo, no colículo inferior esquerdo e no colículo inferior direito de camundongos tratados com reserpina. Além disso, o UFP-101 previu as alterações induzidas pela reserpina na distribuição do tamanho das fibras musculares, de acordo com a avaliação dos cortes histológicos do masseter e do gastrocnêmio. Tanto a indução da fibromialgia pela reserpina, quanto o tratamento crônico com UFP-101,

não alteraram a área mitocondrial. Em resumos, os dados do presente estudo indicam que o bloqueio farmacológico do NOPr reduziu os sintomas de dor, fadiga e adinamia, recuperando também os padrões de ativação cerebral e as alterações musculares esqueléticas no modelo de fibromialgia experimental induzido pela reserpina. Além disso, expressão da ppN/OFQ e do NOPr foi alterada pela indução de fibromialgia, tanto em sítios centrais, quanto periféricos, reforçando a relevância do sistema N/OFQ-NOPr na patofisiologia da fibromialgia.

**Palavras-chave:** fibromialgia; reserpina; nociceptina/orfanina FQ; receptor da nociceptina/orfanina FQ; UFP-101; SB-612111; nocicepção; fadiga.

## **ABSTRACT**

Fibromyalgia is characterized by widespread pain, being accompanied by functional and affective disorders. This study evaluated the implication of nociceptin/orphanin FQ peptide receptor (NOPr) in a mouse model of fibromyalgia. The local Animal Ethics Committee approved the experimental protocols (15/00487). Fibromyalgia was induced in female CF-1 mice (20-24 g, 4 week-old) by reserpine administration (0.25 mg/kg; subcutaneous route), once a day, during 3 consecutive days. Control groups received vehicle. On the fourth day, mice were acutely treated with the selective NOP agonist nociceptin (N/OFQ), or with the selective peptide antagonist UFP-101, given by intraperitoneal (i.p., 0.3-5 nmol/kg), intracerebroventricular (i.c.v., 0.3-1 nmol/site), or intrathecal (i.t., 0.3-5 nmol/site) routes, 30 min before the experimental sessions. In a separate set of experiments, the animals were treated with the peptide UFP-101 (1 nmol/kg) or the non-peptide SB-612111 (6.6  $\mu$ mol/kg) antagonists, given by intraperitoneal route, during three consecutive days, 30 min after daily reserpine injection. At the 4<sup>th</sup> day, mice also received the antagonist, dosed 30 min before evaluations. The animals were subjected to Von Frey, hot-plate, forced swimming, elevated plus-maze, rotarod and grasping tests. Pre-pro-nociceptin (ppN/OFQ) and NOPr expression was determined by RT-qPCR and immunohistochemistry. The [<sup>18</sup>F]-FDG microPET imaging was used to assess the brain activation patterns in reserpine-treated mice. The fiber size distribution of masseter and gastrocnemius muscles was evaluated by histological analysis. The mitochondria area and density in the skeletal muscle were analysed by transmission electron microscopy (TEM). In the acute protocols of treatment, the i.t. or i.p. administration of N/OFQ (1 nmol/site or 1 nmol/kg, respectively) significantly reduced the mechanical allodynia. However, i.p. treatment with N/OFQ at the dose of 5 nmol/kg had an opposite effect, leading to hypernociception. Concerning the UFP-101 effects, this peptide antagonist given i.c.v. (1 nmol/site), i.t. (3 and 5 nmol/site) or i.p. (1, 3 and 5 nmol/kg)

significantly reduced the mechanical hypersensitivity in mice treated with reserpine. The acute treatment with N/OFQ or UFP-101 did not significantly alter the thermal hypersensitivity, when given by i.c.v. or i.p. routes. The i.t. administration of N/OFQ (3 nmol/site) and UFP-101 (5 nmol/site) had a significant inhibitory effect on the thermal nociception. The immobility time was significantly inhibited by N/OFQ, given by i.c.v (1 nmol/site) or i.t. (3 nmol/site) routes. N/OFQ and UFP-101 did not modify any anxiety-related parameter. The chronic treatment with UFP-101 and SB-612111 reduced the mechanical allodynia ( $37 \pm 8\%$  and  $43 \pm 15.2\%$ ) and the thermal hypernociception ( $32.2 \pm 5\%$  and  $45 \pm 17.5\%$ ), besides improving the motor coordination in the rotational apparatus (7 and 2-fold increase in permanence time) and the grasping strength ( $15 \pm 16\%$  and  $9 \pm 5.5\%$ ), respectively. None of the antagonists altered the parameters of anxiety or depression. Reserpine-induced fibromyalgia was associated with an increase in ppN/OFQ mRNA expression in the lumbar spinal cord (day 3) and masseter (days 1 and 2), whereas NOPr mRNA expression was increased in the masseter muscle (day 1). Alternatively, NOPr mRNA expression was reduced in the thalamus/hypothalamus (day 3). The immunohistochemistry analysis revealed an increased expression of NOPr in the dorsal root ganglion (DRG; on day 4). UFP-101 led to a decrease in the [ $^{18}\text{F}$ ]-FDG metabolism in cingulate gyrus, superior colliculus, left midbrain, left inferior colliculus and right inferior colliculus of reserpine-treated mice. Additionally, UFP-101 prevented reserpine-induced changes in fiber size distribution, according to the assessment of masseter and gastrocnemius histological sections. TEM analysis revealed that either the induction of fibromyalgia by reserpine, or the chronic treatment with UFP-101, did not alter the mitochondrial area or density. The expression of nociceptin and NOPr was altered in the mouse model of fibromyalgia induced by reserpine. Remarkably, UFP-101 improved the symptoms of pain, fatigue and adynamia, also recovering the brain activation patterns and the muscle fiber changes in this experimental paradigm. Our

data shed new lights on the mechanisms underlying the fibromyalgia pathogenesis, supporting a role for NOPr in this syndrome.

**Keywords:** fibromyalgia; reserpine; nociceptin/orphanin FQ; nociceptin/orphanin FQ receptor; UFP-101; SB-612111; nociception; fatigue.

## LISTA DE ILUSTRAÇÕES

### **FIGURAS**

<b>Figura 1.</b> Efeitos pleiotrópicos de nociceptina/orfanina FQ (N/OFQ) nos principais sistemas.....	23
<b>Figura 2.</b> Esquema para descrever a relação entre locais anatômicos subjacentes às ações do N/OFQ na dor.....	28
<b>Figura 3.</b> Potenciais processos fisiopatológicos na fibromialgia.....	44
<b>Figura 4.</b> Representação dos mecanismos envolvidos na patogênese da fibromialgia e alvos para o tratamento.....	46
<b>Figura 5.</b> Efeitos centrais e periféricos associados com a liberação de neuropeptídos pelas fibras C terminais.....	47

### **TABELAS**

<b>Tabela 1.</b> Efeitos pró-inflamatórios da ativação ou do bloqueio do NOPr.....	25
<b>Tabela 2.</b> Múltiplos efeitos de ligantes do NOPr sobre o processamento da dor.....	32
<b>Tabela 3.</b> Efeitos de antagonistas do NOPr em modelos pré-clínicos de depressão.....	36
<b>Tabela 4.</b> Efeitos de agonistas não-peptídicos do NOPr em modelos pré-clínicos de ansiedade.....	39

## **LISTA DE ABREVIATURAS**

**ACR** – Colégio Americano de Reumatologia

**ACTH** – Hormônio adrenocorticotrófico

**BDNF** – Fator neurotrófico derivado do cérebro

**CCI** – Injúria crônica do nervo ciático

**CCL2** – Ligante 2 de CC quimiocina

**CCL17** - Ligante 17 de CC quimiocina

**CCL22** - Ligante 22 de CC quimiocina

**CRH** – Hormônio liberador de corticotrofina

**CXCL1** – Ligante 1 de quimiocina CXC

**CXCL9** - Ligante 9 de quimiocina CXC

**CXCL11** - Ligante 11 de quimiocina CXC

**DRG** – Gânglio da raiz dorsal

**ERK** – Quinase regulada por sinal extracelular

**FDA** – *Food and Drug Administration*

**GABA** – Ácido gama-aminobutírico

**GHB** – Gama-hidroxibutirato

**HPA** – Eixo hipotálamo-pituitária-adrenal

**IASP** – *International Association for the Study of Pain*

**ICS** – Frio intermitente induzido pelo estresse

**IFN- $\gamma$**  – Interferon-gama

**JNK** – Proteína Jun N-terminal quinase

**LDH** – Lactato desidrogenase

**LPS** – Lipopolissacarídeo

**MadCAM-1** – Molécula de adesão celular de adressina da mucosa 1

**MAPK** – Proteínas quinases ativadas por mitógeno

**MIF** – Fator de inibição de migração de macrófagos

**MuRF1** - Muscle RING-finger protein-1

**N/OFQ** – Peptídeo nociceptina/orfanina FQ

**NDR** – núcleo dorsal da rafe

**NF-κB** – Fator de transcrição nuclear κB

**NGF** – Fator de crescimento do nervo

**NOPr** – Receptor do peptídeo nociceptina/orfanina FQ

**NRM** – núcleo magno da rafe

**PGD<sub>2</sub>** – prostaglandina D<sub>2</sub>

**PGE<sub>2</sub>** – Prostaglandina E<sub>2</sub>

**PKC** – Proteína quinase C

**PLA<sub>2</sub>** – Fosfolipase A<sub>2</sub>

**PLC** – Fosfolipase C

**ppN/OFQ** – Precursor pré-pró-N/OFQ

**SNC** – Sistema nervoso central

**SNL** – Modelo de ligação do nervo espinhal

**SNRI** – Inibidores da recaptação de serotonina e noradrenalina

**SSRI** – Inibidores seletivos da recaptação de serotonina

**STAT3** – Transdutor de sinal e ativador da transcrição 3

**TCC** – Terapia cognitivo-comportamental

**TNBS** – Colite induzida por ácido trinitrobenzeno sulfônico

**TNF** – Fator de necrose tumoral

**vIPAG** – Substância cinzenta periaquedatal ventrolateral

## **LISTA DE SÍMBOLOS E UNIDADES DE MEDIDAS**

$^{\circ}\text{C}$  Celsius

cm Centímetro

h Hora

kg Quilograma

M Molar

min Minuto

mg Miligramma

mL Mililitro

$\mu\text{g}$  Micrograma

$\mu\text{L}$  Microlitro

pmol Picomol

s Segundo

$\pm$  mais ou menos

$\alpha$  Alpha

$\beta$  Beta

$\gamma$  Gamma

$\delta$  Delta

## SUMÁRIO

<b>1 FUNDAMENTAÇÃO TEÓRICA.....</b>	<b>18</b>
1.1 Sistema nociceptina/orfanina FQ-receptor NOP .....	18
1.2 Transdução de sinal e heterodimerização do receptor NOP .....	18
1.3 Ligantes do receptor NOP .....	20
1.3.1 Agonistas e antagonistas peptídicos .....	20
1.3.2 Antagonistas não peptídicos .....	20
1.3.3 Moléculas com atividade bifuncional.....	21
1.4 Padrão de expressão do receptor NOP e do peptídeo N/OFQ .....	21
1.5 Ação biológica do Sistema N/OFQ-receptor NOP .....	23
1.5.1 Ação biológica da N/OFQ sobre o sistema imune .....	23
1.5.2 Ação biológica do sistema N/OFQ-NOPr no sistema nervoso central.....	27
1.5.2.1 Nocicepção .....	27
1.5.2.2 Ansiedade, estresse e depressão .....	35
1.5.2.3 Ligantes do NOPr na clínica.....	41
1.6 Fibromialgia .....	41
1.6.1 Diagnóstico da fibromialgia .....	43
1.6.2 Fisiopatologia da doença .....	43
1.6.3 Fibromialgia e marcadores inflamatórios .....	46
1.6.4 Tratamento .....	48
1.6.5 Fibromialgia e alterações musculares.....	50
1.6.6 Sistema nociceptina/orfanina FQ-NOPr na fibromialgia .....	51
<b>2 JUSTIFICATIVA .....</b>	<b>52</b>
<b>3 OBJETIVOS .....</b>	<b>53</b>
3.1 Objetivo Geral .....	53
3.2 Objetivos Específicos .....	53
<b>4 MANUSCRITO DO TRABALHO EXPERIMENTAL.....</b>	<b>55</b>
<b>5 CONSIDERAÇÕES FINAIS.....</b>	<b>125</b>
<b>6 PERSPECTIVAS.....</b>	<b>128</b>
<b>REFERÊNCIAS BIBLIOGRÁFICAS .....</b>	<b>130</b>
<b>ANEXO A – APROVAÇÃO DA CEUA.....</b>	<b>146</b>
<b>ANEXO B – ACEITE DO MANUSCRITO – ARTIGO EM PRODUÇÃO .....</b>	<b>147</b>

## **1 FUNDAMENTAÇÃO TEÓRICA**

### **1.1 Sistema nociceptina/orfanina FQ-receptor NOP**

Este sistema é composto pelo peptídeo nociceptina/orfanina FQ (N/OFQ) e pelo receptor de N/OFQ (NOPr) (1). O receptor da N/OFQ foi classificado como um membro da família de receptores opioides, sendo denominado como receptor opioide do tipo 1 (ORL-1). Posteriormente, a IUPHAR (International Union of Basic & Clinical Pharmacology) denominou o receptor de OP4 e, subsequentemente, como receptor do peptídeo nociceptina/orfanina FQ (NOPr) (2). O peptídeo N/OFQ é produzido a partir do precursor pré-pró-N/OFQ (ppN/OFQ), que é constituído por 176 aminoácidos (3). A N/OFQ contém 17 aminoácidos (Phe-Gly-Gly-Phe-Thr-Gly-Ala-Arg-Lys-Ser-Ala-Arg-Lys-Leu-Ala-Asn-Gln) e foi descrita pela primeira vez em 1995, como o ligante endógeno do NOPr. De acordo com a sua estrutura, o peptídeo possui similaridade com peptídeos opioides, como a dinorfina. Por isto, geralmente é relatado como um peptídeo opioide. Entretanto, não possui afinidade pelos receptores opioides MOP ( $\mu$ ), DOP ( $\delta$ ) e KOP ( $\kappa$ ) (1, 4, 5). De maneira semelhante, os opioides endógenos não se ligam ao NOPr, pois apresentam baixa afinidade pelo mesmo (6). Esta seletividade distinta tem sido atribuída a três resíduos do NOPr que diferem dos demais receptores opioides, Ala216 de Lys, Gln280 de His e, Thr305 de Ile, respectivamente. Isto está intimamente relacionado com os motivos estruturais dos peptídeos ligantes, sendo que as partes hidrofóbicas e hidrofílicas das bolsas de ligação de receptores de opioides diferem do NOPr (7, 8).

### **1.2 Transdução de sinal e heterodimerização do receptor NOP**

O NOPr é acoplado à proteína G (GPCR) e é um membro não-opioide da família de receptores opioides, sendo insensível ao antagonista naloxona (6). Quando o NOPr é ativado pelo peptídeo N/OFQ, desencadeia uma cascata de eventos, incluindo a inibição de canais de  $\text{Ca}^+$  e da proteína adenilato ciclase, além da ativação do canal de  $\text{K}^+$  retificador de corrente, reduzindo assim,

a excitabilidade neuronal e a liberação de neurotransmissores na fenda sináptica (4, 5, 9-12). A redução na liberação de neurotransmissores, como catecolaminas (dopamina, e noradrenalina), serotonina (5-HT), acetilcolina e glutamato é observada após a ativação do NOPr (13-16). Esta ação é a base para a modulação de transtornos como ansiedade, depressão e dor.

Os mecanismos pelos quais o NOPr interage com os canais de cálcio ainda não estão bem elucidados. Altier e colaboradores (17) mostraram que a ativação prolongada do receptor por N/OFQ leva à internalização dos canais de cálcio do tipo N (neuronal) em vesículas e, à redução da entrada de cálcio, sendo que este efeito é revertido pelo bloqueio do receptor com o antagonista peptídico III-BTD. Estes dados indicam que ocorre a remoção dos canais da membrana plasmática. Outro trabalho demonstrou que há a heterodimerização do NOPr com o receptor opioide MOP e que a posterior ativação de MOP leva à internalização dos canais de cálcio, somente na presença de NOP. No entanto, esta inibição é menor quando comparada com aquela mediada por NOP apenas (18). Contudo, Murali e cols. (19) verificaram que não houve a internalização destes canais no corpo celular e nos terminais nervosos centrais de neurônios do gânglio da raiz dorsal de ratos. Vale ressaltar que este mecanismo é fundamental no controle da transmissão de sinais nociceptivos.

A participação do NOPr também tem sido descrita em outras vias importantes, como na ativação da proteína quinase C (PKC) (20), da fosfolipase A<sub>2</sub> (PLA<sub>2</sub>) (21) e da fosfolipase C (PLC) (20), na modulação das proteínas quinases ativadas por mitógeno (MAPK), que incluem a quinase regulada por sinal extracelular (ERK) e a proteína Jun N-terminal quinase (JNK), além do fator de transcrição nuclear κB (NF-κB) (20, 22-26). Mais recentemente, o transdutor de sinal e ativador da transcrição 3 (STAT3) tem sido implicado nas vias de transdução deste receptor (27).

### **1.3 Ligantes do receptor NOP**

#### **1.3.1 Agonistas e antagonistas peptídicos**

O [F/G]N/OFQ(1–13)-NH<sub>2</sub> foi descrito como o primeiro ligante do NOPr com eficácia reduzida, sendo gerado a partir de mudanças na ligação entre Phe<sup>1</sup> e Gly<sup>2</sup> (28). Já, o primeiro antagonista seletivo para o NOPr ([Nphe<sup>1</sup>] N/OFQ (1-13) NH<sub>2</sub>) surgiu das modificações realizadas na porção N-terminal de N/OFQ, através da mudança de Phe<sup>1</sup> da cadeia lateral do átomo C para o átomo N (29). Contudo, este peptídeo apresentou uma baixa potência. Outra modificação foi realizada em N/OFQ [Arg<sup>14</sup>, Lys<sup>15</sup>] e forneceu um agonista bastante potente (30X mais potente que N/OFQ) (30). Foi observado um efeito pró-nociceptivo após a administração intracerebroventricular (i.c.v.) deste peptídeo, como avaliado no teste de retirada da cauda, além de produzir inibição da atividade locomotora de camundongos (31).

O antagonista [Nphe<sup>1</sup>, Arg<sup>14</sup>, Lys<sup>15</sup>] N/OFQ-NH<sub>2</sub> foi produzido a partir da junção das duas modificações citadas acima em uma única molécula. Também chamado de UFP-101, este peptídeo é um antagonista altamente seletivo para o NOPr (32, 33), demonstrando várias atividades biológicas, incluindo ação anti-inflamatória (34) e efeitos antiarrítmicos (35), dentre outras.

#### **1.3.2 Antagonistas não peptídicos**

O J-113397 foi descrito como o primeiro antagonista não peptídico com uma alta potência (pA<sub>2</sub> 7.5–8.0) e uma seletividade aceitável para o NOPr (36). No entanto, outro antagonista, o SB-612111, apresentou potência mais elevada (pA<sub>2</sub> 8.0–8.5) que o J-113397, com seletividade comparável ao mesmo (37). Ademais, outra molécula foi caracterizada como um antagonista com melhores propriedades farmacológicas, apresentando alta potência (pA<sub>2</sub> 8.5–9.0), denominado composto 24 (C-24) (38-42).

### **1.3.3 Moléculas com atividade bifuncional**

Em uma tentativa de desenhar novas moléculas com atividade bifuncional, alguns grupos têm utilizado a integração de farmacóforos que têm afinidade por diferentes receptores, como NOP e MOP, simultaneamente (43, 44). Este conceito de ligantes com múltiplos alvos é baseado na terapia com opioides, com a utilização, por exemplo, de buprenorfina, sendo este um agonista de MOP e antagonista de KOP (45, 46). Sobczak e cols. (47) verificaram ação analgésica e inibição da motilidade gastrointestinal para o agonista BU08070, um análogo de buprenorfina, em um modelo de síndrome do intestino irritável. Este agonista possui atividade bifuncional, pois é ligante dos receptores NOP e MOP. Neste estudo, o efeito antinociceptivo do BU08070 foi mediado pelo receptor MOP, enquanto que sua ação sobre o trato gastrointestinal foi mediada pelo NOPr.

### **1.4 Padrão de expressão do receptor NOP e do peptídeo N/OFQ**

O peptídeo N/OFQ e o NOPr são amplamente expressos em células imunes, no sistema nervoso central (SNC) e, em diversos órgãos periféricos (48). Em particular, o NOPr é expresso no sistema aminérgico (núcleos adrenérgico, colinérgico, dopaminérgico e serotoninérgico). Este também é encontrado no sistema límbico, incluindo o complexo amigdaloide, o hipocampo, vários núcleos do hipotálamo, a banda diagonal de Broca, dentre outras áreas envolvidas no processamento das emoções. O receptor também está distribuído pelas vias olfatória, auditiva e visual, da percepção somatossensorial e no controle motor (49).

Nos tecidos periféricos, foi detectada a expressão do NOPr na mucosa intestinal humana (50) e de ratos (51), no miocárdio humano (52) e nos gânglios neuronais simpáticos e sensoriais periféricos de cobaias (53) e ratos (54). Os receptores da nociceptina são encontrados amplamente distribuídos nos neurônios (grandes e pequenos, mielinizados e não mielinizados) do gânglio da raiz dorsal (DRGs), onde 43% de todos os neurônios expressam o NOPr (55). Estes receptores estão presentes tanto nos neurônios peptidérgicos, como não peptidérgicos, importantes para a dor aguda

pelo calor e para a dor mecânica, respectivamente (56-60). Os receptores da nociceptina também estão co-localizados com os receptores opioides MOP nas fibras C peptidérgicas (55, 61).

Em condições patológicas, como no modelo de dor neuropática de ligação do nervo ciático (SNL), a expressão do RNAm da N/OFQ e do NOPr está diminuída no tálamo e no hipotálamo e aumentada no córtex cingulado anterior (62). Ainda neste estudo, um aumento na expressão do RNAm da N/OFQ também foi observado na amígdala. Neste mesmo modelo, Ozawa et al. (2018) relataram a diminuição da imunoreatividade ao NOP-eGFP nas lâminas I e II externas na medula e nos aferentes primários nos DRGs de L4 (lombar 4), sem alteração da expressão na borda ventral da lâmina II interna (63). No modelo induzido por injúria crônica do nervo ciático (CCI), a expressão do NOPr está elevada no núcleo magno da rafe (NRM), substância cinzenta periaquedatal ventrolateral (vlPAG) e núcleo dorsal da rafe (NDR) (64). Em adição, os níveis de proteína do NOPr estão aumentados na medula espinal dorsal ipsilateral de ratos com CCI (65). Outros estudos demonstram a modulação do NOPr em diferentes modelos de dor (66-68). Quanto à localização de NOPr em tecidos humanos, um estudo demonstrou o aumento de fibras positivas para o receptor no sub-urotélio da bexiga de pacientes com bexiga hiperativa e com síndrome da bexiga dolorosa (69). Anand e cols. (2016) mostraram também que 75% a 80% dos neurônios pequenos/médios nos DRGs lombares e sacrais humanos eram positivos para NOPr, e que a imunorreatividade do NOPr foi diminuída nos nervos periféricos lesados e nos neuromas dolorosos (69). Stamer et al. (2011) também demonstrou a modulação da expressão da N/OFQ e do NOPr em células sanguíneas periféricas de pacientes com câncer ou sépticos (70) e Sobczak et al. (2011) do NOPr em amostras do cólon de pacientes com doença inflamatória intestinal (71). Entretanto, a modulação da expressão da N/OFQ e de seu receptor no DRG ainda não foi investigado em modelos de dor disfuncional, como a fibromialgia.

## 1.5 Ação biológica do Sistema N/OFQ-receptor NOP

O sistema N/OFQ- NOPr afeta uma variedade de sistemas biológicos (1) (Figura 1). Assim, muitos grupos têm estudado agonistas e antagonistas do NOPr como potenciais ferramentas terapêuticas. Experimentos *in vivo* demonstram a modulação de uma diversidade de funções biológicas, como vasodilatação (72). Os efeitos dos ligantes desses receptores também têm sido caracterizados na dor, depressão e na ansiedade (1, 7, 73-81). Além disto, a sua ação sobre a aprendizagem e memória foi também descrita (82, 83).

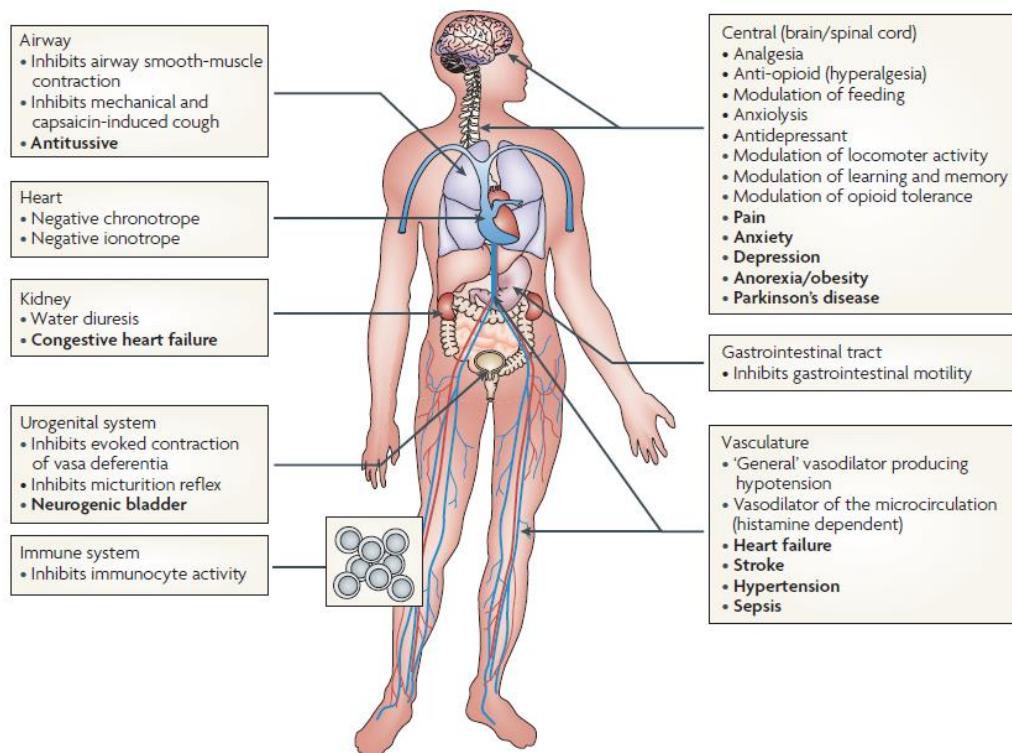


Figura 1: Efeitos pleiotrópicos de nociceptina/orfanina FQ (N/OFQ) nos principais sistemas. Indicações clínicas potenciais estão em negrito.

Extraído de (1) – Publicação aprovada pela revista (Número de licença: 3681510776945).

### 1.5.1 Ação biológica da N/OFQ sobre o sistema imune

Mais recentemente, estudos *in vitro* e *in vivo* mostraram o efeito do sistema N/OFQ-NOP na modulação de funções imunes, sendo que o RNA mensageiro (RNAm) do NOPr foi detectado em linfócitos T citotóxicos e T-helper de camundongos (84), em linhagens de linfócitos T e monócitos

(U937) e, em monócitos e linfócitos humanos (85, 86). A expressão de RNAm também foi relatada em órgãos linfoides de suínos, nos linfonodos, no timo e no baço (87). Além da expressão nos órgãos linfoides, a N/OFQ e o NOPr são expressos na epiderme e na mucosa intestinal, importantes sítios para reconhecimento de抗ígenos (50, 88, 89).

A N/OFQ modula diferentes respostas imunes, induzindo a quimiotaxia de neutrófilos (90), e de monócitos humanos (91), promovendo a migração de leucócitos (90), aumentando a permeabilidade vascular (92), além de induzir hipotensão, rolamento de leucócitos e vazamento macromolecular (34, 72). O peptídeo também estimula a liberação de histamina pelos mastócitos (92).

Vários trabalhos demonstraram que agonistas e antagonistas do NOPr induzem efeitos pró- e anti-inflamatórios, respectivamente, em vários modelos de sepse. Carvalho e cols. (93) utilizaram o modelo de sepse induzida por punção e ligadura cecal em ratos Wistar e verificaram que o antagonista peptídico do NOPr, o UFP-101, administrado por via subcutânea (s.c.), na dose de 0,03 mg/kg, reduziu a mortalidade em 50%. Em adição, houve a diminuição do infiltrado inflamatório no fluido broncoalveolar e no exsudato peritoneal, prevenção da disseminação bacteriana e, diminuição da concentração plasmática das citocinas fator de necrose tumoral (TNF) e interleucina-1 beta (IL-1 $\beta$ ). Foram identificados efeitos anti-inflamatórios para o UFP-101 no modelo de sepse induzida por lipopolissacarídeo (LPS) (34). Neste estudo, o extravasamento plasmático e o rolamento de leucócitos induzidos por LPS foram reduzidos pela co-administração endovenosa de 286,23  $\mu$ g/kg do antagonista.

No modelo de colite em camundongos, foi observada a inibição da produção dos mediadores inflamatórios interferon-gama (IFN- $\gamma$ ), TNF- $\alpha$ , IL-1 $\beta$  e ligante 1 de quimiocina CXC (CXCL1), após o tratamento com o antagonista não-peptídico, SB-612111, na dose de 30 mg/kg (94). De forma interessante, camundongos deficientes para NOPr apresentaram diminuição nos níveis da molécula de adesão celular de adressina da mucosa 1 (MadCAM-1) e, do número de células inflamatórias na mucosa do cólon (95).

Ao contrário dos efeitos anti-inflamatórios dos antagonistas, a administração de N/OFQ pela via endovenosa, nas doses de 1,085 µg/kg a 108,54 µg/kg, em ratos Wistar, ocasionou vasodilatação, hipotensão e adesão de leucócitos (72). Além disto, a aplicação intradérmica do agonista endógeno aumentou a permeabilidade vascular na pele dos ratos, por um mecanismo dependente do receptor H1 de histamina (92).

Na literatura, também são encontrados resultados que discordam dos achados acima citados, como o efeito anti-inflamatório de agonistas não-peptídicos do NOPr. Sobczak e cols. (71) demonstraram atividade anti-inflamatória e antinociceptiva para o SCH 221510, um agonista não peptídico e altamente seletivo para o NOPr (96).

Abaixo estão descritos os efeitos pró-inflamatórios de ligantes do NOPr (48) (Tabela 1).

**Tabela 1:**

**Table 1 Available evidence for a proinflammatory effects of NOP activation  
Effects of NOP activation or NOP blockage**

<b>In vivo and in vitro studies</b>	<b>Effects of NOP activation or NOP blockage</b>	<b>References</b>
C57BL/6J mice (normal and ppN/OFQ knockout)	<ul style="list-style-type: none"> <li>The administration of N/OFQ (55 nmol/kg, i.p.) 30 min prior to Staphylococcal enterotoxin A increased the expression of TNF-<math>\alpha</math> and IFN-<math>\gamma</math> on the spleen</li> <li>N/OFQ-deficient mice displayed attenuated TNF-<math>\alpha</math> and IFN-<math>\gamma</math> mRNA levels triggered by antigen challenge</li> </ul>	Goldfarb, Reinscheid, and Kusnecov (2006)
Rat astrocytes	<ul style="list-style-type: none"> <li>The expression of N/OFQ mRNA and protein was increased by proinflammatory mediators such as TNF-<math>\alpha</math>, IL-1<math>\beta</math>, and LPS</li> </ul>	Buzas, Rosenberger, Kim, and Cox (2002)
Rat splenocytes	<ul style="list-style-type: none"> <li>TNF-<math>\alpha</math> and IL-1-<math>\beta</math> increased the N/OFQ secretion by splenocytes <i>in vitro</i></li> </ul>	Miller and Fulford (2007)
Anesthetized Wistar rats	<ul style="list-style-type: none"> <li>Administration of N/OFQ (0.6–60 nmol/kg i.v.) caused hypotension, vasodilatation, macromolecular leak, and leukocyte adhesion</li> </ul>	Brookes et al. (2007)

Anesthetized Wistar rats	<ul style="list-style-type: none"> <li>Administration of N/OFQ (0.6–60 nmol/kg i.v.) caused hypotension, vasodilatation, macromolecular leak, and leukocyte adhesion</li> </ul>	Brookes et al. (2007)
Wistar rats and isolated mast cell	<ul style="list-style-type: none"> <li>Intradermal application of N/OFQ increased vascular permeability in rat skin by a mechanism dependent of histamine H1 receptor</li> <li><i>In vitro</i> N/OFQ stimulated the release of histamine by rat peritoneal mast cells</li> </ul>	Kimura et al. (2000)
ICR mice and C57BL/6 NOP-deficient mice	<ul style="list-style-type: none"> <li>Intradermal inoculation of N/OFQ presented pruritogenic effect in normal but not in NOP-deficient mice. The leukotriene B<sub>4</sub> receptor antagonist inhibited the itch</li> <li>N/OFQ stimulated the production of leukotriene B<sub>4</sub> by keratinocytes</li> </ul>	Andoh et al. (2004)
Monocytes and neutrophils obtained from healthy subjects	<ul style="list-style-type: none"> <li>NOP activation stimulated the chemotaxis of human monocytes and increased the release of lysozyme by neutrophils</li> </ul>	Trombella et al. (2005)
Neutrophils obtained from healthy volunteers	<ul style="list-style-type: none"> <li>N/OFQ exhibited a potent chemoattractant activity <i>in vitro</i></li> </ul>	Serhan et al. (2001)
BALB/c mice (air pouch model)	<ul style="list-style-type: none"> <li>N/OFQ at low doses (10 ng) induced significant leukocyte recruitment into the air pouch</li> </ul>	Serhan et al. (2001)
Human neutrophils	<ul style="list-style-type: none"> <li>Neutrophils stimulated by fMLP quickly secreted N/OFQ upon exocytosis of granules</li> </ul>	Fiset et al. (2003)
Septic rats (CLP model)	<ul style="list-style-type: none"> <li>Pharmacological blockade of NOP receptor with UFP-101 enhanced the bacterial control and decreased systemic inflammation and mortality of animals, while N/OFQ administration increased animal mortality</li> </ul>	Carvalho et al. (2008)

Colitic mice (DSS model)	<ul style="list-style-type: none"> <li>The NOP receptor antagonist Alt et al. (2012) (SB612111—30 mg/kg) ameliorated the clinical signs of colitis and inhibited the production of CXCL1, IFN-<math>\gamma</math>, TNF-<math>\alpha</math>, IL-1<math>\beta</math>, IL-6, and TNF-<math>\alpha</math></li> </ul>	
Colitic mice (DSS model: wild-type and NOP-deficient C57BL/6 mice)	<ul style="list-style-type: none"> <li>NOP-deficient animals developed attenuated DSS-induced colitis and expressed decreased levels of mucosal addressin (MadCAM-1) and significant reduction in the number of inflammatory cells in colonic mucosa</li> </ul>	Kato et al. (2005)

CXCL1, chemokine (C-X-C motif) ligand 1; fMLP, proinflammatory peptide N-formyl-L-methionyl-L-leucyl-L-phenylalanine; IFN- $\gamma$ , interferon-gamma; IL-1 $\beta$ , interleukin 1 beta; LPSs, lipopolysaccharides; MadCAM-1, mucosal addressin cell adhesion molecule-1; N/OFQ, nociceptin/orphanin FQ peptide; NOP, N/OFQ receptor; UFP-101, University of Ferrara Peptide-101; TNF- $\alpha$ , tumor necrosis factor-alpha.

Extraído de (48) – Publicação aprovada pela revista (Número de licença: 4524251386841).

## 1.5.2 Ação biológica do sistema N/OFQ-NOPr no sistema nervoso central

### 1.5.2.1 Nocicepção

A sensação dolorosa inicia com um estímulo nocivo, detectado na periferia (pele e outros tecidos fora do SNC) pelos nociceptores, que possuem seus corpos celulares no DRG. Estas fibras nervosas sensoriais aferentes são ativadas na maioria das alterações dolorosas (fibras de alto limiar C e A $\delta$ ). Os nociceptores transmitem o impulso para os neurônios do corno dorsal da medula espinhal, e estes, conduzem o impulso até o cérebro como potencial de ação (57, 97). As principais alterações sensoriais são a alodínia (dor provocada por um estímulo não nocivo) e a hiperalgésia (sensibilidade aumentada para um estímulo doloroso) (98), presentes na fibromialgia.

Em relação às ações centrais do sistema N/OFQ–NOP, os principais estudos se concentram no seu papel sobre o processamento da dor. De acordo com o trabalho de Meunier e cols. (4), a denominação da nociceptina foi baseada em seus efeitos hiperalgésicos, após sua administração por via i.c.v. Lambert (2008) demonstrou que a morfina inibe as células ON na medula ventromedial

rostral, sendo incapazes de inibir as células OFF, que reduzem a informação ascendente nociceptiva ao nível medular, originando assim, um efeito de analgesia. É importante destacar que este efeito é revertido pela ação de N/OFQ. Desta maneira, a N/OFQ possui um efeito nociceptivo quando administrado ao nível supra-espinal, mediado pela inibição de células ON (primárias) e células OFF (secundárias) na medula ventromedial rostral. Por outro lado, a analgesia opioide clássica, decorrente da administração da N/OFQ por via intratecal (i.t.), parece estar relacionada com a inibição do afluxo aferente nociceptivo, que ocorre em tecidos periféricos. A inibição deste fluxo aferente também pode ocorrer perifericamente, sobre a pele, a bexiga e/ou sobre células sanguíneas mononucleares (1). A figura abaixo demonstra os prováveis sítios de ação da N/OFQ na modulação da nocicepção (Figura 2).

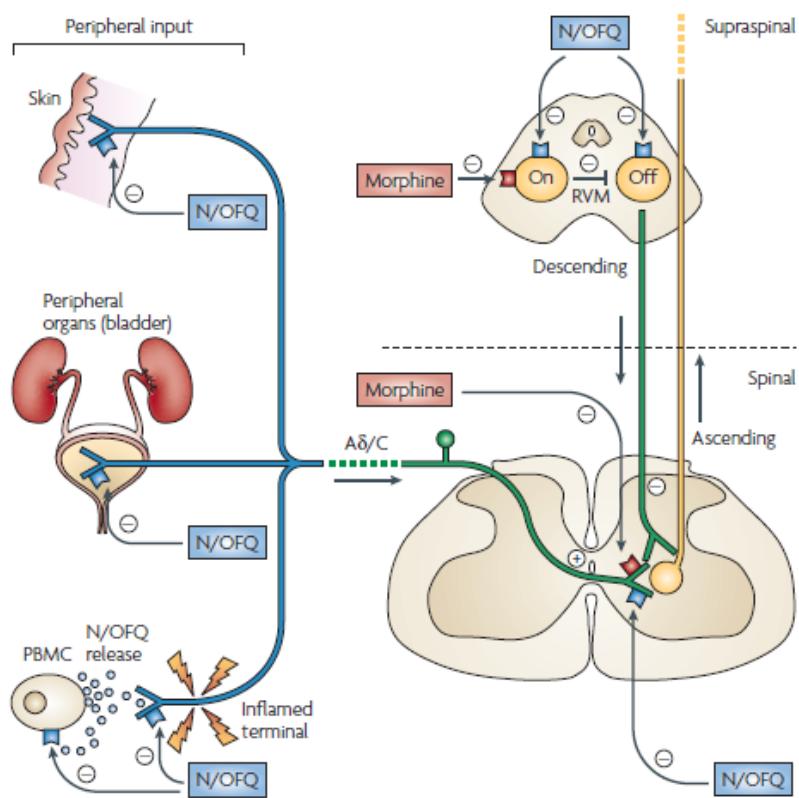


Figura 2: Esquema para descrever a relação entre locais anatômicos subjacentes às ações do N/OFQ na nocicepção.

RVM: medula ventromedial rostral. PBMC: células sanguíneas mononucleares periféricas.

Extraído de (1) – *Publicação aprovada pela revista (Número de licença: 3682510662258)*.

Os efeitos de agonistas do NOPr sobre a dor podem levar tanto à nocicepção quanto à antinocicepção em roedores, dependendo das diferentes vias de administração, das doses utilizadas e da modalidade da dor (7, 73). Com relação à dor aguda, quando da administração de N/OFQ periférica em camundongos, foi observada redução significativa da nocicepção induzida por capsaicina (99). Em contrapartida, o antagonista não peptídico, SB-612111, administrado por via intravenosa, antagonizou a atividade antimorfina e a hiperalgesia térmica induzidas pela N/OFQ (100). É importante ressaltar que os efeitos da administração sistêmica em roedores de agonistas do NOPr são dependentes da soma dos efeitos supra-espinhal e espinal, relativos à sinalização do sistema N/OFQ- NOPr (73).

Este mesmo efeito de analgesia foi verificado em ratos no teste de retirada da cauda, com a administração i.t. ao nível da cauda equina de [Phe<sup>1</sup>psi(CH<sub>2</sub>-NH)Gly<sup>2</sup>]N/OFQ(1-13)-NH<sub>2</sub>, um análogo de N/OFQ (101). Em adição, Tian e cols. (102) demonstraram que a administração i.t. de N/OFQ produziu efeito antinociceptivo em ratos no teste de retirada da cauda, além de potencializar os efeitos da morfina. Corroborando estes achados, Tsai et al. (2018) demonstraram que a injeção i.t. de N/OFQ (1, 2 ou 5 nmol/sítio) levou ao aumento da latência no teste de retirada da cauda (103), sendo este efeito antagonizado por UFP-101 (10 nmol/sítio). Por outro lado, a administração de N/OFQ (10 nmol/sítio) na vIPAG de ratos bloqueou a analgesia provocada por DAMGO (um agonista do receptor MOP, na dose de 1,9 nmol/sítio), enquanto UFP-101 (10 nmol/sítio) potencializou a ação do opioide (0,5 nmol/sítio) (104). Em adição, após a injeção de [Phe<sup>1</sup>psi(CH<sub>2</sub>-NH)Gly<sup>2</sup>]N/OFQ(1-13)-NH<sub>2</sub>, por via i.c.v., foi observada resposta pró-nociceptiva no teste de retirada da cauda em ratos (101) e camundongos (5). Além disso, baixas doses do peptídeo, administradas perifericamente ou por via i.t., induziram algesia em camundongos (105, 106). Neste mesmo teste, Calo e cols. (2002) (32) demonstraram que o antagonista UFP-101 (3 nmol/sítio, i.c.v.) previu o efeito pró-nociceptivo da N/OFQ (1 nmol/sítio, i.c.v.). Em adição, o antagonista produziu efeito antinociceptivo *per se*, apenas na dose de 10 nmol. Quando co-injetados (N/OFQ 1 nmol/sítio + UFP-101 10 nmol/sítio), o efeito pró-nociceptivo do peptídeo natural e o efeito

antinociceptivo do antagonista foram abolidos, atingindo valores aproximados aos observados nos animais controle. O antagonista SB-612111, administrado por via i.p. (3 mg/kg), também foi capaz de antagonizar as ações pró-nociceptiva e antinociceptiva da N/OFQ (1 nmol), por via i.c.v. e i.t., respectivamente (107). Este antagonista, na dose de 10 mg/kg, por via s.c., também foi capaz de potencializar os efeitos antinociceptivos de agonistas de NOP/MOP, a buprenorfina (1 e 3 mg/kg; via s.c.), SR16435 e (3 – 30 mg/kg; via s.c.) e SR16507 (0.3 e 3 mg/kg; via s.c.) (108). Outro agonista bifuncional, o cebranopadol, apresentou efeito antinociceptivo no teste de retirada da cauda nas doses de 0.01 – 1 mg/kg, por via i.v. (109).

Em modelos de dor inflamatória em roedores, agonistas do NOPr, quando administrados por via i.t. ou supra-espinhal, induzem ações antinociceptiva e pró-nociceptivas, respectivamente (7). Agostini e cols. (51) demonstraram que a administração periférica de N/OFQ (2 nmol/kg) teve efeito anti-hiperalgésico no modelo de colite induzida por ácido trinitrobenzeno sulfônico (TNBS) em ratos Wistar. Neste mesmo estudo, a administração periférica de UFP-101 (10 nmol/kg) inibiu a ação da N/OFQ. No teste de formalina em camundongos, a administração de UFP-101, na dose de 10 nmol/sítio, pelas vias i.c.v. e i.t., exerceu efeitos analgésico e algésico, respectivamente (110). Em contrapartida, nos modelos de dor neuropática, efeitos analgésicos são verificados tanto pela via i.t., quanto supra-espinhal. Isto pode ser explicado pela ativação do NOPr nas duas vias (i.t. e supra-espinhal), após administração sistêmica de N/OFQ, onde foi observado um potente efeito analgésico (7). Exemplo disto foi observado com a injeção s.c. dos agonistas não peptídicos, SR14150 (10 mg/kg; via s.c.) e SR16835 (30 mg/kg; via s.c.), que induziram ação anti-alodínica no modelo SNL em camundongos ICR (111). Neste estudo, os autores demonstraram que o circuito envolvido na dor crônica e aguda são diferentes, onde SB-612111 (10 mg/kg; via s.c.) bloqueou o efeito anti-alodínico dos dois agonistas não peptídicos, mas não previneu a ação antinociceptiva de SR14150 (3 e 10 mg/kg; via s.c.). O SB-612111 também foi responsável pela potencialização dos efeitos da morfina de antinociceção (3 mg/kg; via s.c.) e inibiu a alodínia (1 e 3 mg/kg; via s.c.) (111). Um estudo mais recente, relatou os efeitos espinhais nociceptivos para o agonista PWT2-N/OFQ (2,5,

25 e 250 pmol/site) e para a N/OFQ (0,1, 1 e 10 nmol/site), em modelos de dor neuropática e nociceptiva, sendo estes efeitos bloqueados por SB-612111 (112).

A administração i.t. de N/OFQ inibiu a alodínia no modelo de dano por constrição crônica do nervo ciático em ratos (113). Neste mesmo modelo, a infusão do antagonista UFP-101 intra-vIPAG, na dose de 18 µg/1 µl/rato, reverteu a diminuição do limiar alodínico (114). Um estudo recente com o modelo de SNL, relatou o papel do sistema N/OFQ-NOPr na diminuição da hiperalgesia induzida pelo dano. Esta ação é resultante da inibição direta dos neurônios na medula espinhal (63). Outro estudo demonstrou que a administração periodontal de UFP-101 aliviou a dor orofacial induzida pela movimentação dentária experimental em ratos. Este mesmo efeito não foi visto para a administração intraperitoneal do antagonista (115). Além dos efeitos observados para o antagonista UFP-101 sobre modelos de dor crônica, também foram descritos efeitos benéficos para agonista AT-200 (10 mg/kg; s.c.) sobre a hiperalgesia musculoesquelética térmica e mecânica em camundongos com anemia falciforme, sendo esta atividade revertida por SB-612111 (10 mg/kg; s.c.) (116).

Em contrapartida, diferentemente do observado em roedores, a administração i.t. de agonistas do NOPr, independentemente da dose, levam à somente à antinocicepção, em modelos de dor em primatas, como revisado por Kiguchi et al. (2016) (73). Em um modelo de dor aguda (retirada da cauda da água morna à 50 °C, como estímulo nocivo), a administração espinal de N/OFQ (10-100 nmol/sítio) ou PWT2-N/OFQ (doses de 0,3 – 3 nmol/sítio) induziu efeitos antinociceptivos em macacos (112, 117). Este mesmo efeito foi demonstrado para o agonista não peptídico Ro 64-6198 (0,001–0,06 mg/kg, s.c.), além de uma ação anti-alodínica no modelo induzido por capsaicina. Neste estudo, o antagonista J-113397 (0,01–0,1 mg/kg, s.c.) foi capaz de reverter o efeito de Ro 64-6198 (118). As ações supraespinhais da N/OFQ em primatas levam à antinocicepção, diferentemente do que é visto em roedores (73). Ding e cols. (2015) demonstraram que a administração intracisternal da N/OFQ induziu efeitos antinociceptivos de maneira dose-dependente, sendo estes efeitos bloqueados pelo antagonista J-113397 do NOPr (117). As ações

sistêmicas de agonistas do NOPr, de maneira geral, levam à analgesia, independente da modalidade da dor, como visto em roedores (73). Estudos demonstram que a administração sistêmica dos agonistas do NOPr, Ro64-6198 e SCH 221510, induz efeitos anti-alodínico e anti-hiperalgésico em modelos de dor induzidos por capsaicina e por carragenina (73, 118-121).

Abaixo estão descritos os principais efeitos de agonistas e antagonistas do NOPr em modelos de nocicepção (73) (Tabela 2).

**Tabela 2**

Multiple effects of NOP receptor-related ligands on regulating pain processing.

NOP receptor-related ligands	Findings in rodents	Findings in primates
<b><i>NOP Receptor Agonists (Peptides)</i></b>		
N/OFQ	Spinal, Acute pain ↓ (Xu et al., 1996) (Erb et al., 1997) (King et al., 1997) (Yamamoto et al., 1997a)	Spinal, Acute pain ↓ (Ko et al., 2006) (Ko & Naughton, 2009)
	Spinal, Acute pain ↑ (Inoue et al., 1999) (Sakurada et al., 1999)	
	Spinal, Inflammatory pain ↓ (Yamamoto et al., 1997b) (Hao et al., 1998) (Chen & Sommer, 2007)	
	Spinal, Neuropathic pain ↓ (Yamamoto & Nozaki-Taguchi, 1997) (Corradini et al., 2001) (Courteix et al., 2004)	
	Supraspinal, Acute pain ↑ (Meunier et al., 1995) (Reinscheid et al., 1995)	Supraspinal, Acute pain ↓ (Ding et al., 2015b)
	Supraspinal, Inflammatory pain ↑ (Zhu et al., 1997) (Wang et al., 1999a)	

[Phe <sup>1</sup> ψ(CH <sub>2</sub> -NH)Gly <sup>2</sup> ]N/OFQ-(1-13)-NH <sub>2</sub>	Supraspinal, Acute pain ↑ (Calo et al., 1998) (Wang et al., 1999b)	
	Supraspinal, Inflammatory pain ↑ (Bertorelli et al., 1999)	
UFP-112	Spinal, Acute pain ↓ (Rizzi et al., 2007) (Calo et al., 2011)	Spinal, Acute pain ↓ (Hu et al., 2010)
		Spinal, Capsaicin-induced allodynia ↓ (Hu et al., 2010)
PWT2-N/OFQ	Spinal, Acute pain ↓ (Rizzi et al., 2015)	Spinal, Acute pain ↓ (Rizzi et al., 2015)
	Spinal, Neuropathic pain ↓ (Rizzi et al., 2015)	

***NOP Receptor Agonists (Non-peptides)***

Ro64-6198	Spinal, Neuropathic pain ↓ (Obara et al., 2005)	
	Systemic, Acute pain ↓ (Reiss et al., 2008)	Systemic, Acute pain ↓ (Ko et al., 2009)
	Systemic, Acute pain ↑ (Reiss et al., 2008)	Systemic, Inflammatory pain ↓ (Sukhtankar et al., 2014)
		Systemic, Capsaicin-induced allodynia ↓ (Ko et al., 2009)

Ro65-6570	Supraspinal, Neuropathic pain ↓ (Schiene et al., 2013)
	Systemic, Inflammatory pain ↓

NOP receptor-related ligands	Findings in rodents	Findings in primates
	(Schiene et al., 2013)	
	Supraspinal, Neuropathic pain ↓ (Schiene et al., 2013)	
GRT-TA2210	Supraspinal, Neuropathic pain ↓ (Linz et al., 2013)	
	Systemic, Inflammatory pain ↓ (Linz et al., 2013)	
SCH 221510	Systemic, Inflammatory pain ↓ (Sobczak et al., 2013) (Sobczak et al., 2014)	Systemic, Acute pain ↓ (Cremeans et al., 2012)
		Systemic, Inflammatory pain ↓ (Wladischkin et al., 2012)
		Systemic, Capsaicin-induced allodynia ↓ (Wladischkin et al., 2012)

***NOP Receptor Antagonist***

UFP-101	Supraspinal, Acute pain ↓ (Rizzi et al., 2006)
	Supraspinal, Inflammatory pain ↓ (Scoto et al., 2009)
	Supraspinal, Neuropathic pain ↓ (Scoto et al., 2009)

***Mixed NOP/MOP Receptor Agonists***

[Dmt <sup>1</sup> ]N/OFQ(1–13)-NH <sub>2</sub>	Spinal, Acute pain ↓ (Calo et al., 2012)	Spinal, Acute pain ↓ (Molinari et al., 2013)
--	---	---

SR16435	Spinal, Inflammatory pain ↓ (Sukhtankar et al., 2013)
	Spinal, Neuropathic pain ↓ (Sukhtankar et al., 2013)
	Systemic, Acute pain ↓ (Khroyan et al., 2009)

SR14150	Systemic, Neuropathic pain ↓ (Khroyan et al., 2011b)
---------	---

SR16835	Systemic, Neuropathic pain ↓ (Khroyan et al., 2011b)
---------	---

BU08028	Spinal, Inflammatory pain ↓ (Sukhtankar et al., 2013)	Systemic, Acute pain ↓ (Ding et al., 2015a)
	Spinal, Neuropathic pain ↓ (Sukhtankar et al., 2013)	Systemic, Capsaicin-induced allodynia ↓ (Ding et al., 2015a)
	Systemic, Acute pain ↓ (Khroyan et al., 2011a)	

Cebranopadol	Systemic, Acute pain ↓ (Linz et al., 2014)
	Systemic, Inflammatory pain ↓ (Linz et al., 2014)
	Systemic, Neuropathic pain ↓

<b>NOP receptor-related ligands</b>	<b>Findings in rodents</b>	<b>Findings in primates</b>
	(Linz et al., 2014)	

↓, antinociception or antihypersensitivity; ↑, pronociception or hypersensitivity

Extraído de (73) – Publicação aprovada pela revista (Número de licença: 4524260379553).

### 1.5.2.2 Ansiedade, estresse e depressão

Estudos indicam que os efeitos de N/OFQ são comparáveis aos produzidos por estressores, agindo diretamente sobre o eixo HPA e o SNC (77). Vários estudos apontam para um aumento do hormônio adrenocorticotrófico (ACTH) e corticosterona no plasma, após a administração i.c.v. de N/OFQ em ratos (122, 123). O aumento de corticosterona também foi detectado no plasma de ratos, após administração endovenosa do antagonista não peptídico, JTC-801, em condições de repouso (124). A administração por via i.c.v. do antagonista peptídico UFP-101 previniu o aumento da corticosterona induzido pela injeção central de N/OFQ (125). Estes estudos sugerem resultados controversos, mas os efeitos que prevalecem na literatura são de que antagonistas de NOPr previnem o efeito estimulatório da N/OFQ sob o eixo HPA (48).

Vários trabalhos demonstram, de forma consistente, que antagonistas do NOPr produzem melhora dos sintomas da depressão (81, 126). Okawa e cols. (2001) demonstraram a diminuição da liberação de noradrenalina do córtex pré-frontal do cérebro de ratos, após a injeção de 1,825 µg do agonista N/OFQNH<sub>2</sub> dentro do *locus coeruleus*, sendo parcialmente revertida por 138,16 µg do antagonista [Nphe1]N/OFQ(1-13)NH<sub>2</sub> (127). Além disso, ratos e camundongos com deleção gênica do NOPr tiveram redução no tempo de imobilidade no teste de nado forçado (107, 128-131). Contudo, Witkin e cols. não identificaram este mesmo efeito (132). Gavioli e cols. (2003, 2004) (129, 130) relataram que o bloqueio do NOPr pelo antagonista UFP-101, pela via i.c.v. (3-10 nmol/sítio), induz efeitos do tipo antidepressivo em camundongos e ratos. De acordo com Gavioli et al. (2003), a co-injeção com a N/OFQ, na dose de 1 nmol pela mesma via, levou a reversão dos efeitos do UFP-101 (3 e 10 nmol/sítio) (129). Em adição, o antagonista peptídico, UFP-101, apresentou efeito antidepressivo em camundongos nos testes do nado forçado e da suspensão da cauda, com a infusão de 5,72 µg, no hipocampo dorsal (133). A administração de UFP-101, pela via i.c.v., também resultou no aumento do consumo de sacarose por animais com estresse crônico moderado (134, 135). O antagonista SB-612111, pela via i.p., nas doses de 1-10 mg/kg, diminuiu o tempo de imobilidade no teste do nado forçado, sendo este efeito prevenido pela N/OFQ,

administrada centralmente na dose de 1 nmol (107). Importante ressaltar que Holanda et al. (2016) e Medeiros et al. (2015) também relataram efeitos antidepressivos para UFP-101 (3–10 nmol; i.c.v.) e SB-612111 (3–10 mg/kg; i.p.) (136, 137). Ademais, outros antagonistas do NOPr apresentaram ações antidepressivas em modelos de depressão, [Nphe1]-nociceptin (1–13)-NH<sub>2</sub> e J-113397 (138), LY2940094 (139), assim como agonistas parciais, UFP-113 e [F/G]N/OFQ(1–13)NH<sub>2</sub> (140). Por outro lado, os agonistas N/OFQ e Ro65-6570 do NOPr são capazes de prevenir efeitos do tipo-antidepressivo da nortriptilina e da fluoxetina, mas se apresentaram inativos *per se* em camundongos naïve (141).

Abaixo estão descritos as ações de antagonistas do NOPr sobre modelos de depressão (81) (Tabela 3).

**Tabela 3**

Effects of NOP receptor antagonists in preclinical models of depression

Assay	Compound	Species and strain	Effects	References
Forced swimming test	UFP-101	Swiss and NOP(−/−) mouse	↓ Immobility time; no effects in NOP(−/−) mice	Gavioli et al. (2003, 2004)
	UFP-101	Wistar rat	↓ Immobility time	Gavioli et al. (2004)
	[Nphe <sup>1</sup> ]N/OFQ(1–13)-NH <sub>2</sub>	CD-1 mouse	↓ Immobility time	Redrobe et al. (2002)
	UFP-113	Swiss mouse	↓ Immobility time	Asth et al. (2016)
	[F/G]N/OFQ(1–13)NH <sub>2</sub>	Swiss mouse	↓ Immobility time	Asth et al. (2016)
	J-113397	CD-, Swiss, and NOP(−/−) mouse	↓ Immobility time; no effects in NOP(−/−) mice	Redrobe et al. (2002) and Gavioli and Calo' (2006)
	SB-612111	Swiss and NOP(−/−) mouse	↓ Immobility time; no effects in NOP(−/−) mice	Rizzi et al. (2007)
	LY2940094	NIH-Swiss and NOP(−/−) mouse	↓ Immobility time; no effects in NOP(−/−) mice	Post et al. (2016) and Witkin et al. (2016)

Tail suspension test	UFP-101 SB-612111	Swiss mouse Swiss mouse	↓ Immobility time ↓ Immobility time	Gavioli et al. (2004) Rizzi et al. (2007)
DRL-72	LY2940094	SD rat	No effect	Witkin et al. (2016)
Chronic mild stress	UFP-101	Wistar rat	↑ Sucrose solution intake and ↓ immobility time after 21 days of treatment	Vitale et al. (2009, 2017)
Learned helplessness	SB-612111 and UFP-101	Swiss mouse	↑ Escapes and ↓ escape latencies	Holanda et al. (2016, 2018)
LPS-induced depressive-like behavior	SB-612111 and UFP-101	Swiss and CD-1 mouse	↓ Immobility time	Medeiros et al. (2015)

DRL differential reinforcement of low rate schedule, J-113397 1-[(3R,4R)-1-(cyclooctylmethyl)-3-(hydroxymethyl)-4-piperidinyl]-3-ethyl-1,3-dihydro-2H-benzimidazol-2-one, LPS bacterial lipopolysaccharide, LY2940094 [2-[4-[(2-chloro-4,4-difluoro-spiro[5Hthieno[2,3-c]pyran-7,4'-piperidine]-1'-yl)methyl]-3-methylpyrazol-1-yl]-3-pyridyl]methanol, SB-612111 (5S,7S)-7-[[4-(2,6-dichlorophenyl)-1-piperidinyl]methyl]-6,7,8,9-tetrahydro-1-methyl-5H-benzocyclohepten-5-ol, UFP-101 [Nphe<sup>1</sup>, Arg<sup>14</sup>, Lys<sup>15</sup>] N/OFQ-NH<sub>2</sub>

Extraído de (81) – Publicação aprovada pela revista (Número de licença: 4524260904007).

Com relação à ansiedade, a administração central de N/OFQ induziu o comportamento de ansiedade em ratos (142, 143). Contudo, esses dados são exceção à literatura e a maioria dos estudos apontam ausência de efeitos para os antagonistas de NOPr. Desta maneira, a administração central de UFP-101 (1-10 nmol), em ratos Wistar, reduziu a latênciam na esquiva inibitória no teste do labirinto em T elevado, indicando um efeito ansiolítico (144). Em adição, o antagonista, LY2940094, atenuou a imobilidade condicionada ao medo e a hipertermia induzidas pelo estresse (132). Mesmo com os dados que sugerem o impacto negativo do agonista natural N/OFQ sobre a ansiedade, a maior parte dos trabalhos se opõem a estes achados, demonstrando de forma consistente que a administração central de N/OFQ resultou na redução da ansiedade em roedores (140, 145-152). Assim, muitos ligantes não peptídicos do NOPr têm sido desenvolvidos como potenciais candidatos para o tratamento da ansiedade (55). Desta maneira, efeito ansiolítico foi verificado para agonistas em diferentes modelos de ansiedade e em diferentes espécies de roedores (81). Por exemplo, diferentes estudos revelaram um efeito do tipo-ansiolítico para os agonistas Ro

64-6198, Ro 65-6570, SCH 221510, SR-8993, AT-090 e Compostos 1, 1c e 3c, com aumento do tempo no braço aberto no labirinto em cruz elevado (96, 140, 153-158).

Abaixo estão listados os principais efeitos de agonistas do NOPr sobre modelos de ansiedade (81) (Tabela 4).

**Tabela 4**

Effects of non-peptide NOP receptor agonists in preclinical models of anxiety

Assay	Compound	Species and strain	Effects	References
Elevated plus-maze test	Ro 64-6198	Wistar and SD rat	↑ Time spent and distance moved in open arms	Jenck et al. (2000) and Dautzenberg et al. (2001)
	SCH 221510	CD-1 mouse	↑ Time spent in open arms	Varty et al. (2008)
	Compound 1c	Rat	↑ Time spent and distance moved in open arms	Wichmann et al. (1999)
	Compound 3c	Rat	↑ Time spent and distance moved in open arms	Wichmann et al. (2000)
	SCH 221510	Gerbil	↑ Time spent in open arms	Varty et al. (2008)
	Compound 1	Long-Evans and Hooded rat	↑ Time spent in open arms	Ross et al. (2015)
	Ro 65-6570	CD-1 mouse and NOP (-/-) mouse	↑ Time spent and number of entries in open arms; no effects in NOP(-/-) mice	Asth et al. (2016)
	SR-8993	Wistar rat	↑ Time spent in open arms in naive and after chronic alcohol consumption	Aziz et al. (2016)
Isolation-induced vocalizations	Ro 64-6198	CD-1 mouse	↓ Number and duration of vocalization	Varty et al. (2005)
	Ro 64-6198	Hartley guinea pig	↓ Number of vocalization	Varty et al. (2005)
	SCH 221510	Dunkin-Hartley guinea pig	↓ Separation-induced vocalizations in pups	Varty et al. (2008)
	Compounds 15 and 16	Hartley guinea pig	↓ Number of vocalization	Yang et al. (2009)
	SCH 655842	Dunkin-Hartley guinea pig	↓ Separation-induced vocalizations in pups	Lu et al. (2011)

Conditioned lick suppression	Ro 64-6198	CD-1 mouse	↑ Number of punished licks	Varty et al. (2005)
	SCH 221510	CD-1 mouse	↑ Number of punished licks	Varty et al. (2008)
	Compounds 15 and 16	Rat	↑ Number of punished licks	Yang et al. (2009)
	Compound 24	Rat	↑ Number of punished licks	Ho et al. (2009)
	SCH 655842	CD-1 mouse	↑ Number of punished licks	Lu et al. (2011)
	Ro 64-6198	Wistar rat	↑ Number of punished responses	Jenck et al. (2000)
	Ro 64-6198	SD rat	↑ Drinking time	Goeldner et al. (2012)
	SCH 221510	CD-1 mouse	↑ Number of punished licks	Varty et al. (2008)
	MCOPPB	ddY mouse	↑ Number of punished responses	Hirao et al. (2008b)
	PCPB	ddY mouse	↑ Number of punished responses	Hirao et al. (2008a)
Marble-burying test	Ro 64-6198	C57BL/6J mouse	↓ Marble-burying behavior	Nicolas et al. (2006)
	SCH 655842	C57BL/6J mouse	↓ Marble-burying behavior	Lu et al. (2011)
Ultrasound-induced defensive behaviors	Ro 64-6198	Lister-hooded rat	↓ Freezing behavior	Nicolas et al. (2007)
Fear-potentiated auditory startle	Ro 64-6198	Wistar rat	↓ Startle responses	Jenck et al. (2000)
Panic-like anxiety test	Ro 64-6198	Wistar rat	No effects	Jenck et al. (2000)
Open-field test	Ro 64-6198	Mouse	↑ Time spent in the center	Chang et al. (2015)
Social approach-avoidance	Ro 64-6198	Lewis rat	↑ Time spent in the social compartment	Goeldner et al. (2012)
Novelty-induced hypophagia	Ro 64-6198	C57BL/6J mouse	↓ Latency to drink and increase milk intake	Goeldner et al. (2012)
Stress-induced hyperthermia	Ro 64-6198	NMRI mouse	↓ Stress-induced hyperthermia	Goeldner et al. (2012)

*AT-090* 1-((1s,4s)-4-iso-propylcyclohexyl)piperidin-4-yl)indoline-2,3-dione, *MCOPPB* 1-[1-(1-methylcyclooctyl)-4-piperidinyl]-2-[(3R)-3-piperidinyl]-1H-benzimidazole, *PCPB* 2-(3,5-dimethylpiperazin-1-yl)-1-[1-(1-methylcyclooctyl)piperidin-4-yl]-1H-benzimidazole, *Ro 64-6198* (1S,3aS)-8-(2,3,3a,4,5,6-hexahydro-1H-phenalen-1-yl)-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one, *Ro 65-6570* (RS)-8-(1,2-dihydro-1-acenaphthylenyl)-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one, *SCH 221510* 3-endo-8-[bis(2-methylphenyl)methyl]-3-phenyl-8-azabicyclo[3.2.1]octan-3-ol, *SCH 655842* endo-8-[bis(2-chlorophenyl)methyl]-3-phenyl-8-azabicyclo[3.2.1]octane-3-carboxamide, *SR-8993* 1-[1-(cyclooctylmethyl)-4-piperidinyl]-5-fluoro-2-(3R)-3-pyrrolidinyl-1H-benzimidazole

Extraído de (81) – Publicação aprovada pela revista (Número de licença: 4524260904007).

### 1.5.2.3 Ligantes do NOPr na clínica

De maneira interessante, os ligantes do NOPr demonstraram eficácia em ensaios clínicos com pacientes (2, 78). Dentre estes estudos, potenciais para a utilização na clínica, destaca-se o antagonista LY2940094, um novo medicamento disponível na forma oral, para o tratamento da transtorno depressivo maior (139). Este antagonista também foi testado em pacientes com dependência alcóolica, reduzindo os dias de consumo pesado e aumentando os dias de abstinência (159). O agonista bifuncional cebranopadol foi conduzido em um ensaio clínico em fase II para o tratamento de pacientes com dor lombar crônica, revelando eficácia analgésica (160). Outro agonista, o SER100, em um tratamento por via s.c., induziu diminuição da pressão sanguínea diastólica e sistólica em pacientes com hipertensão sistólica isolada (161). Outros efeitos clínicos têm sido descritos para a N/OFQ, como ações benéficas para a incontinência urinária neurogênica. O agonista SCH486757 se mostrou efetivo na tosse subaguda, enquanto o JNJ-19385899 reduziu os sintomas de depressão e da ansiedade (78, 162). Além disto, foram descritos efeitos analgésicos para o antagonista, JTC-801, na dor neuropática e pós-operatória. Finalmente, o antagonista MK-5757 produziu melhora cognitiva dos sintomas esquizofrenia (78).

## 1.6 Fibromialgia

A fibromialgia é uma doença crônica, caracterizada principalmente por dor generalizada. Esta dor é iniciada por um estímulo habitualmente indolor, definindo assim, a alodínia presente na fibromialgia (163). A fibromialgia é uma dor disfuncional, ou seja, não é provocada por uma inflamação periférica, nem por uma lesão no sistema nervoso (164). Além disso, é uma dor presente mesmo sem estímulo, evocada por alta e baixa intensidade, sendo caracterizada por amplificação sensorial (164). Esta síndrome é acompanhada por sintomas como fadiga, dor musculoesquelética, distúrbios do sono, disfunções cognitivas e depressão (165-167). De acordo com a Associação

Internacional para o Estudo da Dor (IASP), a fibromialgia é definida como uma dor nociplástica, ou seja, relacionada com o processamento anormal da nocicepção (168).

Embora o diagnóstico da doença tenha embasamento no critério de classificação do Colégio Americano de Reumatologia (ACR), ainda permanece de difícil resolução (169). Os critérios utilizados em 1990 (ACR 1990) compreendiam: história de dor crônica generalizada (dor afetando os lados esquerdo e direito do corpo, acima e abaixo da cintura, e no esqueleto axial) e, pontos sensíveis à palpação digital (positivo para a doença quando  $\geq 11$  dos 18 sítios dolorosos) (170). No critério de 2010 (ACR 2010), os pontos sensíveis à palpação digital foram eliminados e um questionário com duas escalas foi adicionado: (i) o índice de dor generalizada e, (ii) o escore da gravidade dos sintomas (166). Este último critério sofreu modificações no ano de 2011 (ACR 2010 modificada), onde a estimativa de sintomas somáticos foi eliminada e as duas escalas foram expandidas (171). Em 2016, houve a revisão dos critérios de 2010/2011, sendo a fibromialgia diagnosticada de acordo com os seguintes critérios (1-4) (167):

- (1) Dor generalizada, definida como dor em pelo menos 4 das 5 regiões (região superior esquerda, região superior direita, região inferior esquerda, região inferior direita e região axial), está presente.
- (2) Os sintomas estão presentes em um nível semelhante há pelo menos 3 meses.
- (3) Índice de dor generalizada (WPI)  $\geq 7$  e escore de gravidade dos sintomas (SSS)  $\geq 5$  ou WPI de 4-6 e escore SSS  $\geq 9$ .
- (4) Um diagnóstico de fibromialgia é válido independentemente de outros diagnósticos. Um diagnóstico de fibromialgia não exclui a presença de outras doenças clinicamente importantes.

A prevalência mundial de fibromialgia é de 2,1%, com uma proporção de mulheres para homens de 4: 1 (172). Contudo, estes dados são de difícil caracterização, pois a maioria dos estudos epidemiológicos utiliza diferentes critérios de diagnóstico. Além disto, indivíduos com idade entre 40 e 60 anos são os mais afetados (173-175).

### **1.6.1 Diagnóstico da fibromialgia**

O diagnóstico da fibromialgia demonstra dificuldades, pois não existem achados laboratoriais que permitam a caracterização específica da síndrome. Contudo, estudos de neuroimagem mostram anormalidades no sistema nervoso de pacientes com fibromialgia, como estrutura cerebral alterada (176, 177), atividade metabólica (178) e conectividade funcional de estado de repouso (179), em regiões envolvidas no processamento da dor. Em adição, há um aumento na resposta a uma variedade de estímulos dolorosos (180-182). López-Solà et al. (183) verificaram a existência de uma “assinatura cerebral” em pacientes com fibromialgia, utilizando a ressonância magnética funcional. Nesse estudo, pacientes e grupo controle (indivíduos saudáveis) foram expostos a estímulos dolorosos (pressão) e indolores (táteis, auditivos e visuais) e foi demonstrado que os sujeitos fibromialgicos apresentaram respostas aumentadas nas regiões de integração sensorial (ínsula/opérculo) e auto-referência (por exemplo, pré-frontal medial) e respostas reduzidas no córtex frontal lateral. A ressonância magnética permite elucidar o que está acontecendo em nível cerebral nos pacientes com fibromialgia, refletindo a individualidade da sensação dolorosa inerente a estes sujeitos (183).

### **1.6.2 Fisiopatologia da doença**

A patogênese da doença não é clara, mas sabidamente, o processamento anormal da dor é influente na fibromialgia (184, 185). Em pacientes com a doença, uma pressão mecânica menor é necessária para desencadear a mesma atividade neuronal induzida por dor em relação a um indivíduo saudável. Em pacientes com fibromialgia, esta atividade também é maior (181). Isto se deve à sensibilização central, onde há uma resposta aumentada ao estímulo através da amplificação de sinal no SNC (185-187).

Além da sensibilização central, outras alterações fisiopatológicas têm sido sugeridas, incluindo uma influência genética no controle da dor, que pode ser modulada por fatores como ansiedade, depressão, estresse, trauma, adversidades na infância e/ou infecções. Fatores periféricos

também têm influência, como a dor contínua induzida por co-morbidades. Desequilíbrios de neurotransmissores e mudanças no HPA também podem ser detectados (188) (Figura 3).

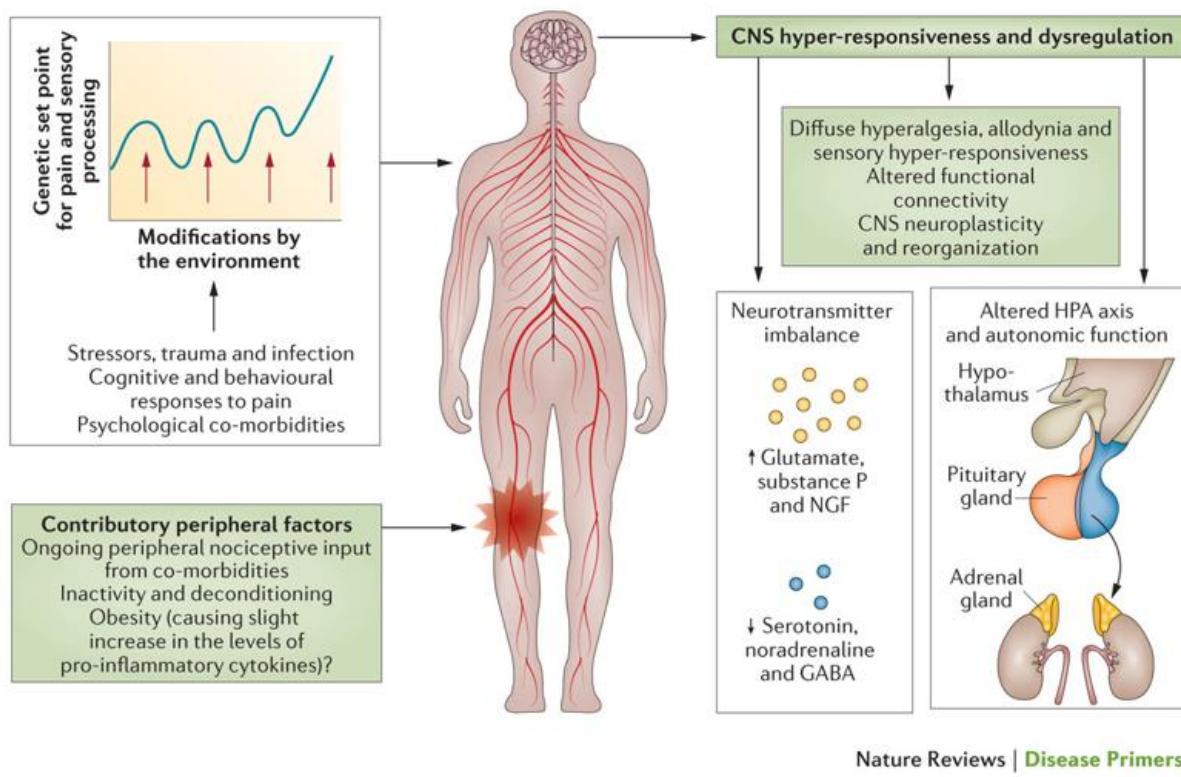


Figura 3: Potenciais processos fisiopatológicos na fibromialgia.

Extraído de (188) - Publicação aprovada pela revista (Número de licença: 3691360912654).

Na via ascendente da dor, os neurônios aferentes primários trazem os estímulos nociceptivos da periferia até o corno dorsal da medula espinhal, onde ocorre a sinapse com neurônios de segunda ordem que se projetam centralmente. Estes estímulos chegam ao hipotálamo e sua percepção ocorre no córtex somato-sensorial. Os neurotransmissores que facilitam a transmissão da dor são a substância P (SP), o glutamato e o fator de crescimento do nervo (NGF). Estes neuropeptídeos estão com os níveis aumentados no fluido cerebroespinal de pacientes com fibromialgia, estando implicados na excitabilidade dos neurônios da medula espinhal (189-192).

A inibição da transmissão da dor no corno dorsal ocorre através das vias descendentes: a medial, que tem origem na medula a partir de neurônios que contêm 5-HT, glutamato e ácido  $\gamma$ -aminobutírico (GABA) e, a via lateral, que se origina do tronco cerebral superior de neurônios que

contêm noradrenalina. No fluído cerebroespinhal de indivíduos com a síndrome, são encontrados níveis reduzidos de 5-HT e noradrenalina (189, 190, 193-195).

Os opioides endógenos também parecem estar envolvidos na fibromialgia, pois suas concentrações estão aumentadas no fluído cerebroespinhal de pacientes com fibromialgia (196). Além disto, estudos demonstram que pacientes com a doença apresentam diminuição da disponibilidade do receptor opioide MOP. Sugere-se que este receptor pode estar altamente ocupado por opioides endógenos, numa tentativa de reduzir a dor, ficando sua disponibilidade reduzida após a estimulação prolongada (197).

Lucas e cols. (198) sugeriram que mastócitos estão envolvidos na fibromialgia, e estas células estão aumentadas na camada papilar da derme de pacientes que tem a doença (199). Além disto, os mastócitos estão envolvidos na urticária crônica, condição frequente em indivíduos com fibromialgia (200). Os mastócitos secretam o peptídeo bradicinina, juntamente com o hormônio liberador de corticotrofina (CRH), histamina, interleucina-1 (IL-1), interleucina-6 (IL-6), prostaglandina D<sub>2</sub> (PGD<sub>2</sub>) e TNF, e estas moléculas podem ativar os nervos sensoriais periféricos diretamente ou, chegar ao cérebro através da circulação sistêmica, criando um circuito de dor auto-sustentável. Estas moléculas são liberadas em resposta ao estímulo dos mastócitos pelos peptídeos do estresse: CRH, NGF, neurotensina (NT) e SP (201) (Figura 4). O CRH encontra-se elevado no fluido cerebroespinhal de pacientes com fibromialgia e está associada com a dor (202).

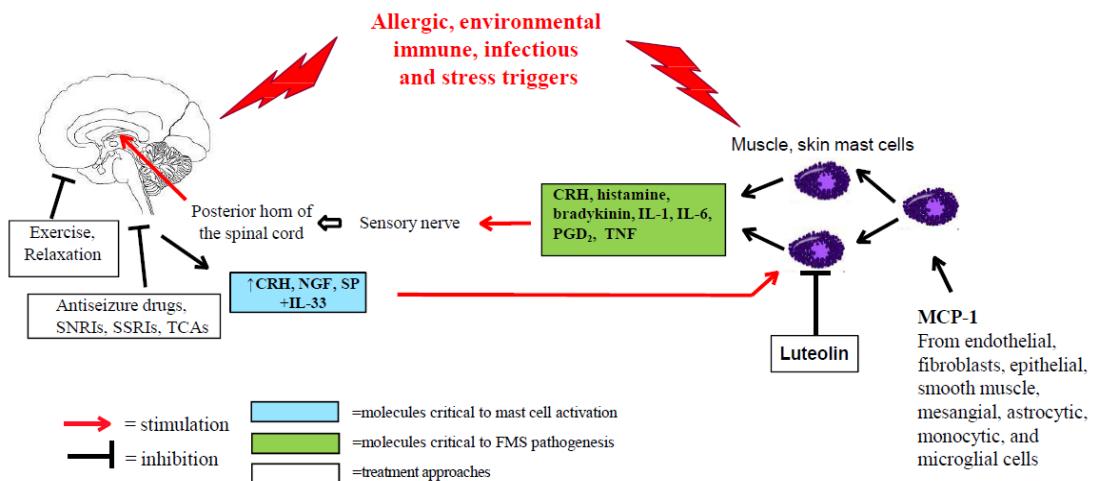


Figura 4: Representação dos mecanismos envolvidos na patogênese da fibromialgia e alvos para o tratamento.

Extraído de (2011) - *Publicação aprovada pela revista (carta de autorização para utilização do*

*doi:10.1124/jpet.115.227298).*

### 1.6.3 Fibromialgia e marcadores inflamatórios

Vários estudos sugerem uma relação entre a sintomatologia da fibromialgia e a ação de diferentes citocinas, pois estas atuam sobre o eixo HPA, linfócitos T e sobre o sistema nervoso simpático (203). As citocinas pró-inflamatórias estão envolvidas na amplificação da dor, através da sensibilização dos neurônios periféricos, aumentando as respostas ao óxido nítrico e à prostaglandina E<sub>2</sub> (PGE<sub>2</sub>). As células gliais ativadas pela SP, pelo glutamato e pelo fator neurotrrófico derivado do cérebro (BDNF) liberam citocinas pró-inflamatórias e vários neuropeptídeos, todos os quais podem contribuir para a amplificação da dor (204, 205) (Figura 5).

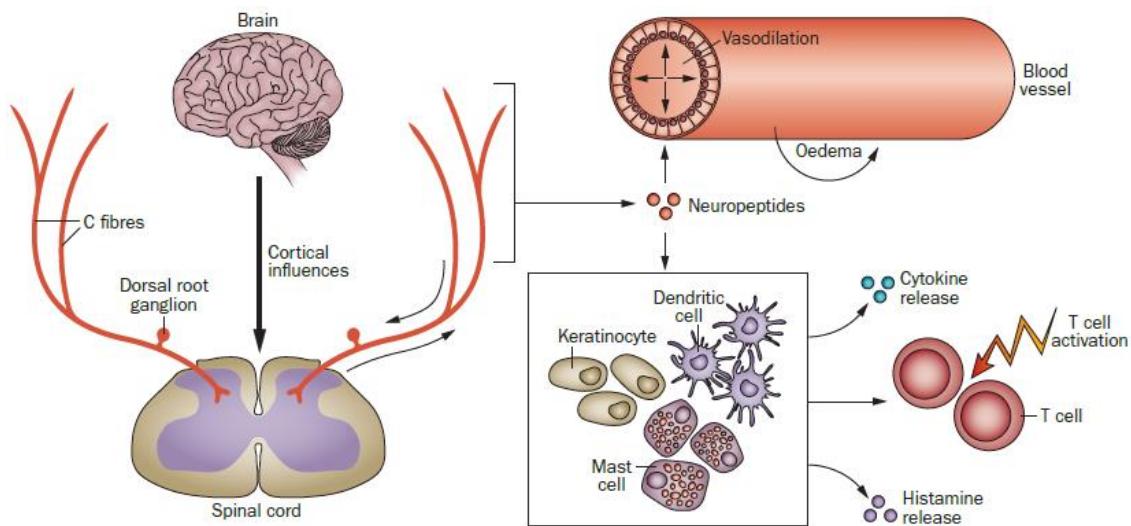


Figura 5: Efeitos centrais e periféricos associados com a liberação de neuropeptídos pelas fibras C terminais.

Extraído de (204) – Publicação aprovada pela revista (Número de licença: 3713771070231).

Sistemicamente, pacientes com fibromialgia, apresentam níveis elevados das citocinas pró-inflamatórias IL-6 e interleucina-8 (IL-8), níveis reduzidos ou inalterados das citocinas anti-inflamatórias interleucina-4 (IL-4) e interleucina-10 (IL-10) e concentrações normais de TNF. Além de níveis diminuídos da citocina pró-inflamatória IL-1 $\beta$  no plasma de pacientes fibromiálgicos. Contudo, muitos dos estudos nesta área apresentam problemas como amostras pequenas e qualidade metodológica baixa e, não levam em consideração que outras doenças (co-morbidades) podem influenciar no resultado da liberação de citocinas (206-210).

Com relação às citocinas pró-inflamatórias, foram detectados altos níveis de IL-1Ra (anticorpo do receptor de IL-1) e níveis inalterados de IL-1 $\beta$  e IL1 em pacientes com fibromialgia (211-213). Diferenças entre o grupo controle e indivíduos com a doença não foram encontradas para TNF (212, 213).

Outros estudos demonstram um aumento de TNF e IL-8 e IL-10 no plasma e soro de pacientes com fibromialgia (208, 211, 214). A concentração elevada de IL-8 no fluído cérebro-espinal medeia a dor simpática, através da ativação de células gliais (215). Quanto à concentração de IL-6, também há discordância entre os estudos, mas níveis aumentados desta interleucina têm sido associados com pacientes que têm a doença (206, 208, 211, 214, 216). Um estudo de

Pernambuco e cols. (217) mostrou níveis aumentados de interleucina-17A (IL-17A), interleucina-2 (IL-2), IL-4, TNF e IFN- $\gamma$  no plasma de pacientes com fibromialgia. A análise de amostras de pele de pacientes com a doença não demonstrou diferenças significativas nas concentrações de IL-10, quando comparadas com o grupo controle (218). Em adição à IL-10, outras citocinas anti-inflamatórias têm sido pouco estudadas, quando comparadas as citocinas pró-inflamatórias (207). Uma concentração reduzida de IL-4 foi encontrada no sangue total (219) e no plasma (220) de pacientes com fibromialgia, além da diminuição de outras citocinas de respostas do tipo Th2, como interleucina-5 (IL-5) e interleucina-13 (IL-13) (220). Yigit e cols. (221) observaram associação entre polimorfismo no gene da IL-4 e fibromialgia.

Alguns autores observaram o aumento de quimiocinas pró-inflamatórias em indivíduos com a síndrome, como altos níveis do ligante 2 de CC quimiocina (CCL2) e eotaxina no plasma (222) e uma concentração elevada de ligante 9 de quimiocina CXC (CXCL9), ligante 22 de CC quimiocina (CCL22), ligante 11 de quimiocina CXC (CXCL11), ligante 17 de CC quimiocina (CCL17) e eotaxina no soro (223). Um trabalho recente (2016), demonstrou que monócitos em repouso ou ativados de pacientes com fibromialgia secretam mais CCL11, CCL22 e CXCL1 quando comparados ao grupo controle (224).

#### **1.6.4 Tratamento**

Grupos na Europa, América do Norte e no Oriente Médio enfatizam que o tratamento da fibromialgia deve englobar terapias farmacológicas e não farmacológicas adaptadas a cada paciente (225). Além disto, por causa da heterogeneidade dos sintomas, um único tratamento não é eficaz para todos os indivíduos, sendo necessária, em alguns casos, uma combinação farmacológica (226). Dentre os tratamentos não farmacológicos, cabe citar a educação do paciente para a realização de atividades físicas e a terapia cognitivo-comportamental (TCC), que são benéficas em relação à dor e ao humor (227-229). A melhora na eficácia da TCC ocorre quando esta faz parte de um programa de tratamento multidisciplinar (educação, exercício e terapia psicológica). Nesse contexto, a

combinação de milnaciprano e TCC e a monoterapia com TCC foram igualmente benéficos para diminuir os sintomas da fibromialgia, sendo que o medicamento em pouco acrescentou no alívio dos sintomas (230, 231).

O antidepressivo tricíclico, amitriptilina, administrado em baixas doses, é efetivo para os sintomas da dor, distúrbios do sono e fadiga, mas tem efeitos limitados devido ao desenvolvimento de taquifilia e efeitos adversos anticolinérgicos e anti-histamínicos (232). A amitriptilina e a ciclobenzaprina apresentam efeitos como boca seca e constipação (233, 234). Outros antidepressivos, como os inibidores seletivos da recaptação de serotonina (SSRI), têm efeito positivo sobre a dor (235), mas medicamentos como a fluoxetina, a paroxetina e a sertralina podem ocasionar disfunção sexual, distúrbios do sono e náuseas (234). Já, os inibidores da recaptação de serotonina e noradrenalina (SNRI), a duloxetina e o milnaciprano, apresentam efeitos como dor de cabeça, náuseas, palpitações, hipertensão e taquicardia (234).

A venlafaxina é um medicamento que tem demonstrado efeitos positivos ou neutros para a fibromialgia; também é bem tolerado pelos pacientes e apresenta um baixo custo (236-239). Entretanto, estudos com o uso deste medicamento na depressão revelaram reações adversas, como insônia, tonturas, sonolência, prisão de ventre, sudorese e aumento ligeiro da pressão arterial (240-242).

Um dos fármacos mais prescritos para tratar casos mais resistentes é o tramadol, um agonista fraco do receptor MOP (243). Contudo, este medicamento apresenta efeitos adversos (hiperalgesia induzida por opioides e sedação), não tendo efeito benéfico sobre a qualidade de vida dos indivíduos, com eficácia moderada sobre a dor (244, 245). Os analgésicos anti-inflamatórios não esteroidais são frequentemente utilizados por pacientes com fibromialgia, mas não há evidências claras da sua eficácia (175, 246, 247).

Os sedativos benzodiazepínicos, assim como, os agentes hipnotícicos não benzodiazepínicos, são empregados para o tratamento de distúrbios do sono e ansiedade em pacientes com fibromialgia, mas o uso crônico pode ocasionar tolerância (248, 249). Estudos controlados não

conseguiram demonstrar efeitos benéficos suficientes do clonazepam para a sua utilização na fibromialgia (246, 250-252). Outra classe estudada para o tratamento da fibromialgia tem sido a dos agonistas dopaminérgicos, que melhoraram os sintomas da doença, como o pramipexol (253).

O oxibato de sódio (um sal de gama-hidroxibutirato; GHB) se mostrou eficaz para o controle dos sintomas da fibromialgia, mas apresentou problemas como vômitos, tontura, diarreia, dor de cabeça, ansiedade e sinusite (254). Por motivos de segurança (potencial droga de abuso) e por não apresentar resultados equivalentes aos medicamentos já aprovados para uso, o oxibato de sódio não foi aprovado pela Food and Drug Administration (FDA) e não é utilizado no Brasil para o tratamento da fibromialgia (249, 255, 256).

Os anticonvulsivantes também são utilizados no tratamento da fibromialgia, mas fármacos como a pregabalina apresentam pouco efeito sobre a fadiga (249). Os fármacos, gabapentina e pregabalina, apresentam baixa eficácia para a fibromialgia e ação limitada para a fadiga, a ansiedade e a depressão (257). Efeitos adversos importantes como sonolência, tontura, ganho de peso e edema são observados com estes medicamentos (258).

Em um estudo duplo-cego, a nabilona (canabinoide sintético) diminuiu significativamente a dor de pacientes com fibromialgia, após um tratamento de quatro semanas. Contudo, após oito semanas os seus efeitos desapareceram (259). Além disto, efeitos adversos como sonolência, boca seca e vertigem foram verificados (260). Outro estudo demonstrou diminuição dos distúrbios do sono, assim como efeitos adversos para este medicamento, incluindo, tonturas, náuseas, boca seca e sonolência (261). Lynch e cols. (262) descreveram que a utilização de canabinoides para indivíduos com fibromialgia apresentam efeitos modestos, porém seguros.

### **1.6.5 Fibromialgia e alterações musculares**

A fibromialgia é caracterizada por dor musculoesquelética, fadiga e perda de força muscular. Esses sintomas têm sido correlacionados com disfunções sensoriais e motoras, envolvendo alterações centrais e periféricas (263). Srikuea e colaboradores descreveram que pacientes com

fibromialgia apresentam maior variabilidade de tamanho e alteração na distribuição do tamanho das fibras musculares, quando comparados com indivíduos saudáveis (264). Alterações na morfologia mitocondrial dos músculos gastrocnêmio e sóleo têm sido investigadas no modelo de fibromialgia induzida por estresse, após indução ao frio intermitente (ICS) (265). Neste estudo, os autores demonstraram diminuição na área transversal das fibras musculares de camundongos machos submetidos ao modelo de fibromialgia por ICS. Além disto, também foi relatada a perda de fibras do tipo IIa no sóleo destes animais. No gastrocnêmio, houve aumento na densidade de células positivas para marcadores inflamatórios e atrogênicos, com a presença de mitocôndrias danificadas. Neste mesmo modelo de fibromialgia, Oezel e cols. (266) observaram aumento da atividade da enzima lactato desidrogenase (LDH) nos extratos mitocondriais dos músculos de camundongos induzidos por ICS.

### **1.6.6 Sistema nociceptina/orfanina FQ-NOPr na fibromialgia**

Poucos trabalhos têm investigado a relação entre o peptídeo N/OFQ e a fibromialgia. Anderberg e colaboradores (267) descreveram diminuição dos níveis plasmáticos de N/OFQ em mulheres fibromiálgicas, na fase lútea do ciclo menstrual, quando comparadas ao grupo controle. No trabalho de Baraniuk et al. (196), verificou-se que não houve diferença significativa nas concentrações de N/OFQ no fluido cérebro-espinal, em pacientes com fibromialgia, comparados com indivíduos controle ou, com diagnóstico de dor lombar crônica.

## 2 JUSTIFICATIVA

Vários estudos têm demonstrado atividades antinociceptivas para agonistas e antagonistas do NOPr, em modelos de dor aguda, inflamatória e neuropática, tanto em roedores quanto em primatas não-humanos. Ademais, os ligantes do NOPr apresentam efeitos sobre alterações comportamentais relacionadas com ansiedade e depressão em roedores. A fibromialgia, representa uma síndrome dolorosa, com co-morbidades, sendo que os pacientes apresentam quadros depressivo-ansiosos. Por outro lado, diversos efeitos adversos têm sido demonstrados pelos tratamentos existentes para controlar as alterações relacionadas à fibromialgia.

Com base no que foi descrito acima, justifica-se a relevância de investigar os possíveis efeitos farmacológicos exercidos por agonistas e antagonistas do NOPr em modelo de fibromialgia. Dessa forma, o presente estudo permitirá obter evidências sobre o papel do NOPr no modelo de fibromialgia induzido por reserpina em camundongos, a fim de identificar novas alternativas potenciais para a compreensão e tratamento desta doença, contribuindo para o avanço científico na área de Farmacologia e Terapêutica Experimental.

### **3 OBJETIVOS**

#### **3.1 Objetivo Geral**

O presente estudo teve como objetivo avaliar o efeito dos ligantes do NOPr em um modelo de fibromialgia em camundongos, bem como, investigar a plasticidade do sistema N/OFQ-NOPr nesse modelo.

#### **3.2 Objetivos Específicos**

- Investigar os efeitos do tratamento agudo com N/OFQ ou com o antagonista peptídico, UFP-101, administrados por diferentes vias, sobre a alodínia mecânica, nocicepção térmica e comportamentos do tipo depressivo-ansioso, em camundongos, no modelo de fibromialgia induzido por reserpina;
- Determinar os níveis N/OFQ em amostras de saliva de pacientes com fibromialgia e, em amostras de cérebro e soro, em animais submetidos ao modelo de fibromialgia;
- Investigar os efeitos do tratamento agudo com a naloxona (antagonista do receptor  $\mu$ ) em combinação com N/OFQ;
- Investigar os efeitos do tratamento agudo da combinação de N/OFQ, com o antagonista UFP-101;
- Investigar os efeitos do tratamento repetido com os antagonistas NOPr, UFP-101 e SB-612111, sobre a alodínia mecânica, nocicepção térmica, e comportamentos do tipo depressivo-ansioso e na fadiga, em camundongos, no modelo de fibromialgia induzido por reserpina;
- Avaliar os efeitos da reserpina e do bloqueio farmacológico do NOPr sobre o tamanho das fibras musculares através de avaliação histológica;

- Avaliar os efeitos da reserpina e dos tratamentos com os antagonistas NOPr sobre a morfologia mitocondrial do músculo, através de microscopia eletrônica de transmissão;
- Avaliar os níveis cerebrais e espinhais de 5-HT e glutamato após o tratamento repetido com o antagonista UFP-101, através de LC-MS/MS;
- Analisar a ativação cerebral, através do microPET/CT, após a indução de fibromialgia por reserpina, avaliando o tratamento repetido com o antagonista UFP-101;
- Avaliar a expressão da ppN/OFQ e do NOPr no cérebro, na medula espinhal e no músculo, através de PCR em tempo real, em camundongos, no modelo de fibromialgia induzido por reserpina;
- Avaliar a expressão do NOPr em amostras de cérebro, DRG, medula e músculo, através de imunoistoquímica em camundongos, no modelo de fibromialgia induzido por reserpina;
- Medir os níveis substância P em amostras de cérebro, medula e músculo, através de ELISA após o tratamento repetido com o antagonista UFP-101;
- Avaliar os efeitos dos tratamentos sobre os níveis de citocinas pró-infamatórias (TNF e IL-1 $\beta$ ) e da citocina anti-inflamatória IL-10, em amostras de cérebro, medula, músculo e soro, através de ELISA, após o tratamento repetido com o antagonista UFP-101;
- Determinar os níveis de glutationa em amostras de cérebro, medula e músculo, após o tratamento crônico com o antagonista UFP-101, em animais submetidos ao modelo de fibromialgia;
- Determinar a atividade da LDH em amostras de soro e músculo, após o tratamento repetido com o antagonista UFP-101.

#### **4 MANUSCRITO DO TRABALHO EXPERIMENTAL**

**Nociceptin/orphanin FQ receptor modulates painful and fatigue symptoms in a mouse model of fibromyalgia**

**Short running title:** NOP receptor and fibromyalgia

Ana Paula Aquistapase Dagnino<sup>1,2</sup>, Rodrigo Braccini Madeira da Silva<sup>3,\*</sup>, Pedro Cesar Chagastelles<sup>3,\*</sup>, Talita Carneiro Brandão Pereira<sup>1,4</sup>, Gianina Teribele Venturin<sup>5</sup>, Samuel Greggio<sup>5,6</sup>, Jaderson Costa da Costa<sup>5</sup>, Maurício Reis Bogo<sup>1,3,4</sup>, Maria Martha Campos<sup>1,2,3,7</sup>

\*These authors contributed equally to this study.

<sup>1</sup>Escola de Ciências, Programa de Pós-Graduação em Biologia Celular e Molecular, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90619-900, Brazil.

<sup>2</sup>Escola de Ciências da Saúde, Centro de Pesquisa em Toxicologia e Farmacologia, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Avenida Ipiranga, 6681, Porto Alegre, RS 90619-900, Brazil.

<sup>3</sup>Escola de Medicina, Programa de Pós-Graduação em Medicina e Ciências da Saúde, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90619-900, Brazil.

<sup>4</sup>Escola de Ciências, Laboratório de Biologia Genômica e Molecular, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90610-000, Brazil.

<sup>5</sup>Centro de Pesquisa Pré-Clínica, Instituto do Cérebro do Rio Grande do Sul – Brain Institute (BraIns), Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90610-000, Brazil.

<sup>6</sup>Escola de Ciências da Saúde, Curso de Graduação em Biomedicina, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90619-900, Brazil.

<sup>7</sup>Escola de Ciências da Saúde, Programa de Pós-Graduação em Odontologia, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Porto Alegre 90619-900, Brazil.

**Correspondence:** Maria M. Campos, Escola de Ciências da Saúde, Pontifícia Universidade Católica do Rio Grande do Sul, Avenida Ipiranga, 6681, Partenon, Porto Alegre, RS 90619-900, Brazil. Telephone number: +555133203677 E-mail: camposmmartha@yahoo.com; [maria.campos@pucrs.br](mailto:maria.campos@pucrs.br)

URL: <http://www.pucrs.br/>

## ABSTRACT

Generalized pain and fatigue are both hallmarks of fibromyalgia, a syndrome with an indefinite aetiology. The treatment options for fibromyalgia are currently limited, probably due to its intricate pathophysiology. Thus, further basic and clinical research on this condition is currently needed. This study investigated the effects of nociceptin/orphanin FQ (N/OFQ) receptor (NOPr) ligands and the modulation of the NOP system in the preclinical mouse model of reserpine-induced fibromyalgia. The effects of administration of the natural agonist N/OFQ and the selective NOPr antagonists (UFP-101 and SB-612111) were evaluated in fibromyalgia-related symptoms in reserpine-treated mice. The expression of preproN/OFQ (ppN/OFQ) and NOPr was assessed in central and peripheral sites at different time-points after reserpine administration. N/OFQ displayed dual effects in the behavioural changes in the reserpine-elicited fibromyalgia model. The peptide NOPr antagonist UFP-101 produced analgesic and anti-fatigue effects, by preventing alterations of brain activity and skeletal muscle metabolism, secondary to fibromyalgia induction. The non-peptide NOPr antagonist SB-612111 mirrored the favourable effects of UFP-101 in painful and fatigue alterations induced by reserpine. A time-related up- or down-regulation of ppN/OFQ and NOPr was observed in supraspinal, spinal and peripheral sites of reserpine-treated mice. Our data shed new lights on the mechanisms underlying the fibromyalgia pathogenesis, supporting a role for N/OFQ-NOP receptor system in this syndrome.

## Introduction

The worldwide prevalence of fibromyalgia is 2.1%, with a female to male ratio of 4:1 [13]. Patients with a fibromyalgia diagnosis display widespread pain, usually evoked by painless stimuli, thus defining the allodynia present in this syndrome [70]. It is accompanied by comorbidities such as fatigue, sleep disturbances, cognitive dysfunctions and depression [83]. A recent note by the International Association for the Study of Pain (IASP) described fibromyalgia as a nociceptive pain, i.e. “pain that arises from altered nociception” [6]. The fibromyalgia pathogenesis is not well defined, but abnormal pain processing is patently present [18]. Besides the involvement of monoamine deficits, the symptoms of fibromyalgia likely rely on the release of neuropeptides and cytokines, which causes central and peripheral alterations [46].

Nociceptin/orphanin FQ (N/OFQ) is a 17-amino acid peptide that exerts its biological actions by activating the opioid-related G protein-coupled nociceptin/orphanin FQ receptor (NOPr) [2]. The effects of NOPr ligands have been characterized in pain, depression and anxiety, among a series of pathophysiological alterations [17,25,39,44,61,65,82,86,87]. Regarding the painful responses, NOPr agonists can elicit either nociception or antinociception, depending on the experimental paradigm [39]. The actions of NOPr ligands have been examined in a series of rodent models of long-lasting pain, as reviewed by Kiguchi et al., 2016 [39]. Notably, chronic neuropathic pain states have been correlated with a plasticity of the N/OFQ-NOPr system. The mRNA levels of the N/OFQ precursor prepronociceptin and the NOPr were decreased in the thalamus and hypothalamus [59], whereas the NOPr mRNA expression was increased in the dorsal root ganglia (DRG) and spinal cord [12] of mice submitted to the chronic constriction injury (CCI) model. In rats, the CCI led to an upregulation of N/OFQ levels in DRG, periaqueductal gray (PAG) and rostral ventromedial medulla (RVM) [16]. Additionally, the spinal nerve ligation induced a decrement of NOPr immunoreactivity in the ipsilateral L4-L5 spinal cord and dorsal root ganglion (DRG) of mice [58].

Despite the substantial evidence showing a relevant role for the N/OFQ-NOPr system in neuropathic pain, only a few studies examined its participation in fibromyalgia. Anderberg and cols. [3] described a decrease of N/OFQ plasma levels in fibromyalgic women in the luteal phase of the menstrual cycle, when compared to controls. Otherwise, Baraniuk and cols. [7] did not identify any significant difference of N/OFQ concentrations in the cerebrospinal fluid of patients with fibromyalgia, in relation to control health subjects or individuals with chronic low back pain. Accordingly, the literature data regarding the role of the NOP system in fibromyalgia is controversial, indicating the needing of additional studies. Considering the previous evidence showing a relevant participation of the NOP system in fibromyalgia-related symptoms and

comorbidities (such as pain, depression and anxiety), we judged relevant to perform additional investigations on this matter. To our knowledge, no previous study assessed the effects of NOP ligands and/or the plasticity of this system in experimental fibromyalgia, justifying the premise of the study.

The biogenic amine depletion induced by reserpine has been firstly validated in rats and after in mice, as an animal model of fibromyalgia. In this experimental paradigm, the repeated administration of reserpine to rodents leads to fibromyalgia-related painful and depression changes (face validity), leading to reduced levels of neurotransmitters (construct validity), being sensitive to the treatment with pregabalin, duloxetine and pramipexole (predictive validity) [5,8,11,21,41,43,54,55]. The present study evaluated the site-specific modulatory effects of NOP<sub>r</sub> ligands in a mouse model of reserpine-induced fibromyalgia, investigating the plasticity of the N/OFQ-NOP<sub>r</sub> system at the supraspinal, spinal and peripheral sites.

## Methods

### *Animals*

The experimental procedures followed the current Brazilian guidelines for the care and use of animals for scientific and didactic procedures, from the National Council for the Control of Animal Experimentation (CONCEA, Brazil, 2014). The local Animal Ethics Committee evaluated and approved all of the protocols (CEUA 15/00487). The animal studies are reported in compliance with the ARRIVE guidelines [40,49].

Female CF-1 specific-pathogen-free mice (2-month old, 20-24 g,  $N = 596$ ) were obtained from the Centre for Experimental Biological Models (CeMBE, PUCRS, Porto Alegre, RS, Brazil). Reserpine leads to fibromyalgia-related symptoms with similar effects in female and male mice [33]. The experimental  $n$  was determined based on previous literature data [43]. The  $n$  per group is indicated within the figures. The dose-response experiments were independently replicated three to four times, explaining the variations of the  $n$ . We initiated with the systemic administration of NOP ligands, by i.p. route, with the dose of 3 nmol/kg (for N/OFQ and UFP-101), including the appropriate negative and positive controls. To complete the dose-response curves, we tested additional doses of NOP ligands, but we included more animals treated with the 3-nmol/kg dose, besides the negative and positive control groups, for purposes of comparison. For this reason, the  $n$  presents such a great variation in this experimental set. We performed an analysis to determine the power of a completed experiment (GraphPad StatMate 2.00; by GraphPad Software Inc.), considering a significance level (alpha=0.05; two-tailed). The analysis revealed power values ranging from 80% to 95% for this experimental set.

The mice were kept in micro-isolator cages (4 per cage), equipped with inlet/outlet air filters, under controlled temperature ( $22 \pm 1^\circ\text{C}$ ) and humidity (50 - 70%), and a light-dark cycle of 12 h (lights on at 7 a.m., lights off at 7 p.m.). The cages were filled with autoclaved wood chip bedding. The animals received pelleted feed and sterile water *ad libitum*. During the experimental procedures, the laboratory temperature was maintained at  $22 \pm 1^\circ\text{C}$ . A period of adaptation to the new environment of at least one hour was used. All the experiments were performed between 7 a.m. and 7 p.m.

The animals were allocated into the experimental groups (with or without fibromyalgia induction) considering their basal responses to Von Frey stimulation before fibromyalgia induction. The mice were treated and assessed in the behavioural tests in the following order: vehicle/saline (negative control group), reserpine/saline (positive control group), mice treated with reserpine/NOPr ligands, one mice from each group per turn. When possible, the collection of samples for biochemical, molecular and histochemical studies was performed after the behavioural

assessments, to reduce the total number of animals included in the study. The investigators were blinded to the experimental groups in either *in vivo* or *ex vivo* assessments. The animals were euthanized by sevoflurane inhalation.

### **Drugs**

Reserpine and naloxone were purchased from Sigma Aldrich Chemical Company (St. Louis MO, USA); N/OFQ and UFP-101 were purchased from Tocris Bioscience (Bristol, UK). Pregabalin (Lyrica) was obtained from Pfizer (Tadworth, UK). SB-612111 was obtained from Santa Cruz Biotechnology (Dallas, Texas, USA). N/OFQ and UFP-101 (1 mM) were diluted in saline solution (0.9% NaCl) and stored at - 20°C. SB-612111 (10 mM) was diluted in 1.2% dimethyl sulfoxide (DMSO) in saline solution (V/V), and stored at - 20°C. Reserpine was dissolved in 0.5% vehicle solution in distilled water (V/V). Pregabalin and naloxone were diluted in saline solution (0.9% NaCl). Reserpine, pregabalin and naloxone were diluted immediately before use. The NOPr ligands were prepared at the desired concentration just before treatments.

### **Reserpine-induced fibromyalgia**

The fibromyalgia mouse model was accomplished as described before [43]. Briefly, an amine depletion was induced by reserpine administration via subcutaneous route (s.c.; 0.25 mg/kg), once a day, for three consecutive days. The control group received the vehicle (0.5% acetic acid). The behavioural tests were carried out on the fourth day (Supplementary Figure 1A). The samples were collected from one to four days after the onset of the induction protocol, depending on the analysis.

### **Dose-related effects of NOPr ligands by different routes of administration**

This part of the study evaluated the acute dose-dependent effects of N/OFQ (NOPr agonist) and UFP-101 (selective peptide NOPr antagonist) on painful-, depression- and anxiety-like changes, in the mouse model of reserpine-induced fibromyalgia. To assess the sites of action of NOPr ligands, N/OFQ or UFP-101 were administered by the intracerebroventricular (i.c.v.), intrathecal (i.t.) or intraperitoneal (i.p.) routes, 15 min (i.c.v. and i.t) or 30 min (i.p.) before the experimental sessions, on the fourth day after the onset of the fibromyalgia induction protocol (Supplementary Figure 1A). Mice were evaluated in behavioural tests, for assessing mechanical and thermal hypersensitivity, depression-related immobility time, and anxiety parameters, as described in the next sections. The *n* for this experimental set was 303 animals (Supplementary Figure 1B).

The doses of N/OFQ and UFP-101 were determined based on prior publications [15,51,56,63,85]. For N/OFQ, the doses were 1 nmol/site (i.c.v.); 0.3, 1 and 3 nmol/site (i.t.); and 0.3, 1, 3 and 5 nmol/kg (i.p.). The doses of UFP-101 were 0.3 and 1 nmol/site (i.c.v.); 1, 3 and 5 nmol/site (i.t.); and 0.3, 1, 3 and 5 nmol/kg (i.p.). An additional dose of N/OFQ was tested by i.c.v. route (3 nmol/site), but the animals presented unspecific central side effects, such as shivering and shaking ( $n = 3$ ). The animals in this group were euthanized, and they were excluded from the study.

For the i.t. injections, a volume of five  $\mu\text{l}$  of saline containing N/OFQ or UFP-101 was injected between the L5 and L6 vertebral spaces. For the i.c.v. injections, a volume of two  $\mu\text{l}$  of saline containing N/OFQ or UFP-101 was injected directly into the lateral ventricle (coordinates from bregma: 1 mm lateral, 1 mm rostral, 3 mm vertical), of animals slightly anaesthetized with sevoflurane (3%) and oxygen (97%). The control groups received the corresponding volumes of vehicle (0.9% NaCl solution) [43].

#### ***Assessment of N/OFQ selectivity and site of action of UFP-101***

To exclude the participation of opioid receptors in the effects of N/OFQ, the animals were pre-treated with the MOP/DOP/KOP antagonist naloxone (5  $\mu\text{mol}/\text{kg}$ ; i.p.) [74], administered 5 min prior to N/OFQ treatment. To assess whether UFP-101 might revert the N/OFQ effects, separate groups of animals received (i) UFP-101 (1 nmol/kg; i.p.) plus N/OFQ (1 nmol/kg or 5 nmol/kg; i.p.; co-treatment); (ii) UFP-101 (1 nmol/kg; i.p.; 15 min prior) plus N/OFQ (1 nmol/site; i.c.v.); or (iii) UFP-101 (1 nmol/kg; i.p.; 15 min prior) plus N/OFQ (1 nmol/site; i.t.). The doses of NOPr ligands were chosen from the dose-response experiments. The behavioural changes were evaluated at the fourth day after the beginning of the protocol for induction of fibromyalgia. One hundred and seven animals were used in this experimental set.

#### ***Repeated treatment with NOPr antagonists***

Based on the dose-response experiments, the effects of the selective peptide NOPr antagonist UFP-101 (1 nmol/kg; i.p.) were evaluated in a protocol of repeated administration. The animals received UFP-101, for three consecutive days, 30 min after the daily reserpine injection. On the fourth day, the animals also received UFP-101, 30 min before the behavioural evaluation. Control animals received saline (0.9% NaCl, 10 ml/kg; i.p.). Pregabalin, an inhibitor of the  $\alpha2\delta$  subunit of voltage-gated calcium channels, was used as a positive control drug (188  $\mu\text{mol}/\text{kg}$ , i.p.), and it was administered at the same schedule of treatment. The pregabalin dose was selected based on a previous publication [71]. Mice were assessed for mechanical and thermal hypersensitivity,

depression-related immobility time, and anxiety parameters. The animals were also submitted to additional tests to analyse fatigue-associated symptoms (*n* for this experimental set of 76 mice).

The effects of the repeated treatment with the selective non-peptide NOPr antagonist SB-612111 (6.6 µmol/kg, i.p.) were also investigated. The antagonist or the vehicle was dosed, for three consecutive days, 30 min after the reserpine injection. On the fourth day, the animals also received SB-612111, 30 min before the behavioural testing. Separate experiments were performed to test the effects of different doses of SB-612111 (2.2, 6.6 and 22 µmol/kg) on the fatigue symptoms evoked by reserpine. The doses of SB-612111 were selected from previous studies [48,62,85]. An experimental *n* of 56 animals was used for this part of the study. A general presentation of the repeated schedule of treatment and the behavioural tests for this experimental set is depicted in the Supplementary Figure 1B, C, D.

## **Behavioural tests**

### ***Mechanical hypersensitivity***

The animals were placed in individual Plexiglas compartments on a metal screen. An adaptation period of 60 min before testing was used. The mechanical allodynia was evaluated using a 0.4-g Von Frey hair filament. The results were expressed as the withdrawal response frequency (%) [24]. The filament was applied ten times to the plantar surface of the right hind paw, with three seconds between each application. The withdrawal response frequency was evaluated before (baseline records), and at the fourth day after the onset of fibromyalgia induction. A significant increase in the response frequency compared to the baseline was considered as an indicative of mechanical hypersensitivity.

### ***Hot-plate test***

The thermal hypersensitivity following heat stimulation was assessed in the hot-plate apparatus (Ugo Basile, Italy), as described previously [42]. The surface of the hot-plate was heated at a constant temperature of  $50 \pm 1^{\circ}\text{C}$ . After the appropriate treatments, the animals were placed in the apparatus, which consists of a metal plate surrounded by a transparent acrylic cylinder. The latency to respond to heat stimulus (hind paw licking, withdrawal of the hind paw, or a jump) was measured before (baseline records) and at the fourth day after initiating the induction of fibromyalgia. The tests were finalised if the animals did not respond within 30 s, to avoid tissue damage.

### ***Forced swimming test***

The experiments were performed using a cylinder (18.5 cm in diameter and 25 cm in height) filled with water at a height of 17 cm. The water was maintained at 23-25°C. The animals were placed in the water and the immobility was defined as the absence of any movements, except those necessary to keep the mouse's head above the water [47]. The time that the mice remained immobile was quantified over a period of 6 min, at the fourth day after initiating the protocol of reserpine treatment, and it was used as an indication of depressive-like behaviour.

#### ***Elevated plus maze***

The mice were placed in the intersection of the four arms of an elevated plus maze and their behaviour was recorded for 5 min [78]. The parameters recorded were the total number of entries, the number of entries in the open arms, and the percent of time spent in the open arms.

#### ***Fatigue evaluation on the rotarod***

A rotarod apparatus (Insight, Ribeirão Preto, Brazil) was used to assess the fatigue-like symptoms in reserpine-treated mice, in the protocols of repeated treatment with the NOPr antagonists. The mice were trained three times, for one min, at 20 rpm, on the day before the first reserpine injection (baseline records). At the fourth day, the mice were submitted to two exercise sections (1 min each) and afterwards, the fatigue analysis was assessed at a speed of 20 rpm. The duration for which the mouse remained on the rod was recorded. A 60-min cut-off time was used [80].

#### ***Grasping strength measurement***

The mice were submitted to the grasping test, as an indication of the grip strength [19]. For this purpose, the animals were repeatedly treated with UFP-101 or SB-612111 as described before. After 30 min of the last treatment, they were lifted by the tail and allowed for grasping a grid connected to an electronic balance. When the first signs of active finger flexion were noted, the grasping strength was registered (in grams). The baseline records were acquired before the first reserpine injection. On this occasion, the mice were submitted to three training assessments. On the fourth day, the mean of three readings was used to calculate the individual grasping strength.

#### ***Kondziela's inverted screen test***

The Kondziela's inverted screen test [20] was performed on day 1 (baseline records) and on day 4, in the groups submitted to the repeated treatment with NOPr antagonists. The mice were placed in the centre of the wire mesh screen and the apparatus was inverted. The time elapsed before the mouse fell from the screen was recorded. A one min cut-off time was used.

### ***Determination of neurotransmitters by LC-MS/MS***

The levels of serotonin (5-HT) and glutamate were analysed in thalamus/hypothalamus, prefrontal cortex and lumbar spinal cord of animals submitted to the repeated treatment with UFP-101 (1 nmol/kg; i.p) in the mouse model of reserpine-induced fibromyalgia, according to the method described by [43]. Tissues were collected on the fourth experimental day. The results were expressed as ng/g tissue.

### ***MicroPET imaging***

These experiments were performed as described previously [67], to assess the supraspinal changes related to fibromyalgia induction, as well as, the effects of NOPr modulation on brain activity. Saline or UFP-101 (1 nmol/kg; i.p) were repeatedly administered for three consecutive days, 30 min after the daily reserpine injection, and 10 minutes after [<sup>18</sup>F]-FDG (250 µCi, i.p.), at the fourth day. After [<sup>18</sup>F]-FDG administration, the animal remained isolated and conscious for 40 min (uptake). For the scanning in Triumph microPET (LabPET-4, TriFoil Imaging, Northridge, CA, USA), the rodent was anesthetized with inhalatory isoflurane and medical oxygen (3-4% induction and 2-3% maintenance dose) and placed in a supine position in the imaging chamber, maintained at a constant temperature of 36°C. The animals were scanned for 10 min, with the brain region positioned in the centre of the microPET field-of-view (FOV). At the end of the scan, the mouse was removed from the device, and kept on a heating surface until complete recovery. The animals were scanned on the fourth day after initiating the protocol for fibromyalgia induction. The reconstruction algorithm for image processing was the MLEM-3D, and the capture of brain [<sup>18</sup>F]-FDG was quantified through the PMOD v3.5 software and Fusion Toolbox (PMOD Technologies, Zurich, Switzerland). Each mouse's uptake of [<sup>18</sup>F]-FDG was normalised to a reference brain region, avoiding unwanted sources of variation related to differences in mouse body weights. Glucose uptake in all brain regions was normalized by the cerebellum and expressed as relative standardized uptake value (SUV<sub>r</sub>) [84].

### ***Dorsal root ganglion (DRG) isolation***

This protocol was accomplished as described before [68], with minor modifications. The spinal cords of vehicle- and reserpine-treated mice were isolated on the fourth experimental day; the surrounding muscles, fat, spinal nerves and other soft tissues were removed. To avoid the damage of DRG, transverse cuts were made through the vertebrae between the discs. Each DRG was collected individually, by using a fine tip scissor. The spinal cord was maintained on ice thoroughly. Once dissected and cleaned, the DRGs of lumbar spinal cord were collected, and fixed in 10%-buffered formalin solution until the immunohistochemistry analysis.

### ***Histological analysis of skeletal muscle***

To assess the effects of UFP-101 on fibromyalgia-related skeletal muscle changes, the masseter, the gastrocnemius and the soleus were collected on the fourth experimental day. The samples were fixed in 10%-buffered formalin solution for 24 h, and embedded in paraffin after dehydration. The histological analysis was performed using haematoxylin-eosin (H&E) staining. The cross-sectional area (CSA) and the frequency (%) of fibres with different diameters were determined using the NIH Image J 1.36b Software. The slides were acquired with a Zeiss AxioImager M2 light microscope (Carl Zeiss, Gottingen, Germany). The images were captured at  $\times 200$  magnification.

### ***Transmission electron microscopy (TEM) for mitochondria evaluation***

Masseter muscles were collected from vehicle and reserpine treated-mice at fourth day, and a mitochondrial analysis was performed according to the method described previously [45]. The samples were cut into small pieces and fixed in a mixture of 4% paraformaldehyde and 2.5% glutaraldehyde, buffered with 0.1 M phosphate (pH 7.3), at room temperature. Then, the samples were post-fixed in osmium tetroxide in the same buffer for 45 min. The dehydration was performed in a graded acetone series (30–100%) and embedded in araldite (Durcupan<sup>TM</sup> ACM, Fluka), for 72 h, at 60°C. Thin sections (70 nm) were stained with 2% uranyl acetate, followed by an immersion in lead citrate. Ultrastructural analysis was performed using the Tecnai G2 T20, FEI transmission electron microscopy and Image Pro Plus software (Image Pro Plus 6.1, Media Cybernetics, Silver Spring, USA). The mitochondrial density and area were analysed as described by [45], with minor modifications. Squares measuring  $27.27 \mu\text{m}^2$  were considered as the full microscopic image of each analysed muscle area (field), and mitochondria located inside each square or intersected by the upper and/or right edges of the squares were counted. Mitochondria intersected by the lower and/or left edges of the squares were not considered. Six to ten muscle regions (field) were analysed per

mouse, with an original magnification  $\times$  8,900. A grid mask (a grid of crosses with equidistant intervals) with an area/point value of  $0.0311418 \mu\text{m}^2$  was placed over the mitochondria images.

### ***ppN/OFQ and NOPr RT-qPCR analysis***

Thalamus/hypothalamus, prefrontal cortex, lumbar spinal cord and masseter muscle were isolated on days 1, 2, or 3 after reserpine induction. The structures were stored in 300  $\mu\text{l}$  of TRIzol Reagent® (Sigma, St. Louis, MO, USA). The total RNA isolated was then quantified by spectrophotometry and the cDNA was synthesized with ImProm-II™ Reverse Transcription System (Promega, Madison, WI, USA), in accordance with manufacturer's instructions, after DNase I treatment (DNase I Amplification Grade; Sigma-Aldrich, EUA). Quantitative PCR was performed using SYBR® Green I (Invitrogen, Carlsbad, CA, USA). Reactions were performed in a volume of 25  $\mu\text{l}$  using 12.5  $\mu\text{l}$  of diluted cDNA (1:50) and 200 nM of each reverse and forward primers (ppN/OFQ-F 5'-AGCACCTGAAGAGAATGCCG-3'; ppN/OFQ-R 5'-CATCTCGCACTTGCACCAAG-3'; NOPr-F 5'-ATGACTAGGGCGTGGACCTGC-3'; NOPr-R 5'-GATGGGCTCTGTGGACTGACA-3' [31]. PCR cycling conditions followed an initial 5 min at 95°C polymerase activation step, plus 40 cycles of 15 s at 95°C for denaturation, 35 s at 60°C for annealing and 15 s at 72°C for elongation. Finally, a melting curve analysis was included with fluorescence measures from 60 to 99°C. The Cq values were obtained with 7500 Fast Real-Time PCR System v.2.0.6 (Applied Biosystems, Carlsbad, CA, USA) and relative expression levels were determined using *ppia* and *hpert* as reference genes [60] through  $2^{-\Delta\Delta\text{Cq}}$  method. The efficiency per sample was calculated using LinRegPCR 11.0 software (<http://LinRegPCR.nl>).

### ***Immunohistochemistry***

The expression of NOPr was evaluated by immunohistochemistry, according to the method described previously [67]. The brain, the lumbar spinal cord (L1-L6), the masseter muscle and the DRGs were collected on the fourth day, from reserpine- or vehicle-treated mice, and fixed in 10%-buffered formalin solution, for 24 h. The immunopositivity to NOPr was assessed in paraffin-embedded tissue sections (4  $\mu\text{m}$ ), using a polyclonal rabbit anti-NOPr antibody (1:400; Alomone, Jerusalem, Israel, Catalog Number AOR-015). Images were examined with a Zeiss AxioImager M2 light microscope (Carl Zeiss, Gottingen, Germany). For analysis, the images were captured in  $\times$  100 (brain, spinal cord and masseter), in  $\times$  200 (DRG) or in  $\times$  400 (masseter) magnification. The schematic representations of brain and spinal cord were captured in  $\times$  8 and  $\times$  32 magnification, respectively (ZEISS Stemi DV4 Stereo Microscope). The number of NOPr positive cells was quantified in lumbar spinal cord (laminas I – VI in the dorsal horn) and in brain areas (thalamus and

agranular insular cortex). For each mouse, three images were taken. To determine the NOPr positive neurons in DRGs, digitized RGB (24-bit) images were analysed by using the NIH ImageJ 1.36b Software. A specific macro was created to quantify the positive areas in DRG and masseter based on pixel colour, according to a previous study [24]. For this purpose, an image from the vehicle-injected group, without reserpine induction was chosen and this macro was applied to all images from the experimental groups, as presented in the Supplementary Figure 2. To confirm the selectivity of the anti-NOPr antibody, we have used an internal antigen control (2.5 µg/ml; nociceptin receptor 337-352 peptide; Alomone, Jerusalem, Israel, Catalog Number AOR015AG0140), by using brain, spinal cord and DRG slides, according to the manufacturer's instructions. The co-incubation of the internal antigen with the primary antibody blocked the immunolabelling in all the evaluated anatomical structures, indicating the specificity of the tested antibody (Supplementary Figure 3). Furthermore, no immunolabelling was observed when the primary antibody was omitted (results not shown).

### **Biochemical parameters**

The biomarkers described in the next sections (cytokine and substance P levels, glutathione (GSH) and lactate dehydrogenase (LDH) contents) were selected based on the pathophysiology of fibromyalgia [10,23,57,64,76]. These experiments were performed to gain further insights into the beneficial effects of UFP-101 in painful- and fatigue-like symptoms in the reserpine mouse model of fibromyalgia.

### **Cytokine levels**

Brain, spinal cord, masseter muscle and serum were collected from animals that received a repeated treatment with UFP-101 (1 nmol/kg) or pregabalin (188 µmol/kg), given i.p., and the respective controls. The samples were stored in an ultra-freezer for analysis of the levels of TNF, IL-1 $\beta$  and IL-10. The tissues were homogenised according to the methodology described previously [67]. The blood was collected from the abdominal aorta, centrifuged, and the serum was frozen. Cytokine levels were measured by ELISA (sandwich enzyme-linked immunosorbent assays) kits according to the manufacturer's recommendations (R&D Systems; Minneapolis, USA) and expressed in pg/100 mg tissue (brain, spinal cord and masseter muscle) or in pg/ml (serum).

### **Substance P (SP) levels**

An ELISA assay for SP detection was performed using a commercially available kit, according to the manufacturer's recommendations (Cayman Chemical, Michigan, USA). Samples of brain, spinal cord and masseter muscle were homogenised, and further processed by using a SPE

(C-18) purification protocol prior to the ELISA assay, as described in the instructions accompanying the kit. For this experimental set, the samples were obtained from the experimental groups submitted to the chronic treatment with UFP-101 or pregabalin, and the respective control groups on the fourth day.

### ***Fibromyalgia induction and oxidative stress***

GSH contents in brain, spinal cord and muscles were determined as described before [66], as a measure of oxidative stress. The samples were collected from mice chronically treated with saline or UFP-101 (1 nmol/kg, i.p.), at the fourth day. The tissues were homogenised in saline (1:10 w/v) on ice with an Ultra Turrax (brain and spinal cord) or with a glass-Teflon homogeniser (masseter muscle). Homogenates were centrifuged at 3000 rpm for 10 min. Next, 250 µl of 4% sulfosalicylic acid were added to 250 µl of the supernatant, and centrifuged at 3000 rpm for 10 min. The supernatant (250 µl) was mixed with one ml of 0.1 M phosphate buffer (pH 8) and 5 µl of 0.01 M 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB, Sigma-Aldrich, USA). The absorbance was read at 412 nm in a spectrophotometer (GENESYS™ 10S UV-Vis, Thermo Fisher Scientific, USA). The protein content was determined by the biuret method (total protein monoreagent kit, Bioclin, Brazil) and the values of GSH contents were normalised by the protein content of the same sample.

### ***Fibromyalgia induction and lactate dehydrogenase activity in serum and mitochondrial extracts of muscle***

LDH contents in serum and mitochondrial extracts of muscles were determined as described before [26], as a biochemical marker for fatigue. The samples were collected from mice chronically treated with saline or UFP-101 (1 nmol/kg, i.p.), at the fourth day. Then, the serum was separated from the blood sample by centrifugation at 3000 rpm at 4°C for 15 min. The masseter muscle was homogenised in Isolation Buffer 1 on ice with a glass-Teflon homogeniser. Homogenates were centrifuged at 700×g for 10 min at 4°C. Next, the supernatant was centrifuged at 10,500 ×g for 10 min at 4°C. The pellet was re-suspended in 500 µl of Isolation Buffer 2 and was again centrifuged at 10,500×g for 10 min at 4°C. The final mitochondrial pellet was re-suspended in 100 µl of Isolation Buffer 2. The LDH activity in the serum and muscle were measured using a commercially available kit, according to the manufacturer's recommendations (Labtest, Minas Gerais, Brazil). The absorbance was read at 340 nm in a spectrophotometer (SpectraMax M2/M2e Microplate Readers, Molecular Devices, USA). The protein content was determined by the biuret method (total protein monoreagent kit, Bioclin, Brazil) and the values of LDH were normalised by the protein content of the same sample.

### **N/OFQ levels**

The N/OFQ levels were analysed in serum and brain of vehicle and reserpine-treated mice (collected on the fourth day), or in the saliva of fibromyalgia patients included in a major study designed to investigate the salivary levels of inflammatory mediators in chronic pain states, such as fibromyalgia and temporomandibular disorder (Human Research Ethical Committee: 844208) by ELISA assay. The *n* of control and fibromyalgia patients was 10 per group. The EIA kit [reference number EK-021-55; Nociceptin/Orphanin FQ (Human, Rat, Mouse, OX); Phoenix Pharmaceuticals, INC, USA] presents a sensitivity of 0.18 ng/ml. Brain samples were homogenised, and an ELISA assay was performed using a commercially available kit, according to the manufacturer's recommendations. The absorbance was read at 450 nm and the results were calculated from a standard curve ranging from 0.01 to 100 ng/ml.

### **Data and statistical analysis**

The results were expressed as the mean  $\pm$  standard error of the mean (SEM). Brown-Forsythe and Bartlett tests were used for checking normality of data. Statistical analysis was performed by Student *t* test (comparison between the vehicle group *vs.* the reserpine treated-group). Kruskal Wallis (non-parametric data) and one-way ANOVA (parametric data) were used for comparison of the vehicle group *vs.* reserpine treated-group *vs.* NOPr ligand treated-group. Two-way ANOVA was used for comparison of the vehicle group *vs.* reserpine treated-group *vs.* NOPr ligand treated-group, by analysing data at the baseline and on the fourth day. When the interaction of factors was statistically significant (*P* values less than 0.05), pairwise comparisons were conducted by using Dunn's or Bonferroni's *post-hoc* tests, after Kruskal Wallis and ANOVA, respectively. ELISA assays (N/OFQ, substance P and cytokine levels) were run in duplicate (for brain samples) or in a single reaction (for serum, spinal cord and masseter muscle), with an experimental *n* = 5-6 per group. All tests were performed using GraphPad Software® version 6.0 (GraphPad Software Inc., San Diego, CA, USA).

## Results

### **Dose-related acute effects of N/OFQ and UFP-101 administered by different routes**

Previous studies to validate the reserpine-induced fibromyalgia model revealed the development of mechanical and thermal hypersensitivity, associated with depression-like behaviour in mice [43]. The first part of the study was designed to evaluate the dose-response effects of N/OFQ and UFP-101 on these reserpine-elicited behavioural changes. The possible sites of action of NOPr ligands were investigated by dosing the agonist or the antagonist by i.c.v., i.t. or i.p. routes of administration. As expected, the fibromyalgia induction by the repeated administration of reserpine evoked a significant increase of the frequency withdrawal after the stimulation with the 0.4-g von Frey hair (Figure 1). The i.c.v. administration of N/OFQ (1 nmol/site) produced a slight, but not significant reduction of this response (Figure 1A). However, the i.t. (Figure 1B) or i.p. (Figure 1C) administration of N/OFQ (1 nmol/site or 1 nmol/kg) significantly reduced the fibromyalgia-related mechanical hypersensitivity (by  $62 \pm 10\%$  and  $36 \pm 11\%$ , respectively). Instead, reserpine-induced mechanical allodynia was significantly enhanced by the i.p. treatment with N/OFQ at the dose of 5 nmol/kg (Figure 1C; by  $27 \pm 6\%$ ). Concerning the UFP-101 effects, this peptide antagonist given i.c.v. (1 nmol/site; Figure 1D) or i.t. (3 and 5 nmol/site; Figure 1E) significantly reduced the mechanical hypersensitivity in mice treated with reserpine (by  $46 \pm 14\%$ ,  $53 \pm 14\%$  and  $51 \pm 13\%$ , respectively). All the tested i.p. doses of UFP-101 (1, 3 and 5 nmol/kg), apart from the 0.3 nmol/kg dose, significantly prevented the mechanical hypernociception in mice submitted to the fibromyalgia model (Figure 1F). In this case, the effects of UFP-101 lacked a typical dose-related profile (inhibition percentages of  $43 \pm 10\%$ ,  $33 \pm 5\%$  and  $28 \pm 10\%$ , respectively).

The induction of fibromyalgia by the acute administration of reserpine significantly reduced the latency time to reaction after heat stimulation, in the hot-plate apparatus (Figure 2). The acute treatment with N/OFQ or UFP-101 did not significantly alter the thermal hypersensitivity, when given by i.c.v. (Figure 2A, D) or i.p. (Figure 2C, F) routes. Noteworthy, the spinal administration of N/OFQ (3 nmol/site; i.t.) had a significant inhibitory effect on the thermal nociception (by  $49 \pm 13\%$ ; Figure 2B). Additionally, the higher tested dose of UFP-101 given i.t. (5 nmol/site) significantly prevented the thermal hypernociception in reserpine-treated mice (by  $69 \pm 13\%$ ; Figure 2E).

Reserpine-elicited fibromyalgia led to a slight increase of immobility time in the forced-swimming test, as an indicative of depressive-like behaviour (Figure 3). This parameter was significantly inhibited by N/OFQ, given by i.c.v (1 nmol/site; Figure 3A) or i.t. (3 nmol/site; Figure 3B) routes, with inhibition percentages of  $27 \pm 7\%$  and  $31 \pm 5\%$ , respectively. The systemic

administration of N/OFQ (Figure 3C) or all the routes of administration and doses tested for UFP-101 failed to significantly affect the immobility time of animals submitted to the reserpine fibromyalgia model (Figure 3D, E, F).

Considering the relevance of NOPr in anxiety, we decided to assess the effects of NOPr ligands in the plus maze paradigm in the mouse model of fibromyalgia induced by reserpine. Overall, the induction of fibromyalgia led to a diminished locomotor activity, as indicated by a significant reduction in the total number of entries in open and close arms. However, the NOPr ligands, given by i.c.v., i.t. or i.p. routes, at different doses, did not elicit any significant alteration of the locomotor index. The anxiety-related parameters, namely the entries and the time spent in open arms, were also unaffected (Supplementary Table 1).

To assess whether the effects of N/OFQ might involve the activation of opioid receptors under fibromyalgia induction by reserpine, we performed a separate series of experiments in which the animals received naloxone in combination with N/OFQ. The mice were evaluated in the same behavioural tasks as described above. The treatment with naloxone (5 µmol/kg; i.p.) did not significantly alter the effects of N/OFQ (1 nmol/kg; i.p.) in any of the evaluated parameters (Supplementary Figure 4).

The co-treatment with UFP-101 (1 nmol/kg; i.p.) significantly prevented the pro-nociceptive effect of N/OFQ (5 nmol/kg; i.p.), according to the assessment of mechanical hypersensitivity in animals subjected to the fibromyalgia model induced by reserpine (Supplementary Figure 5). However, the pre-treatment with UFP-101 (1 nmol/kg, i.p.), dosed 15 min before, failed to significantly alter the effects of N/OFQ given by i.c.v. or i.t. routes, as evaluated by von Frey and hot plate tests (Supplementary Figure 6).

#### ***The chronic administration of UFP-101 improves fibromyalgia-related pain and fatigue***

Based on the dose-response experiments, UFP-101 (1 nmol/kg), dosed by i.p. route, was tested in a repeated treatment scheme, in which the peptide NOPr antagonist was administered daily during the fibromyalgia induction protocol. The effects of UFP-101 were compared to those displayed by the positive control drug pregabalin (188 µmol/kg, i.p.). The administration of reserpine led to mechanical and thermal hypersensitivity in saline-treated control mice (Figure 4A, B). The repeated treatment with UFP-101 significantly reduced the mechanical allodynia (37 ± 8%, Figure 4A), and increased the latency time in the hot-plate test (32.2 ± 5%, Figure 4B). UFP-101 failed to alter the depression-like behaviour in the forced swimming test (Figure 4C). The administration of pregabalin resulted in a similar inhibition of mechanical and thermal

hypernociception secondary to fibromyalgia induction ( $45.5 \pm 10\%$  and  $32 \pm 7\%$ , respectively), without affecting the immobility time (Figure 4A, B, C).

Fatigue is a frequent complaint of patients with fibromyalgia diagnosis [77]. Thus, we tested the effects of repeated administration of the NOPr antagonist UFP-101 (1 nmol/kg; i.p.) on fatigue-related symptoms in reserpine-treated mice. The induction of fibromyalgia by reserpine led to a reduction in the time that animals remained in the rotarod apparatus adjusted for provoking a fatigue status. Noteworthy, the treatment with UFP-101 significantly improved the permanence time in the apparatus (with a 7-fold increase). However, the repeated scheme of treatment with the clinically used drug pregabalin (188  $\mu$ mol/kg; i.p.) lacked any significant effect in this experimental paradigm (Figure 4D). The chronic treatment with UFP-101, but not pregabalin, partially improved the grasping strength of mice subjected to fibromyalgia induction by reserpine ( $15 \pm 16\%$ ; Figure 4E). There was no difference among the experimental groups regarding the latency to fall in the Kondziela's inverted screen test (Figure 4F). In relation to the anxiety parameters, the reserpine administration decreased the total number of entries when compared to the vehicle control group, without differences for the UFP-101 and pregabalin treatments (Figure 4G, H, I).

#### ***Skeletal muscle changes related to fibromyalgia are restored by NOPr inhibition***

Considering the favourable effects of NOPr antagonism on fatigue and reduced grip strength in the reserpine fibromyalgia model, we carried out a histological analysis of masseter, gastrocnemius, and soleus sections. The induction of fibromyalgia by reserpine led to a decrease in the frequency of fibres with  $301\text{-}400 \mu\text{m}^2$  and  $401\text{-}500 \mu\text{m}^2$ , associated with an elevation in the frequency of fibres with  $601\text{-}700 \mu\text{m}^2$  in the masseter muscle (Figure 5A). In the gastrocnemius muscle, we observed an increase in the frequency of fibres with  $401\text{-}500 \mu\text{m}^2$  and  $501\text{-}600 \mu\text{m}^2$ , and a reduction of fibres with  $901\text{-}1000 \mu\text{m}^2$  and  $1001\text{-}1100 \mu\text{m}^2$  (Figure 5C). Notably, the repeated treatment with UFP-101 (1 nmol/kg; i.p.) significantly rescued the changes elicited by reserpine in the frequency of fibres with  $301\text{-}400 \mu\text{m}^2$  (masseter) and  $401\text{-}500 \mu\text{m}^2$  (gastrocnemius) (Figure 5A, C). No change was found in the cross-section area (Figure 5B, D) or weight (data not shown) of masseter and gastrocnemius muscles. The soleus muscle was also analysed for these parameters and there was no difference among the experimental groups (data not shown).

#### ***Mitochondrial analysis in reserpine-elicited fibromyalgia***

To evaluate the effects of the repeated treatment with UFP-101 (1 nmol/kg; i.p.) on the area and density of mitochondria in the skeletal muscle, we performed an ultrastructural analysis of the masseter muscle (Supplementary Figure 7). The examination of transmission electron microscopy

images revealed that either the induction of fibromyalgia by reserpine, or the chronic treatment with UFP-101, did not alter the mitochondrial area (Supplementary Figure 7D) and density (Supplementary Figure 7E).

### ***5-HT depletion and glutamate levels are not altered by UFP-101***

As demonstrated before [43], the induction of fibromyalgia by reserpine elicits a marked reduction of 5-HT levels in brain and spinal cord. Accordingly, reserpine-evoked fibromyalgia was associated with a diminishment of 5-HT levels in the pre-frontal cortex, thalamus/hypothalamus, and lumbar spinal cord, when compared with vehicle control mice. The repeated i.p. treatment with UFP-101 (1 nmol/kg) or pregabalin (188 µmol/kg) did not prevent the reduction of 5-HT in reserpine-injected mice (Figure 6A, B, C). There was no significant alteration of glutamate levels in the same anatomical structures (Figure 6D, E, F).

### ***Assessment of brain activity by microPET imaging***

Considering the marked behavioural changes associated with the fibromyalgia model induced by reserpine, we performed a microPET scan to evaluate the brain metabolism. An analysis of the whole brain did not show any significant difference in the glucose metabolism among the experimental groups (Figure 7A). However, the induction of fibromyalgia by reserpine caused a significant increment of glucose metabolic rates, according to the assessment of specific cerebral areas, such as cingulate gyrus (CG), superior colliculus (SC), and left or right inferior colliculus (LIC and RIC). The repeated i.p. administration of UFP-101 (1 nmol/kg), but not pregabalin (188 µmol/kg), reduced the glucose metabolism toward the values observed in the vehicle-treated control group (Figure 7B, C). There was no difference of brain activity among the experimental groups, in the following brain structures: striatum (RSTR and LSTR), cortex (CTX), hippocampus (RHIP and LHIP), thalamus (THA), basal forebrain/septum (BFS), hypothalamus (HYP), amygdala (RAMY and LAMY), brainstem (BS), olfactory areas (OLF) and right midbrain (RMID) (Figure 7B).

### ***ppN/OFQ and NOPr expression***

To further examining the relevance of the NOP system in fibromyalgia, we evaluated the time-related central and peripheral expression of ppN/OFQ and NOPr mRNA in mice submitted to the reserpine model. Reserpine-induced fibromyalgia was associated with an increase in ppN/OFQ mRNA expression in the lumbar spinal cord on day three (Figure 8C) and in the masseter on days one and two (Figure 8D), whereas NOPr mRNA expression was increased in the masseter muscle on day one (Figure 8H). Alternatively, the NOPr mRNA expression was reduced in the

thalamus/hypothalamus on day three (Figure 8F). The ppN/OFQ or NOPr mRNA expression in the pre-frontal cortex, the ppN/OFQ mRNA expression in the thalamus/hypothalamus, or the NOPr mRNA expression in the lumbar spinal cord were not significantly altered by the protocol of fibromyalgia induction used in this study (Figure 8A, B, E).

To extend the PCR data, the immunopositivity for NOPr was assessed in the agranular insular cortex, thalamus, lumbar spinal cord, lumbar DRGs and masseter muscle, according to the evaluation at the fourth day after initiating the protocol of fibromyalgia induction. There was a significant decrease in the immunolabelling for NOPr in the agranular insular cortex of reserpine-treated mice when compared to the vehicle control group (Figure 9A, B, C). Otherwise, the immunohistochemistry analysis revealed an increased expression of NOPr in the lumbar DRGs of reserpine-treated mice (Figure 9G, H, I). Additionally, reserpine slightly reduced the immunopositivity for NOPr in thalamus (Supplementary Figure 8A, B, C), while it failed to significantly affect NOPr distribution in the lumbar spinal cord (Figure 9D, E, F) or masseter (Supplementary Figure 8D, E, F).

### ***Analysis of inflammatory changes***

An inflammatory status has been correlated with the pathophysiology of fibromyalgia [46]. Thus, the variations of some inflammatory mediators were examined in the different experimental groups. No significant differences in SP levels were observed between reserpine- and vehicle-treated mice, in brain, spinal cord, and masseter, on the fourth day (Supplementary Figure 9A, B, C). Concerning the UFP-101 effects (1 nmol/kg; i.p.; four days), this NOPr antagonist decreased the SP levels in the masseter, when compared to the vehicle control group or to the reserpine-treated mice (by  $42 \pm 1.5\%$  and  $45 \pm 1.5\%$ , respectively) (Supplementary Figure 9C).

In a separate experimental set, the levels of TNF, IL-1 $\beta$ , and IL-10 were evaluated in brain, spinal cord, masseter and serum, at the fourth experimental day. The induction of fibromyalgia by reserpine did not cause any evident change of cytokine levels, in all the analysed samples. The repeated scheme of treatment with UFP-101 failed to significantly alter the production of cytokines, except by a reduction of the IL-1 $\beta$  levels in brain when compared to the pregabalin treatment. Unexpectedly, pregabalin (188  $\mu$ mol/kg; i.p.) caused an elevation in the levels of TNF and IL-10 in brain homogenates (Supplementary Table 2).

### ***Effects of treatment with UFP-101 on glutathione levels***

The glutathione levels were measured in brain (Supplementary Figure 10A), spinal cord (Supplementary Figure 10B), and masseter muscle (Supplementary Figure 10C) of mice, as an

indicative of oxidative stress. There were no significant differences in the levels of glutathione when comparing reserpine- and vehicle-treated mice, in any of the tested samples. In addition, the repeated treatment with UFP-101 (1 nmol/kg; i.p.) failed to affect the glutathione levels of mice submitted to the reserpine fibromyalgia model.

#### ***Effects of treatment with UFP-101 on lactate dehydrogenase activity***

LDH activity was quantified in serum (Supplementary Figure 11A) and mitochondrial extracts from muscle tissue (Supplementary Figure 11B). There were no significant differences in the LDH activity when comparing reserpine- and vehicle-treated mice, in any of the tested samples. Its activity was increased in mitochondrial extracts of UFP-101-treated mice, when compared with reserpine and vehicle groups (Supplementary Figure 11B).

#### ***N/OFQ concentrations in fibromyalgia mice and patients***

To obtain additional evidence on the relevance of the N/OFQ-NOP system in fibromyalgia, we assessed the N/OFQ levels in brain or serum obtained from mice subjected to the fibromyalgia model induced by reserpine, as well as in the saliva of a small sample of patients with fibromyalgia diagnosis. There was a slight but not statistically significant decrease of the N/OFQ levels in the serum of reserpine-treated mice in comparison to the vehicle control animals, whereas this peptide was undetectable in brain samples (Supplementary Figure 12A). N/OFQ was detected in the saliva of control or fibromyalgia patients, without any evident difference between the groups (Supplementary Figure 12B).

#### ***Effects of the non-peptide NOPr antagonist SB-612111 on fibromyalgia signs***

We performed a separate set of experiments to test the effects of SB-612111, a selective non-peptide NOPr antagonist in the reserpine-elicited fibromyalgia model. As described above, the reserpine administration led to an increase of the response frequency to mechanical stimulation, associated with a decrease in the latency to respond to heat stimulus, compared to the vehicle control groups (Figure 10A, B). The repeated treatment with SB-612111 (6.6 µmol/kg; i.p.) reduced both the mechanical allodynia ( $43 \pm 15.2\%$ , Figure 10A) and the thermal hypernociception ( $45 \pm 17.5\%$ , Figure 10B) induced by reserpine. The induction of fibromyalgia by reserpine evoked depression-like behaviour, an effect that was not altered by SB-612111, at 6.6 µmol/kg, given i.p. (Figure 10C). The repeated treatment with SB-612111 (2.2 µmol/kg; i.p.) recovered the fatigue (with a 2-fold increase) and the loss of grip strength ( $9 \pm 5.5\%$ ) caused by reserpine, according to the evaluation in the rotarod (Figure 10D) and the grasping tests (Figure 10E). The upper doses of

SB-611211 (6.6 and 22  $\mu\text{mol/kg}$ ; i.p.) failed to significantly alter both fatigue-related symptoms (Figure 10D, E). There was no difference among the experimental groups in the latency to fall in the Kondziela's inverted screen test (Figure 10F). Regarding the anxiety parameters, the reserpine administration diminished the total number of entries when compared to the vehicle control group, without any significant effect for the SB-612111 treatment (Figure 10G, H, I).

## Discussion

Herein, we evaluated the dose-related effects of the natural agonist N/OFQ, administered acutely by different routes (i.c.v., i.t and i.p), after completing the protocol of fibromyalgia induction by reserpine. This experimental set showed that i.c.v. administration of N/OFQ (1 nmol/site) did not markedly affect the mechanical allodynia or the thermal hypersensitivity secondary to fibromyalgia induction. When dosed spinally (0.3 to 3 nmol/site), N/OFQ exhibited a U-shaped profile regarding its inhibitory effects on mechanical hypersensitivity, whereas only the higher dose (3 nmol/site) prevented the thermal nociception. Spinal analgesic effects for N/OFQ have been described in rodents elsewhere [56,75]. Finally, the systemic administration of N/OFQ (i.p.; 0.3 to 5 nmol/kg) failed to alter the thermal hypersensitivity, although it potentiated the mechanical allodynia at 9  $\mu\text{g} \cdot \text{site}^{-1}$ . These results suggest that N/OFQ exhibits dual effects in the nociplastic fibromyalgia-like pain caused by reserpine in mice, extending and confirming the previous notion that N/OFQ might produce pro- or antinociceptive effects, depending on the dose and the route of treatment [1,14,39,52,63,65].

Next, we tested the dose-related effects of the peptide antagonist UFP-101, given acutely by different routes of administration. The supraspinal treatment with UFP-101 (1 nmol/site) prevented the mechanical allodynia, without any change of the thermal nociception. A previous study demonstrated analgesic effects for UFP-101, dosed i.c.v., in the thermal tail-flick test - in this case, a higher dose was used (10 nmol/site) [15]. Another study showed that UFP-101, given i.c.v., inhibited the **second phase** of formalin-induced nociception, at 10 nmol, but not at 1 nmol/site, supporting our results [63]. The same publication demonstrated **pronociceptive effects for UFP-101 (10 nmol/site), administered spinally, in the second phase of the formalin model** [63]. Herein, when given i.t., either dose of UFP-101 (3 or 5 nmol/site) reduced the mechanical allodynia, whereas only the higher dose (5 nmol/site) inhibited the thermal hypersensitivity. The acute i.p. treatment with UFP-101 (1 to 5 nmol/kg) markedly prevented the tactile allodynia, without altering the thermal hypernociception, regardless of the tested dose.

To assess the possible site (s) of action of UFP-101 when given systemically, the i.p. effects of this antagonist were tested against N/OFQ, dosed by different routes. UFP-101 (1 nmol/kg; i.p.) significantly prevented the pro-nociceptive systemic effects of N/OFQ (5 nmol/kg), whereas it failed to modify the analgesic effects of N/OFQ given i.t. or i.c.v. (1 nmol/site). Thus, UFP-101, given systemically, probably acts peripherally to induce analgesia in the reserpine-induced fibromyalgia model. In our study, naloxone failed to interfere with N/OFQ peripheral analgesic actions, discarding an interference with opioid receptors.

UFP-101 (i.p.; 1 nmol/kg) was also tested in a schedule of repeated administration, throughout the four-day protocol of fibromyalgia induction. In this protocol, UFP-101 reduced both the mechanical and the thermal hypernociception, at a similar inhibition grade as observed for the reference drug pregabalin. The different effects observed for UFP-101 in thermal nociception, when given in acute or repeated schemes, might be explained by a time-related sensitisation of the N/OFQ-NOPr system in fibromyalgia [72].

The pharmacological or genic inhibition of NOPr induced antidepressant-like effects in control rodents, whereas N/OFQ lacked any effect *per se* [34,50,62]. Herein, the administration of N/OFQ, at 1 nmol/site (i.c.v.) or 3 nmol/site (i.t.), recovered the depressive-like behaviour induced by reserpine, whereas UFP-101 failed to alter the immobility time in any of the tested schemes. Literature data showed antidepressant effects for UFP-101 administered i.c.v., in doses ranging from 1 to 10 nmol/site in naïve animals, with consistent effects only for the higher tested dose [27,28,29,50]. This might partly explain the absence of antidepressant effects for UFP-101 in our experimental model. This discrepancy might also be explained by the modulation of the N/OFQ-NOPr system after fibromyalgia induction.

The N/OFQ levels were decreased in the plasma of patients with fibromyalgia [3]. We found a slight decrement of N/OFQ levels in the serum of reserpine-treated mice, whereas the N/OFQ levels were undetectable in brain samples from vehicle- or reserpine-treated mice. The analysis of N/OFQ levels by radioimmunoassay (RIA) also detected very low levels of the peptide in brain (in fmol) [22,32,79], partly supporting our data, considering the sensitivity of the ELISA kit used by us (0.18 ng/ml). Strikingly, N/OFQ was detected in the saliva of control or fibromyalgia patients, without any evident difference between the groups. Previous studies on chronic pain demonstrated a good correlation between plasma and salivary levels of calcitonin gene related peptide (CGRP) and SP, with higher levels of both peptides in saliva [36].

Fibromyalgia encompasses musculoskeletal pain and fatigue, which is correlated with an impaired sensory-motor function [35,69]. The NOPr inhibition improved the motor disorders in the Parkinsonism model induced by high doses of reserpine [4]. Reserpine-induced fibromyalgia was associated with a marked reduction of the permanence time in the rotarod apparatus, adjusted for inducing fatigue, an effect that was prevented by the repeated treatment with UFP-101 (1 nmol/kg; i.p.). The daily UFP-101 administration also improved the grip strength, supporting its anti-fatigue effects. The non-peptide NOPr antagonist SB-612111 paralleled the favourable effects of UFP-101 in fibromyalgia-related pain and physical distress. Conversely, the reference drug pregabalin failed to rescue the fatigue or the grip strength.

The animals submitted to the reserpine model presented a mild reduction of the total number of entries in the plus-maze, an effect that was unaltered by the acute treatment with UFP-101 (0.3 to 5 nmol/kg), irrespective of the administration route. The repeated systemic treatment with UFP-101 (1 nmol/kg) or pregabalin (188 µmol/kg) also failed to modify the same parameter. Additionally, no experimental group displayed changes in the Kondziela's inverted screen test. It seems that UFP-101 is able to alleviate pain and fatigue alterations featuring the fibromyalgia symptomatology, not simply by altering the locomotor activity of mice.

Srikuea et al. (2013) [69] described an altered distribution of skeletal muscle fibre sizes in fibromyalgia women. Fibromyalgia induction by reserpine led to changes in the fibre size distribution, and the daily treatment with UFP-101 (1 nmol/kg; i.p.) partially prevented these muscle morphological changes. We also performed an ultrastructural analysis of the muscle mitochondrial areas and densities, but there was no difference of these parameters between control and fibromyalgic mice, irrespective of UFP-101 treatment. The tested experimental groups did not exhibit any difference in glutathione levels, which is a measure of oxidative stress. Nonetheless, the repeated administration of UFP-101 led to an increase in muscle LDH activity, what might partly support the preventive effects of NOPr blockade in fibromyalgia-related muscle dysfunction. The induction of fibromyalgia by reserpine did not evoke changes of SP levels in the brain, spinal cord or masseter, but the daily treatment with UFP-101 led to a reduction of SP production in the masseter. This might be an additional mechanism to explain the UFP-101 effects on fibromyalgia-related fatigue. Interestingly, a randomized clinical trial correlated the benefits of acupuncture in alleviating the fibromyalgia symptoms with a reduction of SP serum levels [38]. Central or peripheral changes in cytokine production have been described in fibromyalgia patients [73]. Furthermore, the N/OFQ-NOPr system has been suggested to influence the balance between pro- and anti-inflammatory cytokines [9]. Herein, we did not detect any significant alteration of TNF, IL-1 $\beta$  or IL-10 in the brain, spinal cord or muscle samples of reserpine-treated mice, regardless of the administration of UFP-101.

The animals submitted to the reserpine model presented a reduction of the brain and spinal 5-HT levels, an event that was not modified by UFP-101. The same brain and spinal cord samples were used for analysis of glutamate levels, lacking any significant difference. A recent study [41] showed reduced levels of 5-HT, noradrenaline, and dopamine, without any alteration of glutamate and GABA, in the spinal cord reserpine-treated rats. The authors showed a recovery of monoamine levels by duloxetine, whereas pregabalin failed to modify these changes. Apparently, UFP-101, as pregabalin, displays beneficial effects on fibromyalgia symptoms, independent on the ability to restore the monoamine levels.

A functional MRI study demonstrated different patterns of brain activation following innocuous or noxious stimulation in rats submitted to the reserpine model of fibromyalgia [81]. Herein, a micro-PET scanning analysis demonstrated an increased brain activity in the mouse CG, SC, LIC and RIC, secondary to the induction of fibromyalgia, an effect that was restored by UFP-101 treatment. Neuroimaging studies revealed a reduction of fibromyalgia-related brain hyperactivity in patients submitted to pharmacological and non-pharmacological treatment protocols [37,53]. Thus, the inhibition of brain hyperactivity by UFP-101 might underlie its beneficial effects in fibromyalgia-related painful and fatigue symptoms.

The analgesic effects observed for daily UFP-101 or SB-612111 might be explained by the increased NOPr immunoreactivity in the DRG of reserpine-treated mice. The small unmyelinated DRG neurons are peptidergic and non-peptidergic C nociceptors, primordial to heat and mechanical pain, respectively [72]. An upregulation of NOPr in DRG has also been demonstrated in other chronic pain models [12,16]. The induction of fibromyalgia by reserpine led to an increase of ppN/OFQ mRNA in the spinal cord, further supporting that changes in the N/OFQ-NOPr tonus underlie dysfunctional pain. Alternatively, reserpine-treated mice showed a reduction of immunolabelling for NOPr in the agranular insular cortex, which is a pivotal region for pain control [30]. A decrease of NOPr expression was previously demonstrated in thalamus and hippocampus in a mouse model of neuropathic pain caused by sciatic nerve lesion [59]. Curiously, the levels of ppN/OFQ mRNA were significantly increased in masseter, from the first to the third day after reserpine injection, what might further support the anti-fatigue effects of the daily treatment with NOPr antagonists.

Collectively, these results help to pave the way on the pathophysiology of fibromyalgia, indicating that NOPr antagonism might be an alternative for managing of fibromyalgia-related pain and tenderness.

## Acknowledgements

We would like to thank Mrs. Janaína Pasetti Nunes for her valuable technical assistance in histological processing (Laboratory of Oral Pathology/PUCRS). The authors are grateful to Mrs. Moema Queiroz Vieira (Central Laboratory of Microscopy and Microanalysis; LabCEMM/PUCRS) for the technical assistance in transmission electron microscopy experiments. We also thank the staff of the Centre of Experimental Biological Models (CeMBE/PUCRS) for animal care. This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) Finance Code 001, Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), and Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), in addition to a Financiadora de Estudos e Projetos (FINEP) research grant “Expansão da Infraestrutura Multusuária de Pesquisa na PUCRS (MULTIPUCRS)” # 01.13.0292-00 (Brazil). A.P.A.D. is a PhD student receiving grants from CAPES and T.C.B.P is recipient of PNPD/CAPES fellowship. M.M.C. (CNPq, 303842-2014-8) and M.R.B. (CNPq, 303776/2013-7) are Research Career Awardees of the National Research Council of Brazil (CNPq).

## Conflict of interest statement

The authors declare no conflict of interest.

## References

- [1] Agostini S, Eutamene H, Broccardo M, Improta G, Petrella C, Theodorou V, Bueno L. Peripheral anti-nociceptive effect of nociceptin/orphanin FQ in inflammation and stress-induced colonic hyperalgesia in rats. *Pain* 2009;141:292-299.
- [2] Alexander SP, Christopoulos A, Davenport AP, Kelly E, Marrion NV, Peters JA, Faccenda E, Harding SD, Pawson AJ, Sharman JL, Southan C, Davies JA, Collaborators C. The concise guide to pharmacology 2017/18: G protein-coupled receptors. *Br J Pharmacol* 2017;174 Suppl 1:S17-S129.
- [3] Anderberg UM, Liu Z, Berglund L, Nyberg F. Plasma levels on nociceptin in female fibromyalgia syndrome patients. *Z Rheumatol* 1998;57 Suppl 2:77-80.
- [4] Arcuri L, Mercatelli D, Morari M. Parkinson's disease: no NOP, new hope. *Oncotarget* 2017;8:8995-8996.
- [5] Arora V, Chopra K. Possible involvement of oxido-nitrosative stress induced neuro-inflammatory cascade and monoaminergic pathway: underpinning the correlation between nociceptive and depressive behaviour in a rodent model. *J Affect Disord* 2013;151:1041-1052.
- [6] Aydede M, Shriver A. Recently introduced definition of "nociceptive pain" by the International Association for the Study of Pain needs better formulation. *Pain* 2018;159:1176-1177.
- [7] Baraniuk JN, Whalen G, Cunningham J, Clauw DJ. Cerebrospinal fluid levels of opioid peptides in fibromyalgia and chronic low back pain. *BMC Musculoskelet Disord* 2004;5:48.
- [8] Blasco-Serra A, Escrihuela-Vidal F, Gonzalez-Soler EM, Martinez-Exposito F, Blasco-Ausina MC, Martinez-Bellver S, Cervera-Ferri A, Teruel-Marti V, Valverde-Navarro AA. Depressive-like symptoms in a reserpine-induced model of fibromyalgia in rats. *Physiol Behav* 2015;151:456-462.
- [9] Bodera P, Stankiewicz W, Kocik J. Interactions of orphanin FQ/nociceptin (OFQ/N) system with immune system factors and hypothalamic-pituitary-adrenal (HPA) axis. *Pharmacol Rep* 2014;66:288-291.
- [10] Bonaterra GA, Then H, Oezel L, Schwarzbach H, Ocker M, Thieme K, Di Fazio P, Kinscherf R. Morphological Alterations in Gastrocnemius and Soleus Muscles in Male and Female Mice in a Fibromyalgia Model. *PLoS One* 2016;11:e0151116.
- [11] Brederson JD, Jarvis MF, Honore P, Surowy CS. Fibromyalgia: mechanisms, current treatment and animal models. *Curr Pharm Biotechnol* 2011;12:1613-1626.
- [12] Briscini L, Corradini L, Ongini E, Bertorelli R. Up-regulation of ORL-1 receptors in spinal tissue of allodynic rats after sciatic nerve injury. *Eur J Pharmacol* 2002;447:59-65.

- [13] Cabo-Meseguer A, Cerdá-Olmedo G, Trillo-Mata JL. Fibromyalgia: Prevalence, epidemiologic profiles and economic costs. *Med Clin (Barc)* 2017;149:441-448.
- [14] Calo G, Guerrini R, Rizzi A, Salvadori S, Regoli D. Pharmacology of nociceptin and its receptor: a novel therapeutic target. *Br J Pharmacol* 2000;129:1261-1283.
- [15] Calo G, Rizzi A, Rizzi D, Bigoni R, Guerrini R, Marzola G, Martí M, McDonald J, Morari M, Lambert DG, Salvadori S, Regoli D. [Nphe1,Arg14,Lys15]nociceptin-NH<sub>2</sub>, a novel potent and selective antagonist of the nociceptin/orphanin FQ receptor. *Br J Pharmacol* 2002;136:303-311.
- [16] Chen Y, Sommer C. Nociceptin and its receptor in rat dorsal root ganglion neurons in neuropathic and inflammatory pain models: implications on pain processing. *J Peripher Nerv Syst* 2006;11:232-240.
- [17] Chiou LC, Liao YY, Fan PC, Kuo PH, Wang CH, Riemer C, Prinssen EP. Nociceptin/orphanin FQ peptide receptors: pharmacology and clinical implications. *Curr Drug Targets* 2007;8:117-135.
- [18] Clauw DJ. Fibromyalgia: a clinical review. *JAMA* 2014;311:1547-1555.
- [19] da Silveira NS, de Oliveira-Silva GL, Lamanes Bde F, Prado LC, Bispo-da-Silva LB. The aversive, anxiolytic-like, and verapamil-sensitive psychostimulant effects of pulegone. *Biol Pharm Bull* 2014;37:771-778.
- [20] Deacon RM. Measuring the strength of mice. *J Vis Exp* 2013;76:1-4.
- [21] DeSantana JM, da Cruz KM, Sluka KA. Animal models of fibromyalgia. *Arthritis Res Ther* 2013;15:222.
- [22] Devine DP, Hoversten MT, Ueda Y, Akil H. Nociceptin/orphanin FQ content is decreased in forebrain neurones during acute stress. *J Neuroendocrinol* 2003;15:69-74.
- [23] Fatima G, Das SK, Mahdi AA. Some oxidative and antioxidative parameters and their relationship with clinical symptoms in women with fibromyalgia syndrome. *Int J Rheum Dis* 2017;20:39-45.
- [24] Freitas RD, Costa KM, Nicoletti NF, Kist LW, Bogo MR, Campos MM. Omega-3 fatty acids are able to modulate the painful symptoms associated to cyclophosphamide-induced-hemorrhagic cystitis in mice. *J Nutr Biochem* 2016;27:219-232.
- [25] Fulford AJ. Endogenous nociceptin system involvement in stress responses and anxiety behavior. *Vitam Horm* 2015;97:267-293.

- [26] Garcia-Cazarin ML, Snider NN, Andrade FH. Mitochondrial isolation from skeletal muscle. *J Vis Exp* 2011;49:1-4.
- [27] Gavioli EC, Calo G. Nociceptin/orphanin FQ receptor antagonists as innovative antidepressant drugs. *Pharmacol Ther* 2013;140:10-25.
- [28] Gavioli EC, Marzola G, Guerrini R, Bertorelli R, Zucchini S, De Lima TC, Rae GA, Salvadori S, Regoli D, Calo G. Blockade of nociceptin/orphanin FQ-NOP receptor signalling produces antidepressant-like effects: pharmacological and genetic evidences from the mouse forced swimming test. *Eur J Neurosci* 2003;17:1987-1990.
- [29] Gavioli EC, Vaughan CW, Marzola G, Guerrini R, Mitchell VA, Zucchini S, De Lima TC, Rae GA, Salvadori S, Regoli D, Calo G. Antidepressant-like effects of the nociceptin/orphanin FQ receptor antagonist UFP-101: new evidence from rats and mice. *Naunyn Schmiedebergs Arch Pharmacol* 2004;369:547-553.
- [30] George O, Koob GF. Individual differences in prefrontal cortex function and the transition from drug use to drug dependence. *Neurosci Biobehav Rev* 2010;35:232-247.
- [31] Goldfarb Y, Reinscheid RK, Kusnecov AW. Orphanin FQ/nociceptin interactions with the immune system in vivo: gene expression changes in lymphoid organs and regulation of the cytokine response to staphylococcal enterotoxin A. *J Neuroimmunol* 2006;176:76-85.
- [32] Granholm L, Roman E, Nylander I. Single housing during early adolescence causes time-, area- and peptide-specific alterations in endogenous opioids of rat brain. *Br J Pharmacol* 2015;172:606-614.
- [33] Hernandez-Leon A, De la Luz-Cuellar YE, Granados-Soto V, Gonzalez-Trujano ME, Fernandez-Guasti A. Sex differences and estradiol involvement in hyperalgesia and allodynia in an experimental model of fibromyalgia. *Horm Behav* 2018;97:39-46.
- [34] Holanda VAD, Santos WB, Asth L, Guerrini R, Calo G, Ruzza C, Gavioli EC. NOP agonists prevent the antidepressant-like effects of nortriptyline and fluoxetine but not R-ketamine. *Psychopharmacology (Berl)* 2018;235:3093-3102.
- [35] Holtermann A, Gronlund C, Roeleveld K, Gerdle B. The relation between neuromuscular control and pain intensity in fibromyalgia. *J Electromyogr Kinesiol* 2011;21:519-524.
- [36] Jang MU, Park JW, Kho HS, Chung SC, Chung JW. Plasma and saliva levels of nerve growth factor and neuropeptides in chronic migraine patients. *Oral Dis* 2011;17:187-193.
- [37] Jorge LL, Amaro E, Jr. Brain imaging in fibromyalgia. *Curr Pain Headache Rep* 2012;16:388-398.

- [38] Karatay S, Okur SC, Uzkeser H, Yildirim K, Akcay F. Effects of Acupuncture Treatment on Fibromyalgia Symptoms, Serotonin, and Substance P Levels: A Randomized Sham and Placebo-Controlled Clinical Trial. *Pain Med* 2018;19:615-628.
- [39] Kiguchi N, Ding H, Ko MC. Central N/OFQ-NOP Receptor System in Pain Modulation. *Adv Pharmacol* 2016;75:217-243.
- [40] Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: The ARRIVE guidelines for reporting animal research. *J Pharmacol Pharmacother* 2010;1:94-99.
- [41] Kiso T, Moriyama A, Furutani M, Matsuda R, Funatsu Y. Effects of pregabalin and duloxetine on neurotransmitters in the dorsal horn of the spinal cord in a rat model of fibromyalgia. *Eur J Pharmacol* 2018;827:117-124.
- [42] Klein CP, Cintra MR, Binda N, Diniz DM, Gomez MV, Souto AA, de Souza AH. Coadministration of Resveratrol and Rice Oil Mitigates Nociception and Oxidative State in a Mouse Fibromyalgia-Like Model. *Pain Res Treat* 2016;2016:3191638.
- [43] Klein CP, Sperotto ND, Maciel IS, Leite CE, Souza AH, Campos MM. Effects of D-series resolvins on behavioral and neurochemical changes in a fibromyalgia-like model in mice. *Neuropharmacology* 2014;86:57-66.
- [44] Lambert DG. The nociceptin/orphanin FQ receptor: a target with broad therapeutic potential. *Nat Rev Drug Discov* 2008;7:694-710.
- [45] Lima KG, Krause GC, da Silva EFG, Xavier LL, Martins LAM, Alice LM, da Luz LB, Gassen RB, Filippi-Chiela EC, Haute GV, Garcia MCR, Funchal GA, Pedrazza L, Reghelin CK, de Oliveira JR. Octyl gallate reduces ATP levels and Ki67 expression leading HepG2 cells to cell cycle arrest and mitochondria-mediated apoptosis. *Toxicol In Vitro* 2018;48:11-25.
- [46] Littlejohn G, Guymer E. Neurogenic inflammation in fibromyalgia. *Semin Immunopathol* 2018;40:291-300.
- [47] Maciel IS, Silva RB, Morrone FB, Calixto JB, Campos MM. Synergistic effects of celecoxib and bupropion in a model of chronic inflammation-related depression in mice. *PLoS One* 2013;8:e77227.
- [48] Marti M, Mela F, Budri M, Volta M, Malfacini D, Molinari S, Zaveri NT, Ronzoni S, Petrillo P, Calo G, Morari M. Acute and chronic antiparkinsonian effects of the novel nociceptin/orphanin FQ receptor antagonist NiK-21273 in comparison with SB-612111. *Br J Pharmacol* 2013;168:863-879.

- [49] McGrath JC, Lilley E. Implementing guidelines on reporting research using animals (ARRIVE etc.): new requirements for publication in BJP. *Br J Pharmacol* 2015;172:3189-3193.
- [50] Medeiros IU, Ruzza C, Asth L, Guerrini R, Romao PR, Gavioli EC, Calo G. Blockade of nociceptin/orphanin FQ receptor signaling reverses LPS-induced depressive-like behavior in mice. *Peptides* 2015;72:95-103.
- [51] Micheli L, Di Cesare Mannelli L, Guerrini R, Trapella C, Zanardelli M, Cicoccioppo R, Rizzi A, Ghelardini C, Calo G. Acute and subchronic antinociceptive effects of nociceptin/orphanin FQ receptor agonists infused by intrathecal route in rats. *Eur J Pharmacol* 2015;754:73-81.
- [52] Mogil JS, Pasternak GW. The molecular and behavioral pharmacology of the orphanin FQ/nociceptin peptide and receptor family. *Pharmacol Rev* 2001;53:381-415.
- [53] Murga I, Guillen V, Lafuente JV. Cerebral magnetic resonance changes associated with fibromyalgia syndrome. *Med Clin (Barc)* 2017;148:511-516.
- [54] Nagakura Y, Oe T, Aoki T, Matsuoka N. Biogenic amine depletion causes chronic muscular pain and tactile allodynia accompanied by depression: A putative animal model of fibromyalgia. *Pain* 2009;146:26-33.
- [55] Nagakura Y, Ohsaka N, Azuma R, Takahashi S, Takebayashi Y, Kawasaki S, Murai S, Miwa M, Saito H. Monoamine system disruption induces functional somatic syndromes associated symptomatology in mice. *Physiol Behav* 2018;194:505-514.
- [56] Nazzaro C, Rizzi A, Salvadori S, Guerrini R, Regoli D, Zeilhofer HU, Calo G. UFP-101 antagonizes the spinal antinociceptive effects of nociceptin/orphanin FQ: behavioral and electrophysiological studies in mice. *Peptides* 2007;28:663-669.
- [57] Oezel L, Then H, Jung AL, Jabari S, Bonaterra GA, Wissniowski TT, Onel SF, Ocker M, Thieme K, Kinscherf R, Di Fazio P. Fibromyalgia syndrome: metabolic and autophagic processes in intermittent cold stress mice. *Pharmacol Res Perspect* 2016;4:e00248.
- [58] Ozawa A, Brunori G, Cippitelli A, Toll N, Schoch J, Kieffer BL, Toll L. Analysis of the distribution of spinal NOP receptors in a chronic pain model using NOP-eGFP knock-in mice. *Br J Pharmacol* 2018;175:2662-2675.
- [59] Palmisano M, Mercatelli D, Caputi FF, Carretta D, Romualdi P, Candeletti S. N/OFQ system in brain areas of nerve-injured mice: its role in different aspects of neuropathic pain. *Genes Brain Behav* 2017;16:537-545.

- [60] Pernot F, Dorandeu F, Beup C, Peinnequin A. Selection of reference genes for real-time quantitative reverse transcription-polymerase chain reaction in hippocampal structure in a murine model of temporal lobe epilepsy with focal seizures. *J Neurosci Res* 2010;88:1000-1008.
- [61] Reinscheid RK. The Orphanin FQ / Nociceptin receptor as a novel drug target in psychiatric disorders. *CNS Neurol Disord Drug Targets* 2006;5:219-224.
- [62] Rizzi A, Gavioli EC, Marzola G, Spagnolo B, Zucchini S, Ciccocioppo R, Trapella C, Regoli D, Calo G. Pharmacological characterization of the nociceptin/orphanin FQ receptor antagonist SB-612111 [(-)-cis-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl)methyl]-6,7,8,9-tetrahydro-5H-benzocyclohepten-5-ol]: in vivo studies. *J Pharmacol Exp Ther* 2007;321:968-974.
- [63] Rizzi A, Nazzaro C, Marzola GG, Zucchini S, Trapella C, Guerrini R, Zeilhofer HU, Regoli D, Calo G. Endogenous nociceptin/orphanin FQ signalling produces opposite spinal antinociceptive and supraspinal pronociceptive effects in the mouse formalin test: pharmacological and genetic evidences. *Pain* 2006;124:100-108.
- [64] Rodriguez-Pinto I, Agmon-Levin N, Howard A, Shoenfeld Y. Fibromyalgia and cytokines. *Immunol Lett* 2014;161:200-203.
- [65] Schroder W, Lambert DG, Ko MC, Koch T. Functional plasticity of the N/OFQ-NOP receptor system determines analgesic properties of NOP receptor agonists. *Br J Pharmacol* 2014;171:3777-3800.
- [66] Shim JY, Kim MH, Kim HD, Ahn JY, Yun YS, Song JY. Protective action of the immunomodulator ginsan against carbon tetrachloride-induced liver injury via control of oxidative stress and the inflammatory response. *Toxicol Appl Pharmacol* 2010;242:318-325.
- [67] Silva RBM, Greggio S, Venturin GT, da Costa JC, Gomez MV, Campos MM. Beneficial Effects of the Calcium Channel Blocker CTK 01512-2 in a Mouse Model of Multiple Sclerosis. *Mol Neurobiol* 2018.
- [68] Sleigh JN, Weir GA, Schiavo G. A simple, step-by-step dissection protocol for the rapid isolation of mouse dorsal root ganglia. *BMC Res Notes* 2016;9:82.
- [69] Srikuea R, Symons TB, Long DE, Lee JD, Shang Y, Chomentowski PJ, Yu G, Crofford LJ, Peterson CA. Association of fibromyalgia with altered skeletal muscle characteristics which may contribute to postexertional fatigue in postmenopausal women. *Arthritis Rheum* 2013;65:519-528.
- [70] Staud R. Is it all central sensitization? Role of peripheral tissue nociception in chronic musculoskeletal pain. *Curr Rheumatol Rep* 2010;12:448-454.

- [71] Takeuchi Y, Takasu K, Ono H, Tanabe M. Pregabalin, S-(+)-3-isobutylgaba, activates the descending noradrenergic system to alleviate neuropathic pain in the mouse partial sciatic nerve ligation model. *Neuropharmacology* 2007;53:842-853.
- [72] Toll L, Bruchas MR, Calo G, Cox BM, Zaveri NT. Nociceptin/Orphanin FQ Receptor Structure, Signaling, Ligands, Functions, and Interactions with Opioid Systems. *Pharmacol Rev* 2016;68:419-457.
- [73] Totsch SK, Sorge RE. Immune System Involvement in Specific Pain Conditions. *Mol Pain* 2017;13:1744806917724559.
- [74] Trevisan G, Rossato MF, Walker CI, Oliveira SM, Rosa F, Tonello R, Silva CR, Machado P, Boligon AA, Martins MA, Zanatta N, Bonacorso HG, Athayde ML, Rubin MA, Calixto JB, Ferreira J. A novel, potent, oral active and safe antinociceptive pyrazole targeting kappa opioid receptors. *Neuropharmacology* 2013;73:261-273.
- [75] Tsai CY, Poon YY, Huang YH, Chan SH. Activation of spinal nociceptin receptors induces cardiovascular depression and antinociception in an independent manner in mice. *J Pain Res* 2018;11:2699-2708.
- [76] Tsilioni I, Russell IJ, Stewart JM, Gleason RM, Theoharides TC. Neuropeptides CRH, SP, HK-1, and Inflammatory Cytokines IL-6 and TNF Are Increased in Serum of Patients with Fibromyalgia Syndrome, Implicating Mast Cells. *J Pharmacol Exp Ther* 2016;356:664-672.
- [77] Vincent A, Benzo RP, Whipple MO, McAllister SJ, Erwin PJ, Saligan LN. Beyond pain in fibromyalgia: insights into the symptom of fatigue. *Arthritis Res Ther* 2013;15:221.
- [78] Walf AA, Frye CA. The use of the elevated plus maze as an assay of anxiety-related behavior in rodents. *Nat Protoc* 2007;2:322-328.
- [79] Walker JR, Terenius L, Koob GF. Conditioned opioid withdrawal decreases nociceptin/orphanin FQ levels in the frontal cortex and olfactory tubercle. *Neuropsychopharmacology* 2002;27:203-211.
- [80] Wang J, Li LZ, Liu YG, Teng LR, Lu JH, Xie J, Hu WJ, Liu Y, Liu Y, Wang D, Teng le S. Investigations on the antifatigue and antihypoxic effects of Paecilomyces hepiali extract. *Mol Med Rep* 2016;13:1861-1868.
- [81] Wells JA, Shibata S, Fujikawa A, Takahashi M, Saga T, Aoki I. Functional MRI of the Reserpine-Induced Putative Rat Model of Fibromyalgia Reveals Discriminatory Patterns of Functional Augmentation to Acute Nociceptive Stimuli. *Sci Rep* 2017;7:38325.

- [82] Witkin JM, Statnick MA, Rorick-Kehn LM, Pintar JE, Ansonoff M, Chen Y, Tucker RC, Ciccocioppo R. The biology of Nociceptin/Orphanin FQ (N/OFQ) related to obesity, stress, anxiety, mood, and drug dependence. *Pharmacol Ther* 2014;141:283-299.
- [83] Wolfe F, Clauw DJ, Fitzcharles MA, Goldenberg DL, Hauser W, Katz RL, Mease PJ, Russell AS, Russell IJ, Walitt B. 2016 Revisions to the 2010/2011 fibromyalgia diagnostic criteria. *Semin Arthritis Rheum* 2016;46:319-329.
- [84] Zanirati G, Azevedo PN, Venturin GT, Greggio S, Alcara AM, Zimmer ER, Feltes PK, DaCosta JC. Depression comorbidity in epileptic rats is related to brain glucose hypometabolism and hypersynchronicity in the metabolic network architecture. *Epilepsia* 2018;59:923-934.
- [85] Zaratin PF, Petrone G, Sbacchi M, Garnier M, Fossati C, Petrillo P, Ronzoni S, Giardina GA, Scheideler MA. Modification of nociception and morphine tolerance by the selective opiate receptor-like orphan receptor antagonist (-)-cis-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl)methyl]-6,7,8,9-tetrahydro-5H-benzocyclohepten-5-ol (SB-612111). *J Pharmacol Exp Ther* 2004;308:454-461.
- [86] Zaveri N. Peptide and nonpeptide ligands for the nociceptin/orphanin FQ receptor ORL1: research tools and potential therapeutic agents. *Life Sci* 2003;73:663-678.
- [87] Zaveri NT. Nociceptin Opioid Receptor (NOP) as a Therapeutic Target: Progress in Translation from Preclinical Research to Clinical Utility. *J Med Chem* 2016;59:7011-7028.

## Figure Legends

**Figure 1:** Dose-related effects of N/OFQ or UFP-101 on the mechanical hypersensitivity in the mouse model of fibromyalgia induced by reserpine. Effects of acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v.; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on hindpaw withdrawal threshold (response frequency in percentage) to tactile stimulation. The mechanical hypersensitivity was assessed by using the Von Frey filaments before (baseline), and at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 15 min (i.c.v. and i.t.) or 30 min (i.p.) before the behavioural evaluations. Each column represents the mean  $\pm$  SEM. \* $p$  < 0.05 when compared to the control vehicle/saline group, indicating the development of mechanical allodynia; # $p$  < 0.05 when compared to the reserpine/saline group. Statistical analysis was performed by two-way ANOVA, followed by Bonferroni's *post hoc* test (A-F). (A)  $n$  = 6-8 mice/group; (B)  $n$  = 8-12 mice/group; (C)  $n$  = 8-25 mice/group; (D)  $n$  = 8-11 mice/group; (E)  $n$  = 8-12 mice/group; (F)  $n$  = 8-26 mice/group.

**Figure 2:** Dose-related effects of N/OFQ or UFP-101 on the thermal hypersensitivity in reserpine-treated mice. Effects of acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v.; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on the latency time (s) in response to hot thermal stimulation. Thermal hypersensitivity was assessed in the hot-plate test, at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 15 min (i.c.v. and i.t.) or 30 min (i.p.) before the behavioural evaluations. Each column represents the mean  $\pm$  SEM. \* $p$  < 0.05 when compared to the control vehicle/saline group, indicating the development of thermal sensitivity; # $p$  < 0.05 when compared to the reserpine/saline group. Statistical analysis was performed by two-way ANOVA followed by Bonferroni's *post hoc* test (A-F). (A)  $n$  = 6-8 mice/group; (B)  $n$  = 8-12 mice/group; (C)  $n$  = 8-25 mice/group; (D)  $n$  = 8-11 mice/group; (E)  $n$  = 8-12 mice/group; (F)  $n$  = 8-26 mice/group.

**Figure 3:** Dose-related effects of the acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v.; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on depression-like behaviour in reserpine-treated mice submitted to the forced swimming test. The immobility time (s) was assessed at fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 30 min before the behavioural evaluations. Each column represents the mean  $\pm$  SEM.

\* $p < 0.05$  when compared to the control vehicle/saline group, indicating the development of depressive behaviour; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (B, C, E and F) or one-way ANOVA followed by Bonferroni's *post hoc* test (A and D). (A)  $n = 6\text{-}8$  mice/group; (B)  $n = 8\text{-}12$  mice/group; (C)  $n = 8\text{-}25$  mice/group; (D)  $n = 8\text{-}11$  mice/group; (E)  $n = 8\text{-}12$  mice/group; (F)  $n = 8\text{-}26$  mice/group.

**Figure 4:** Effects of the repeated treatment with UFP-101 or pregabalin on painful-, fatigue-, depressive- and anxiety-like behaviours in reserpine-treated mice. The effects of both drugs were also assessed on the fatigue and grip strength in the fibromyalgia model elicited by reserpine. Effects of the repeated administration of UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188  $\mu$ mol/kg, intraperitoneal), on hind paw mechanical allodynia (A), latency time (s) in response to hot thermal stimulation (B), immobility time in the forced swimming test (C), rotating time (D), grasping strength (E), latency to fall (F), and on plus maze parameters (G-I). All the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). UFP-101 or pregabalin were dosed for three consecutive days, 30 min after reserpine injection. At the fourth day, mice also received UFP-101 or pregabalin, 30 min before evaluations. Each column represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (D, F, H and I), one-way (C and G) or two-way (A, B and E) ANOVA followed by Bonferroni's *post hoc* test. (A-C)  $n = 8\text{-}11$  mice/group; (D-E)  $n = 8\text{-}10$  mice/group; (F-I)  $n = 8$  mice/group.

**Figure 5:** Histological analysis of masseter (A and B) and gastrocnemius (C and D) muscles of reserpine-treated mice. Effects of intraperitoneal (i.p.) repeated treatment with UFP-101 (1 nmol/kg) on fibre size distribution (A and C) and mean fibre cross-sectional area (B and D). UFP-101 was administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days) and 30 min before the muscle collection, at the fourth day. Each point or column represent the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  compared to reserpine/saline group. Statistical analysis was performed by one-way (cross sectional area) or two-way (frequency of fibres) ANOVA followed by Bonferroni's *post hoc* test. (A-D)  $n = 5$  mice/group.

**Figure 6:** Serotonin and glutamate levels in prefrontal cortex (A and D), thalamus/hypothalamus (B and E) and lumbar spinal cord (C and F) of reserpine-treated mice. UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188 µmol/kg, intraperitoneal) were administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection at the fourth day. Serotonin (A-C) and glutamate (D-F) levels are expressed in ng/g tissue. Each bar represents the mean ± SEM. \* $p < 0.05$  when compared to the control vehicle/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (A and C) or one-way ANOVA followed by Bonferroni's *post hoc* test (B, D, E and F). (A and D)  $n = 7\text{-}8$  mice/group; (B, C, E and F)  $n = 8$  mice/group.

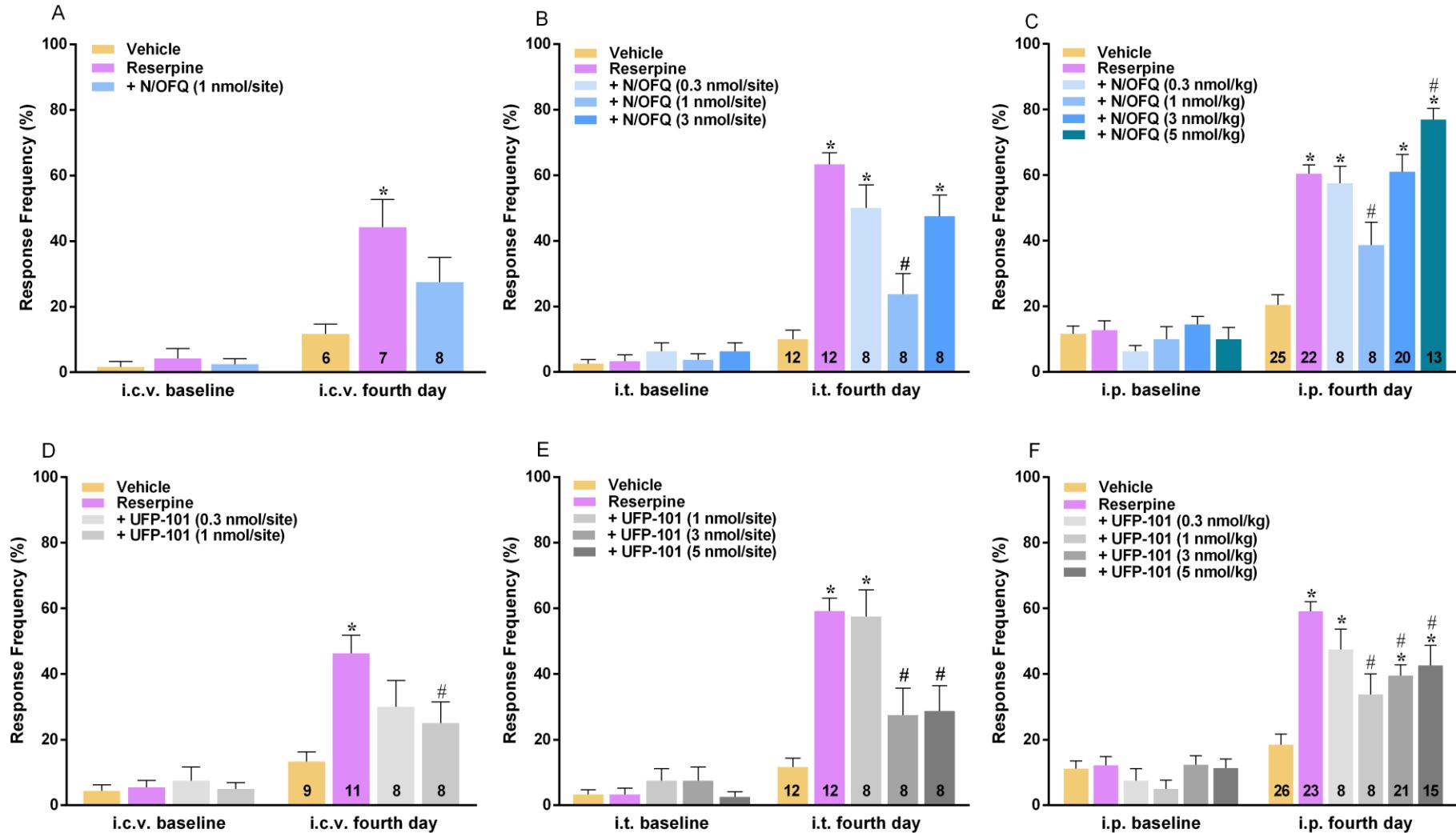
**Figure 7:** Effects of the repeated treatment with UFP-101 or pregabalin on the [<sup>18</sup>F]-FDG hypermetabolism in the whole brain (A) or in several brain structures (B) in the fibromyalgia-like model induced by reserpine in mice. UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188 µmol/kg, intraperitoneal) were administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection at the fourth day. Representative images of the coronal plane of the vehicle control group (CI), the reserpine-treated group (CII), the group treated with UFP-101 (CIII), or the group treated with pregabalin (CIV). Each column represents the mean ± SEM. \* $p < 0.05$  when compared to the control vehicle/saline group. Differences in the standardised uptake value ratio (SUVr) per areas of brain and in whole brain were determined by one-way (A) or two-way ANOVA (B), followed by Bonferroni's *post hoc* tests, respectively. (A-B)  $n = 7\text{-}8$  mice/group. Abbreviations: RSTR, right striatum region; LSTR, left striatum region; CTX, cortex; RHIP, right hippocampal region; LHIP, left hippocampal region; THA, thalamus; BFS, basal forebrain/septum; HYP, hypothalamus; RAMY, right amygdala; LAMY, left amygdala; BS, brainstem; CG, cingulate gyrus; SC, superior colliculus; OLF, olfactory areas; RMID, right midbrain; LMID, left midbrain; LIC, left inferior colliculus; RIC, right inferior colliculus.

**Figure 8:** The ppN/OFQ (A-D) and NOP receptor (E-H) mRNA expression was measured by RT-qPCR in prefrontal cortex (A and E), thalamus/hypothalamus (B and F), lumbar spinal cord (C and G) and masseter muscle (D and H) tissues, at days 1, 2 and 3 after the administration of reserpine (0.25 mg/kg; subcutaneous). Each scatter dot plot represents the mean ± SEM of 6-8 samples. \* $p < 0.05$  when compared to the control vehicle/saline group. Statistical analysis was performed by unpaired Student *t* test.

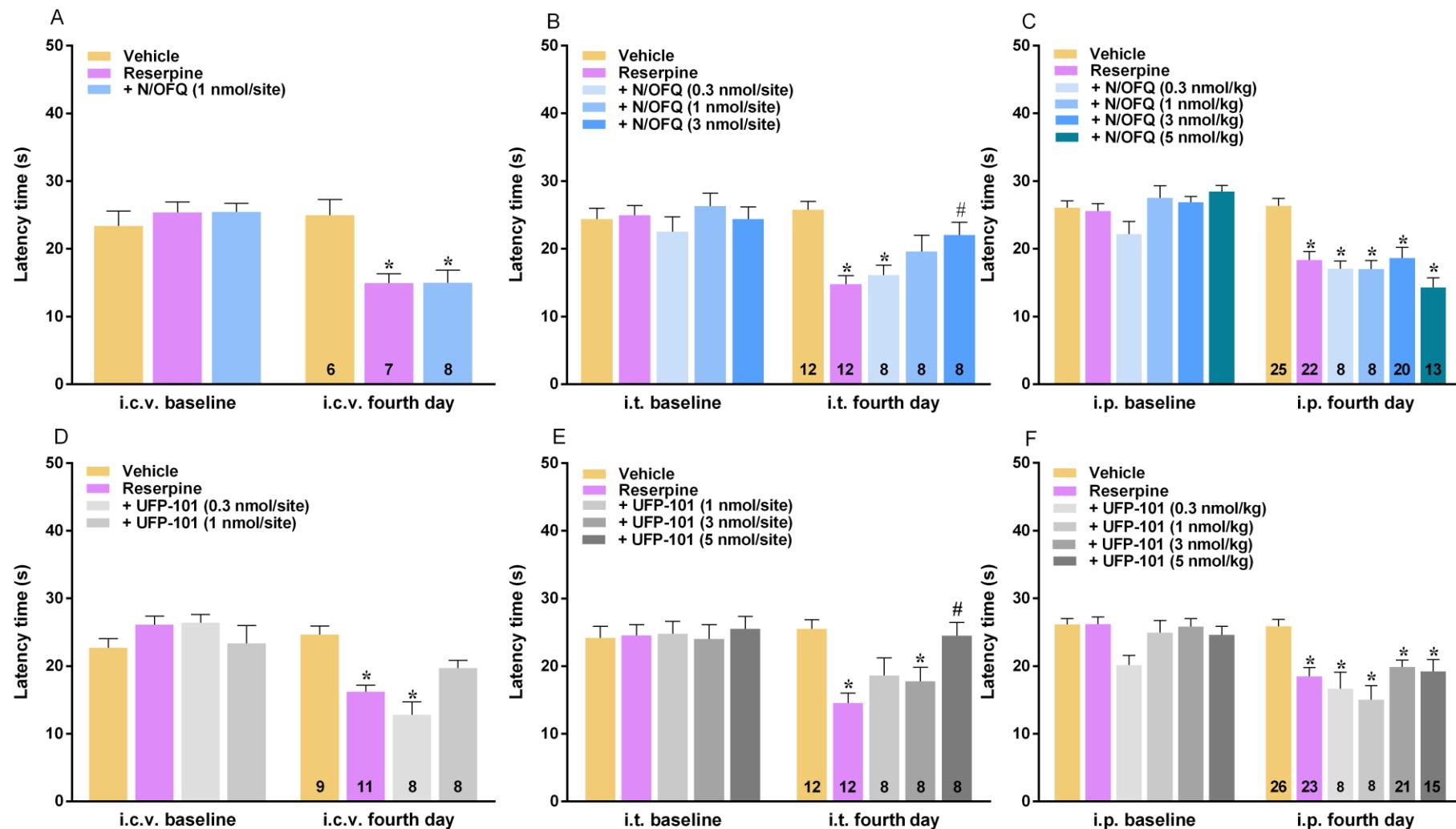
**Figure 9:** Quantitative immunohistochemistry analysis for NOP receptor in the agranular insular cortex (A-C), the lumbar spinal cord (D-F) and the dorsal root ganglia (DRG; G-I) of vehicle- and reserpine-treated mice. The samples were collected at the fourth day after the onset of reserpine administration. NOP receptor immunopositivity was quantified in the regions corresponding to the laminas I to VI (D) of the spinal cord. Representative images for NOP receptor immunolabelling in the agranular insular cortex of the vehicle/saline control (B) or reserpine-treated group (C); in the spinal cord of the vehicle/saline control (E) or reserpine-treated group (F); and the DRG of the vehicle/saline control (H) or reserpine-treated group (I). The DRG images were acquired in 200-x magnification. The schematic representations of brain and lumbar spinal cord were captured in  $\times 8$  and  $\times 32$  magnification, respectively. Red continuous lines delimit the regions of interest analysed in the brain and lumbar spinal cord. Scale bar (—) represents 2 mm, 1 mm and 100  $\mu\text{m}$ , for brain, spinal cord and DRG, respectively. \* $p < 0.05$  when compared to the vehicle/saline control group. Statistical analysis was performed by Student  $t$  test. (A and D)  $n = 5$  mice/group; (G)  $n = 5\text{-}6$  mice/group.

**Figure 10:** Effects of the repeated treatment with SB-612111 on fibromyalgia-like symptoms in reserpine-treated mice. Effects of the repeated administration of SB-612111 (6.6  $\mu\text{mol/kg}$ ; intraperitoneal) on hind paw mechanical allodynia (A), latency time in response to heat stimulation (B), immobility time (C), and on plus maze parameters (G-I). Effects of the repeated administration of SB-612111 (2.2, 6.6 and 22  $\mu\text{mol/kg}$ ; intraperitoneal) on the rotating time (D), grasping strength (E), and latency to fall (F). All the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). SB-612111 was dosed for three consecutive days, 30 min after reserpine injection. At the fourth day, mice also received SB-612111, 30 min before evaluations. Each bar represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis (F, H), one-way (C, D, G and I) or two-way (A, B and E) ANOVA followed by Bonferroni's *post hoc* test. (A-C, G-I)  $n = 10$  mice/group; (D-F)  $n = 8\text{-}14$  mice/group.

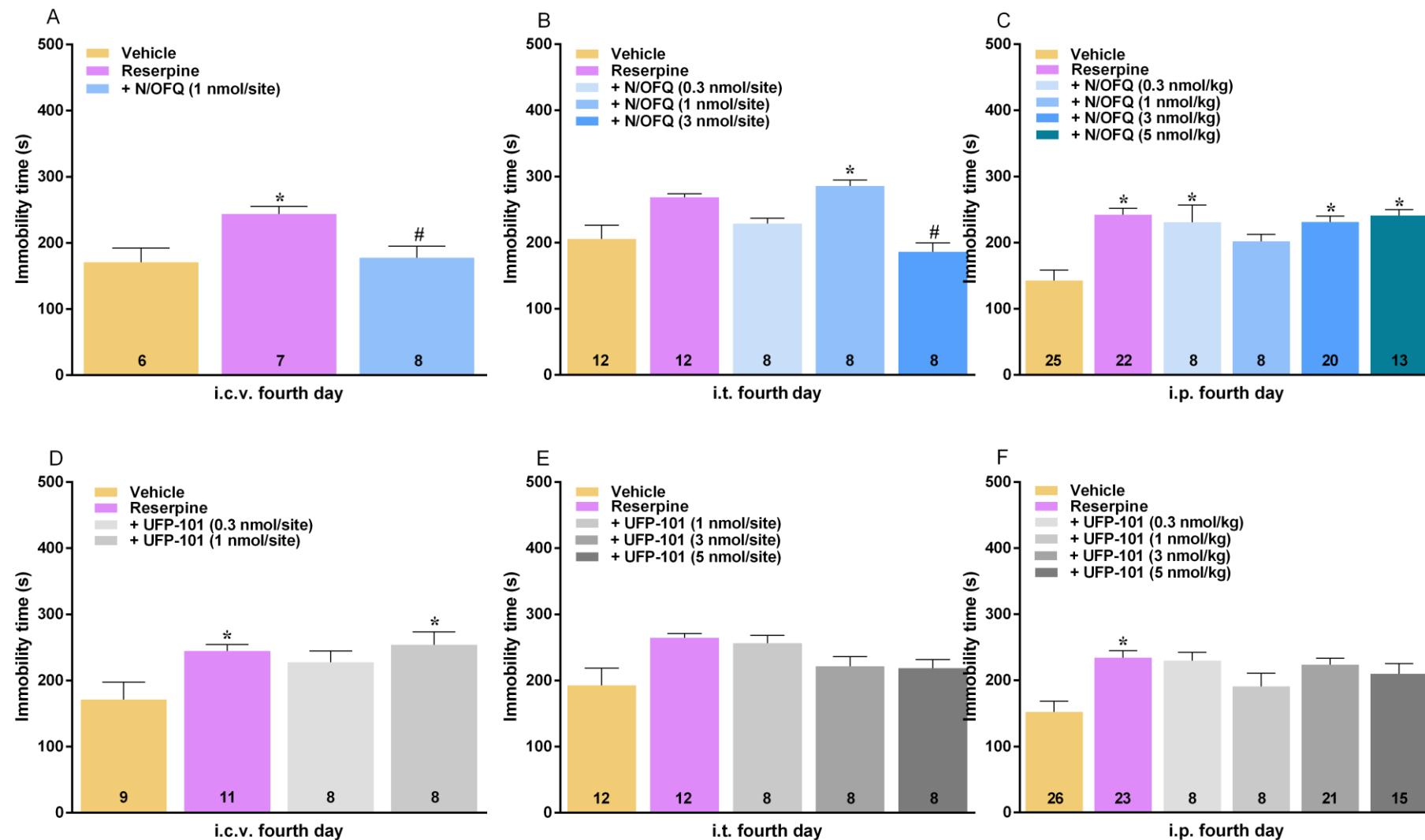
## Figures and Legends



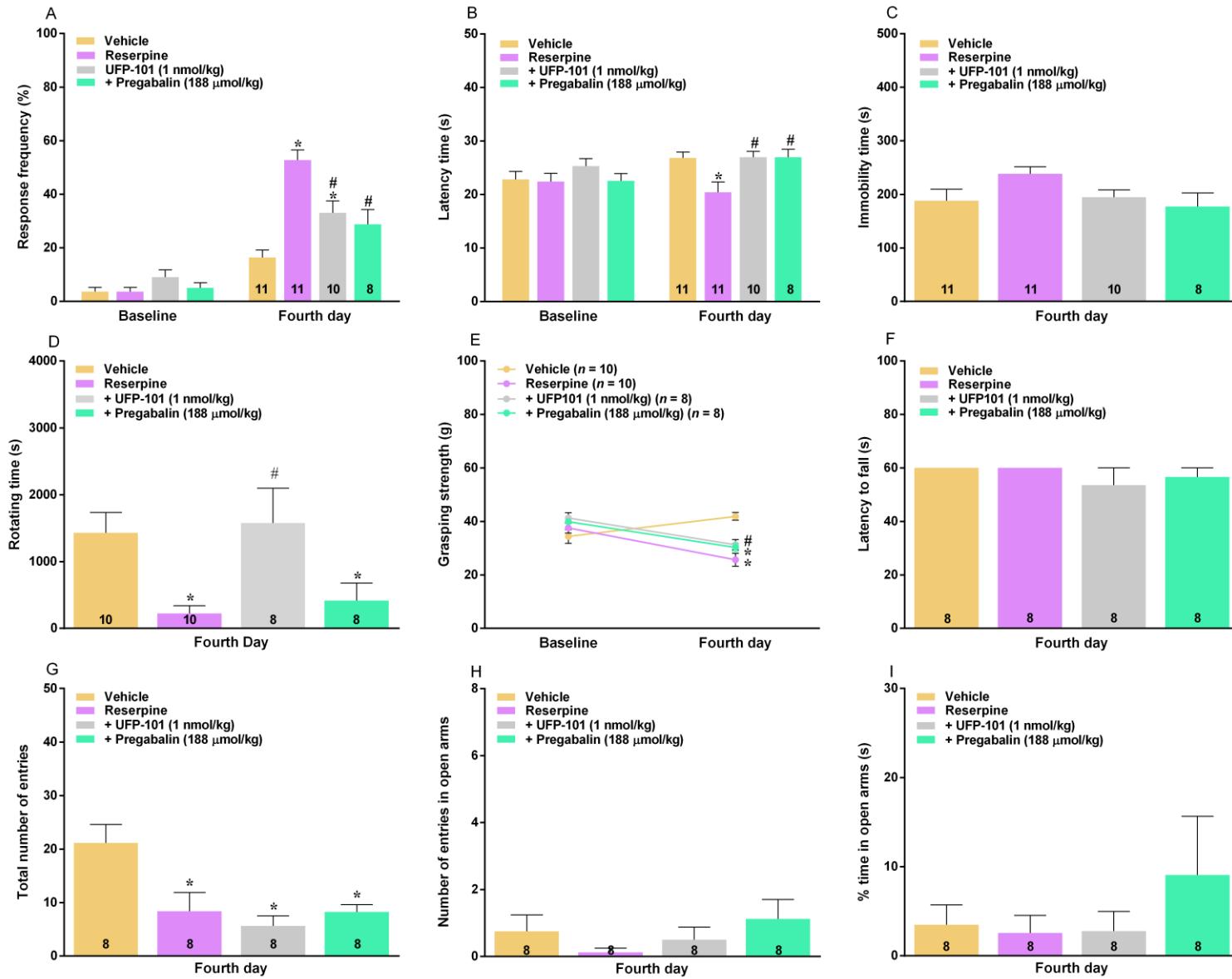
**Figure 1:** Dose-related effects of N/OFQ or UFP-101 on the mechanical hypersensitivity in the mouse model of fibromyalgia induced by reserpine. Effects of acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v.; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on hindpaw withdrawal threshold (response frequency in percentage) to tactile stimulation. The mechanical hypersensitivity was assessed by using the Von Frey filaments before (baseline), and at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 15 min (i.c.v. and i.t.) or 30 min (i.p.) before the behavioural evaluations. Each column represents the mean  $\pm$  SEM. \* $p$  < 0.05 when compared to the control vehicle/saline group, indicating the development of mechanical allodynia; # $p$  < 0.05 when compared to the reserpine/saline group. Statistical analysis was performed by two-way ANOVA, followed by Bonferroni's *post hoc* test (A-F). (A)  $n$  = 6-8 mice/group; (B)  $n$  = 8-12 mice/group; (C)  $n$  = 8-25 mice/group; (D)  $n$  = 8-11 mice/group; (E)  $n$  = 8-12 mice/group; (F)  $n$  = 8-26 mice/group.



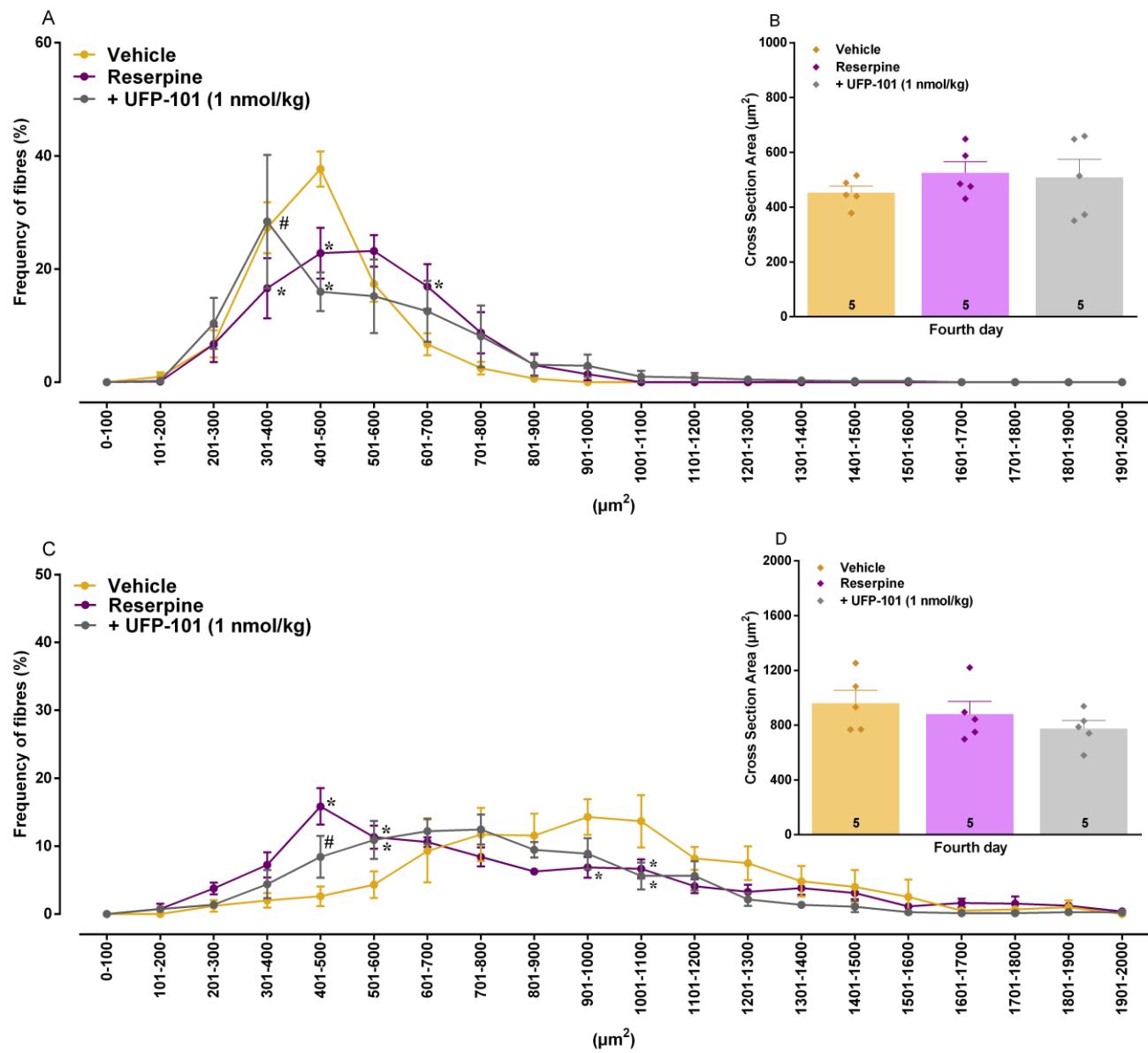
**Figure 2:** Dose-related effects of N/OFQ or UFP-101 on the thermal hypersensitivity in reserpine-treated mice. Effects of acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v.; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on the latency time (s) in response to hot thermal stimulation. Thermal hypersensitivity was assessed in the hot-plate test, at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 15 min (i.c.v. and i.t.) or 30 min (i.p.) before the behavioural evaluations. Each column represents the mean  $\pm$  SEM. \* $p$  < 0.05 when compared to the control vehicle/saline group, indicating the development of thermal sensitivity; # $p$  < 0.05 when compared to the reserpine/saline group. Statistical analysis was performed by two-way ANOVA followed by Bonferroni's *post hoc* test (A-F). (A)  $n$  = 6-8 mice/group; (B)  $n$  = 8-12 mice/group; (C)  $n$  = 8-25 mice/group; (D)  $n$  = 8-11 mice/group; (E)  $n$  = 8-12 mice/group; (F)  $n$  = 8-26 mice/group.



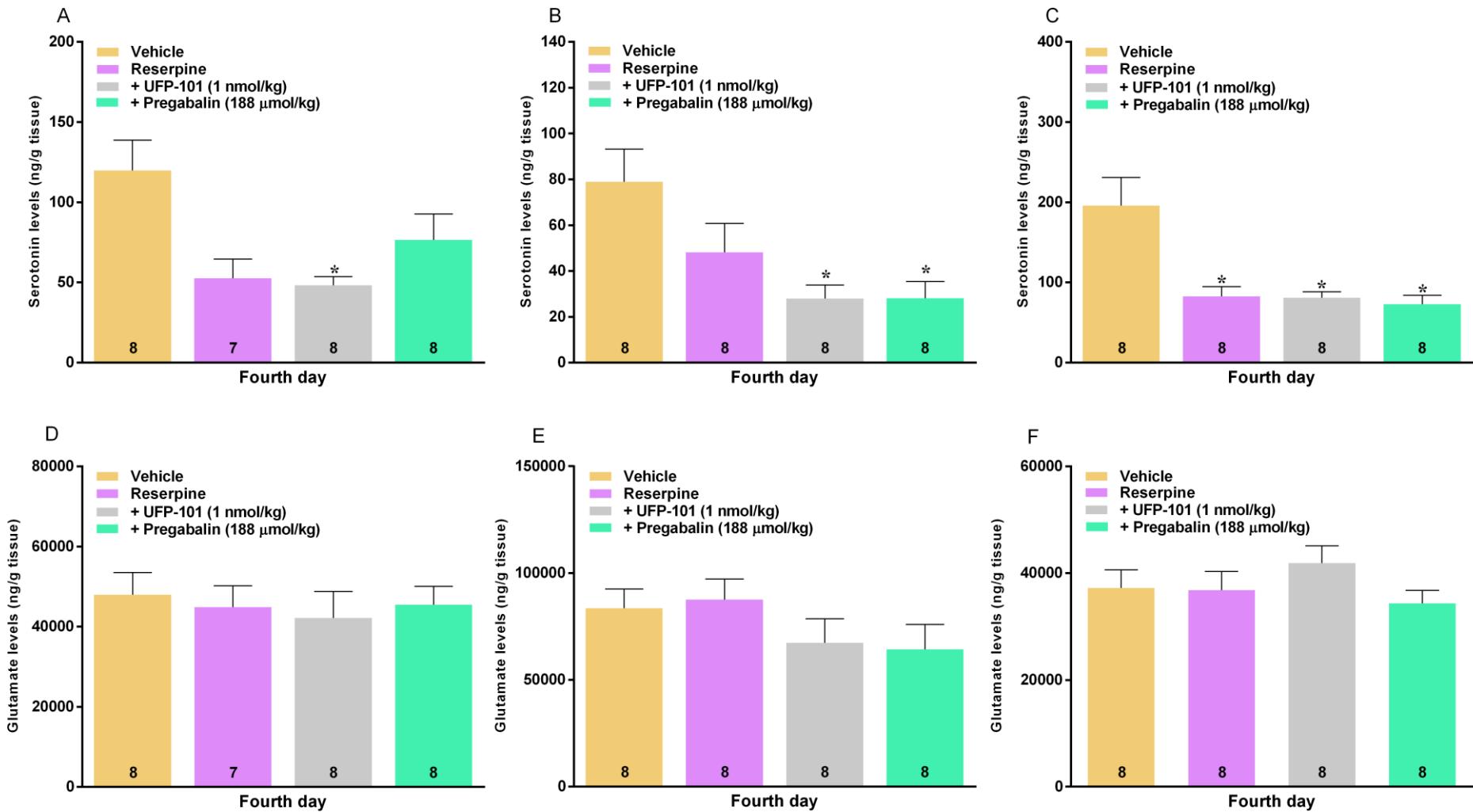
**Figure 3:** Dose-related effects of the acute treatment with N/OFQ (A-C) or UFP-101 (D-F), dosed by intracerebroventricular (i.c.v; A and D), intrathecal (i.t.; B and E) or intraperitoneal (i.p.; C and F) routes, on depression-like behaviour in reserpine-treated mice submitted to the forced swimming test. The immobility time (s) was assessed at fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). The NOPr ligands were administered in a single dose, 30 min before the behavioural evaluations. Each column represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group, indicating the development of depressive behaviour; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (B, C, E and F) or one-way ANOVA followed by Bonferroni's *post hoc* test (A and D). (A)  $n = 6\text{-}8$  mice/group; (B)  $n = 8\text{-}12$  mice/group; (C)  $n = 8\text{-}25$  mice/group; (D)  $n = 8\text{-}11$  mice/group; (E)  $n = 8\text{-}12$  mice/group; (F)  $n = 8\text{-}26$  mice/group.



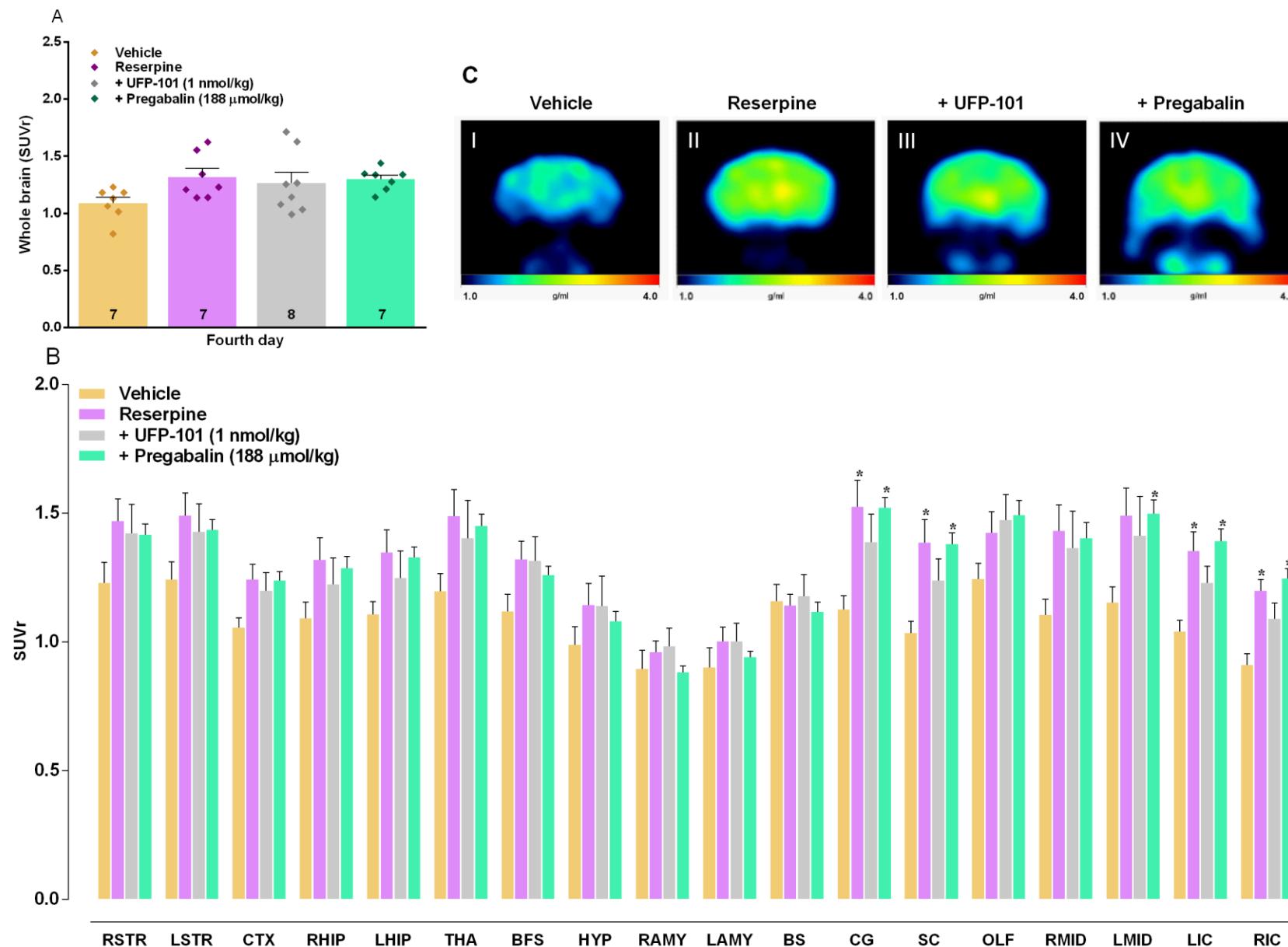
**Figure 4:** Effects of the repeated treatment with UFP-101 or pregabalin on painful-, fatigue-, depressive- and anxiety-like behaviours in reserpine-treated mice. The effects of both drugs were also assessed on the fatigue and grip strength in the fibromyalgia model elicited by reserpine. Effects of the repeated administration of UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188 µmol/kg, intraperitoneal), on hind paw mechanical allodynia (A), latency time (s) in response to hot thermal stimulation (B), immobility time in the forced swimming test (C), rotating time (D), grasping strength (E), latency to fall (F), and on plus maze parameters (G-I). All the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). UFP-101 or pregabalin were dosed for three consecutive days, 30 min after reserpine injection. At the fourth day, mice also received UFP-101 or pregabalin, 30 min before evaluations. Each column represents the mean ± SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (D, F, H and I), one-way (C and G) or two-way (A, B and E) ANOVA followed by Bonferroni's *post hoc* test. (A-C)  $n = 8\text{-}11$  mice/group; (D-E)  $n = 8\text{-}10$  mice/group; (F-I)  $n = 8$  mice/group.



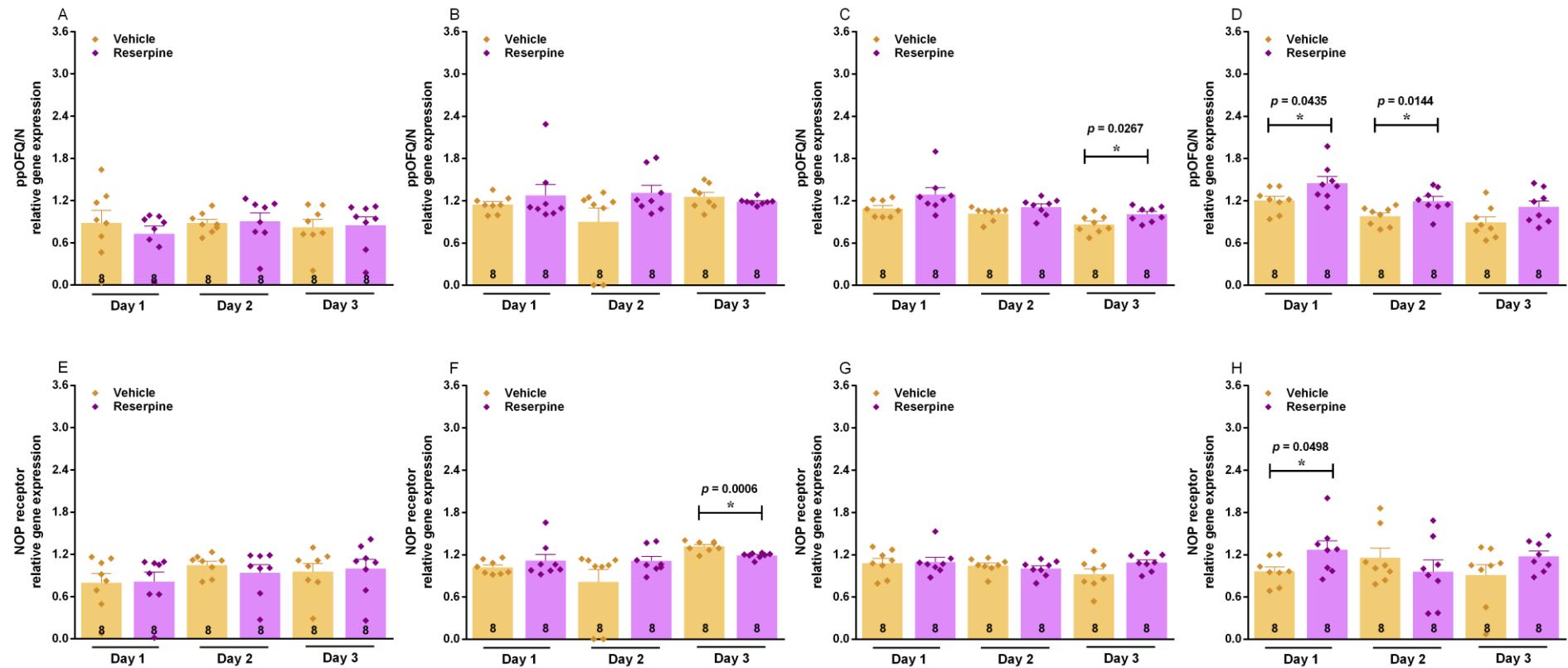
**Figure 5:** Histological analysis of masseter (A and B) and gastrocnemius (C and D) muscles of reserpine-treated mice. Effects of intraperitoneal (i.p.) repeated treatment with UFP-101 (1 nmol/kg) on fibre size distribution (A and C) and mean fibre cross-sectional area (B and D). UFP-101 was administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days) and 30 min before the muscle collection, at the fourth day. Each point or column represent the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  compared to reserpine/saline group. Statistical analysis was performed by one-way (cross sectional area) or two-way (frequency of fibres) ANOVA followed by Bonferroni's *post hoc* test. (A-D)  $n = 5$  mice/group.



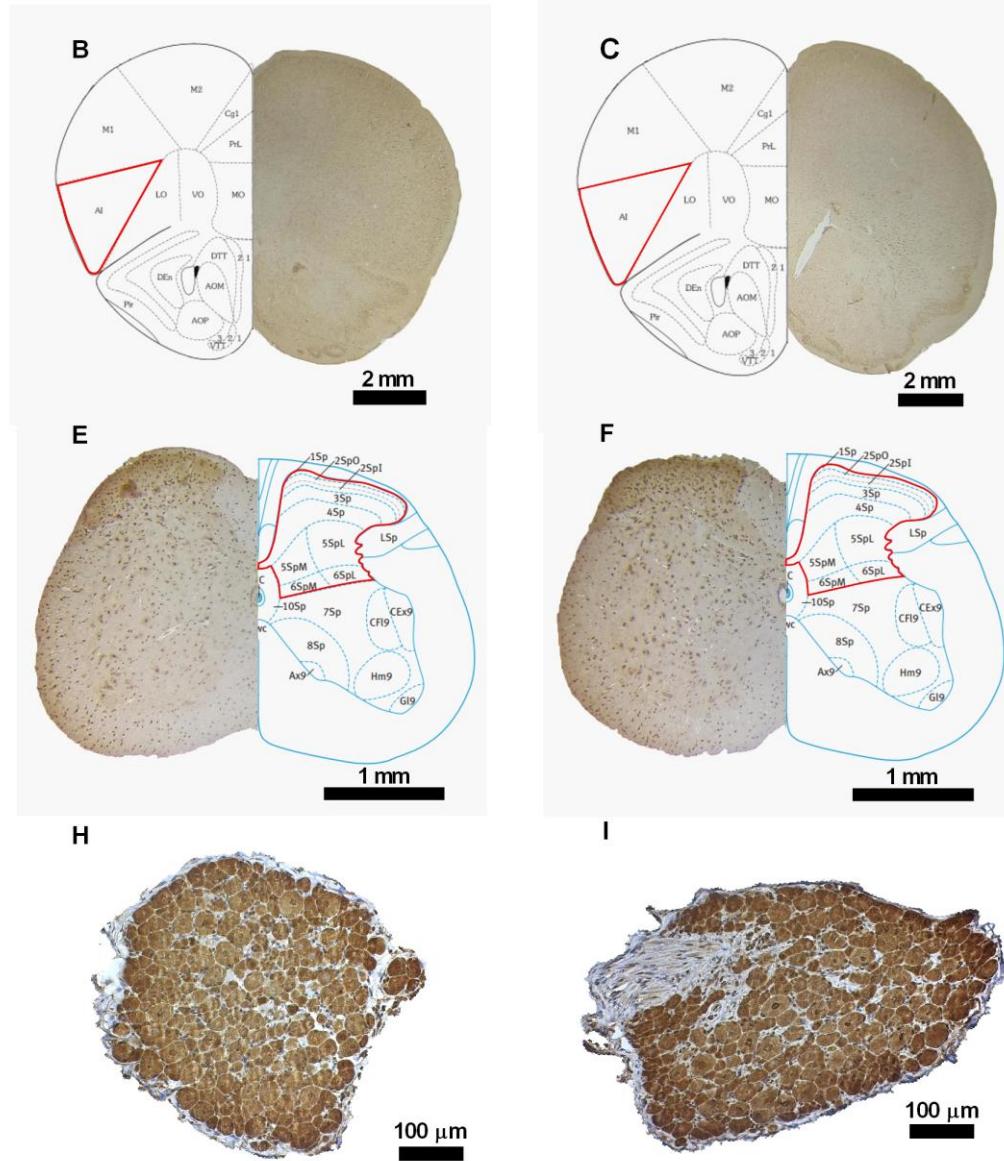
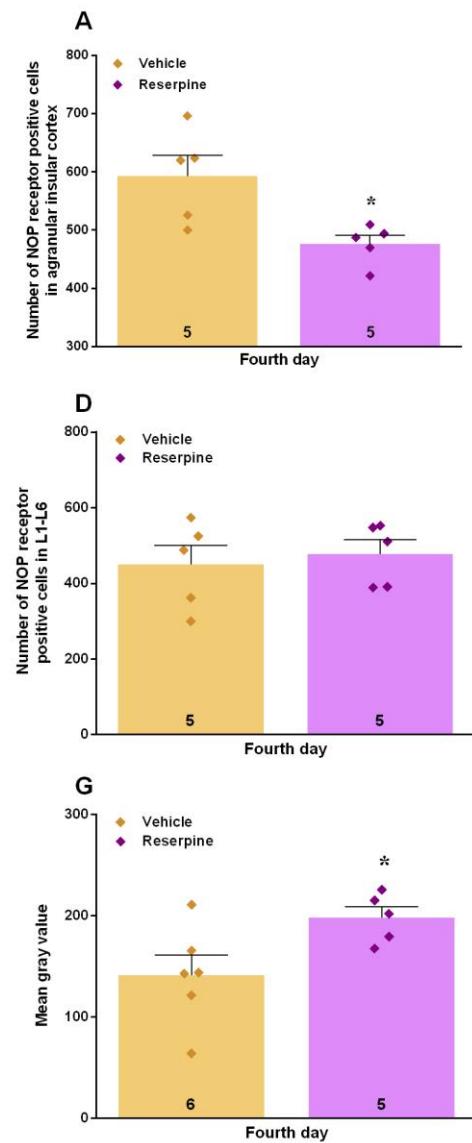
**Figure 6:** Serotonin and glutamate levels in prefrontal cortex (A and D), thalamus/hypothalamus (B and E) and lumbar spinal cord (C and F) of reserpine-treated mice. UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188 µmol/kg, intraperitoneal) were administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection at the fourth day. Serotonin (A-C) and glutamate (D-F) levels are expressed in ng/g tissue. Each bar represents the mean ± SEM. \* $p < 0.05$  when compared to the control vehicle/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (A and C) or one-way ANOVA followed by Bonferroni's *post hoc* test (B, D, E and F). (A and D)  $n = 7-8$  mice/group; (B, C, E and F)  $n = 8$  mice/group.



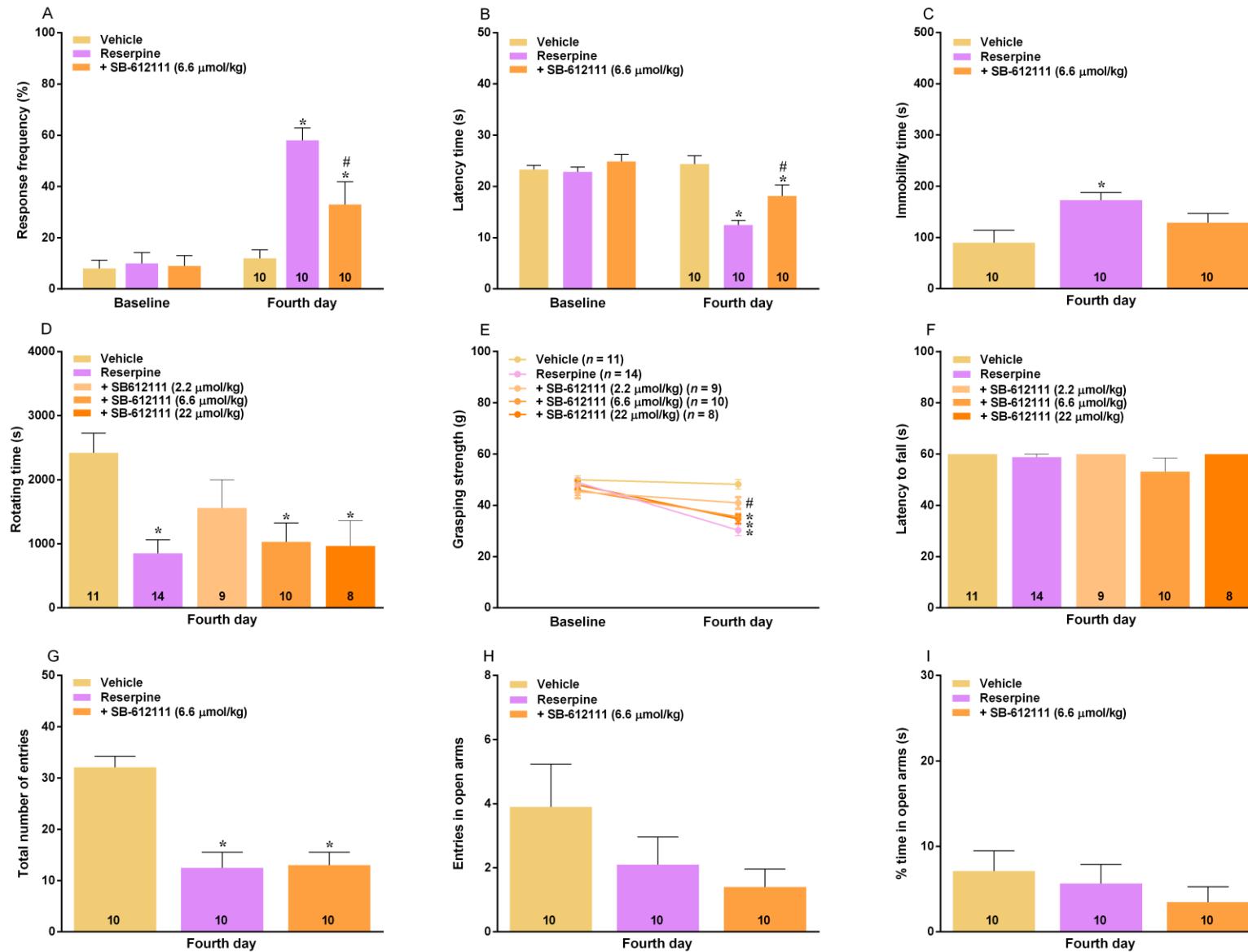
**Figure 7:** Effects of the repeated treatment with UFP-101 or pregabalin on the [<sup>18</sup>F]-FDG hypermetabolism in the whole brain (A) or in several brain structures (B) in the fibromyalgia-like model induced by reserpine in mice. UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188 µmol/kg, intraperitoneal) were administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection at the fourth day. Representative images of the coronal plane of the vehicle control group (C1), the reserpine-treated group (C2), the group treated with UFP-101 (C3), or the group treated with pregabalin (C4). Each column represents the mean ± SEM. \**p* < 0.05 when compared to the control vehicle/saline group. Differences in the standardised uptake value ratio (SUVr) per areas of brain and in whole brain were determined by one-way (A) or two-way ANOVA (B), followed by Bonferroni's *post hoc* tests, respectively. (A-B) *n* = 7-8 mice/group. Abbreviations: RSTR, right striatum region; LSTR, left striatum region; CTX, cortex; RHIP, right hippocampal region; LHIP, left hippocampal region; THA, thalamus; BFS, basal forebrain/septum; HYP, hypothalamus; RAMY, right amygdala; LAMY, left amygdala; BS, brainstem; CG, cingulate gyrus; SC, superior colliculus; OLF, olfactory areas; RMID, right midbrain; LMID, left midbrain; LIC, left inferior colliculus; RIC, right inferior colliculus.



**Figure 8:** The ppN/OFQ (A-D) and NOP receptor (E-H) mRNA expression was measured by RT-qPCR in prefrontal cortex (A and E), thalamus/hypothalamus (B and F), lumbar spinal cord (C and G) and masseter muscle (D and H) tissues, at days 1, 2 and 3 after the administration of reserpine (0.25 mg/kg; subcutaneous). Each scatter dot plot represents the mean  $\pm$  SEM of 6-8 samples. \* $p < 0.05$  when compared to the control vehicle/saline group. Statistical analysis was performed by unpaired Student  $t$  test.



**Figure 9:** Quantitative immunohistochemistry analysis for NOP receptor in the agranular insular cortex (A-C), the lumbar spinal cord (D-F) and the dorsal root ganglia (DRG; G-I) of vehicle- and reserpine-treated mice. The samples were collected at the fourth day after the onset of reserpine administration. NOP receptor immunopositivity was quantified in the regions corresponding to the laminas I to VI (D) of the spinal cord. Representative images for NOP receptor immunolabelling in the agranular insular cortex of the vehicle/saline control (B) or reserpine-treated group (C); in the spinal cord of the vehicle/saline control (E) or reserpine-treated group (F); and the DRG of the vehicle/saline control (H) or reserpine-treated group (I). The DRG images were acquired in 200-x magnification. The schematic representations of brain and lumbar spinal cord were captured in  $\times 8$  and  $\times 32$  magnification, respectively. Red continuous lines delimit the regions of interest analysed in the brain and lumbar spinal cord. Scale bar (—) represents 2 mm, 1 mm and 100  $\mu\text{m}$ , for brain, spinal cord and DRG, respectively. \* $p < 0.05$  when compared to the vehicle/saline control group. Statistical analysis was performed by Student  $t$  test. (A and D)  $n = 5$  mice/group; (G)  $n = 5\text{-}6$  mice/group.



**Figure 10:** Effects of the repeated treatment with SB-612111 on fibromyalgia-like symptoms in reserpine-treated mice. Effects of the repeated administration of SB-612111 (6.6  $\mu\text{mol/kg}$ ; intraperitoneal) on hind paw mechanical allodynia (A), latency time in response to heat stimulation (B), immobility time (C), and on plus maze parameters (G-I). Effects of the repeated administration of SB-612111 (2.2, 6.6 and 22  $\mu\text{mol/kg}$ ; intraperitoneal) on the rotating time (D), grasping strength (E), and latency to fall (F). All the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). SB-612111 was dosed for three consecutive days, 30 min after reserpine injection. At the fourth day, mice also received SB-612111, 30 min before evaluations. Each bar represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis (F, H), one-way (C, D, G and I) or two-way (A, B and E) ANOVA followed by Bonferroni's *post hoc* test. (A-C, G-I)  $n = 10$  mice/group; (D-F)  $n = 8-14$  mice/group.

## Supporting Information

**Supplementary Table 1.** Effects of the acute treatment with N/OFQ or UFP-101 on the locomotor activity and anxiety parameters in reserpine-treated mice. The values represent the mean  $\pm$  SEM.

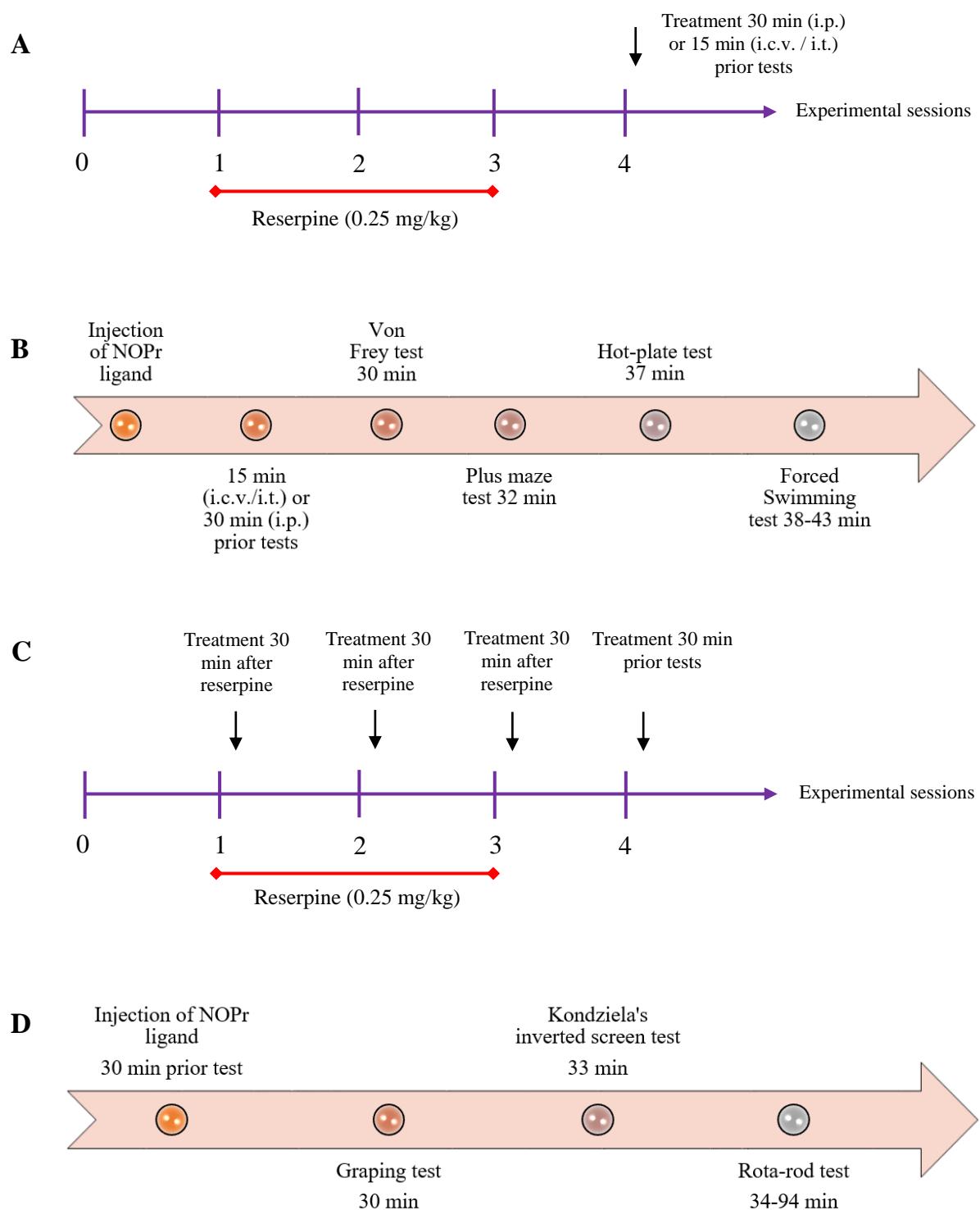
I.C.V.	Vehicle	Reserpine	N/OFQ (1 nmol/site)			
Total number entries	20 $\pm$ 1.1	8.4 $\pm$ 3.6 *	8 $\pm$ 1.4 *			
Entries in open arms	0.8 $\pm$ 0.5	1.3 $\pm$ 1	1.5 $\pm$ 0.8			
%Time in open arms	1.6 $\pm$ 0.9	12.9 $\pm$ 9.6	3.8 $\pm$ 1.7			
I.C.V.	Vehicle	Reserpine	UFP-101 (0.3 nmol/site)	UFP-101 (1 nmol/site)		
Total number entries	16.3 $\pm$ 3.1	6.7 $\pm$ 2.6 *	8.6 $\pm$ 2	3.9 $\pm$ 1.5 *		
Entries in open arms	0.8 $\pm$ 0.3	0.4 $\pm$ 0.4	1.1 $\pm$ 0.5	1.5 $\pm$ 0.8		
%Time in open arms	1.7 $\pm$ 0.5	0.4 $\pm$ 0.2	3.5 $\pm$ 1.5	14 $\pm$ 9.2		
I.T.	Vehicle	Reserpine	N/OFQ (0.3 nmol/site)	N/OFQ (1 nmol/site)	N/OFQ (3 nmol/site)	
Total number entries	23.2 $\pm$ 1.7	11.6 $\pm$ 3.1 *	10.5 $\pm$ 2.4 *	14.4 $\pm$ 3.3	8.1 $\pm$ 1.6 *	
Entries in open arms	1.1 $\pm$ 0.6	2 $\pm$ 0.9	1.2 $\pm$ 0.4	2.9 $\pm$ 1.5	1.9 $\pm$ 0.7	
%Time in open arms	3.3 $\pm$ 1.6	9.2 $\pm$ 4.7	3 $\pm$ 1.1	6.2 $\pm$ 3.8	1.7 $\pm$ 0.7	
I.T.	Vehicle	Reserpine	UFP-101 (1 nmol/site)	UFP-101 (3 nmol/site)	UFP-101 (5 nmol/site)	
Total number entries	20.2 $\pm$ 2.6	11.4 $\pm$ 3.2	13.6 $\pm$ 3.3	10.5 $\pm$ 1.5	13.2 $\pm$ 4.2	
Entries in open arms	0.5 $\pm$ 0.3	1.9 $\pm$ 0.9	2 $\pm$ 1	0.9 $\pm$ 0.4	5.6 $\pm$ 1.8	
%Time in open arms	1.9 $\pm$ 0.9	4.9 $\pm$ 2.1	3.1 $\pm$ 1.5	4.2 $\pm$ 1.8	35.7 $\pm$ 12.9	
I.P.	Vehicle	Reserpine	N/OFQ (0.3 nmol/kg)	N/OFQ (1 nmol/kg)	N/OFQ (3 nmol/kg)	N/OFQ (5 nmol/kg)
Total number entries	22.5 $\pm$ 2	14 $\pm$ 2 *	6.4 $\pm$ 2.3 *	13.1 $\pm$ 2.8	8.4 $\pm$ 1.9 *	14.4 $\pm$ 1.8
Entries in open arms	2.1 $\pm$ 0.5	2.6 $\pm$ 0.7	0.9 $\pm$ 0.9	1.4 $\pm$ 0.6	0.9 $\pm$ 0.3	2.8 $\pm$ 0.7
%Time in open arms	3.4 $\pm$ 1	6.7 $\pm$ 1.5	3.2 $\pm$ 2.8	4.3 $\pm$ 2	2 $\pm$ 1	7 $\pm$ 2
I.P.	Vehicle	Reserpine	UFP-101 (0.3 nmol/kg)	UFP-101 (1 nmol/kg)	UFP-101 (3 nmol/kg)	UFP-101 (5 nmol/kg)
Total number entries	23.8 $\pm$ 2	12.1 $\pm$ 2.1 *	13 $\pm$ 1.8	4.2 $\pm$ 1.5 *	16.2 $\pm$ 4	12.7 $\pm$ 2.2 *
Entries in open arms	2.4 $\pm$ 0.6	2.3 $\pm$ 0.7	0.1 $\pm$ 0.1	1.2 $\pm$ 0.6	3.3 $\pm$ 1.3	3.1 $\pm$ 0.5
%Time in open arms	4.7 $\pm$ 1.4	6.2 $\pm$ 1.6	0.4 $\pm$ 0.2	8 $\pm$ 4.4	9.3 $\pm$ 2.9	12 $\pm$ 5.4

N/OFQ and UFP-101 were dosed by intracerebroventricular (i.c.v.), intrathecal (i.t.) or intraperitoneal (i.p.) routes, at the fourth day after the onset of fibromyalgia induction, 30 min before the plus maze test. \* $p$  < 0.05 when compared to the control vehicle/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test, or by one-way ANOVA followed by Bonferroni's *post hoc* test. (N/OFQ i.c.v. treatment)  $n$  = 6-8 mice per group; (UFP-101 i.c.v. treatment)  $n$  = eight mice per group; (N/OFQ i.t. treatment)  $n$  = 8-10 mice per group; (UFP-101 i.t. treatment)  $n$  = 7-10 mice per group; (N/OFQ i.p. treatment)  $n$  = 8-17 mice per group; (UFP-101 i.p. treatment)  $n$  = 8-18 mice per group.

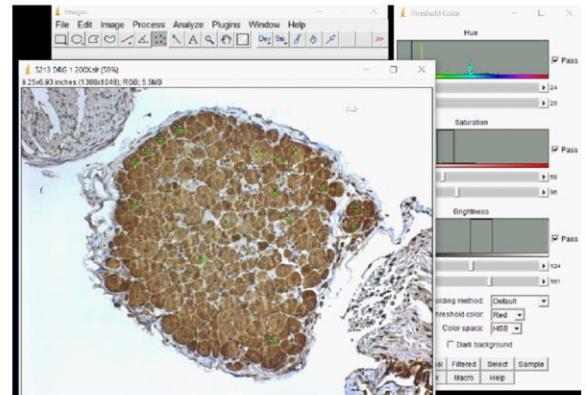
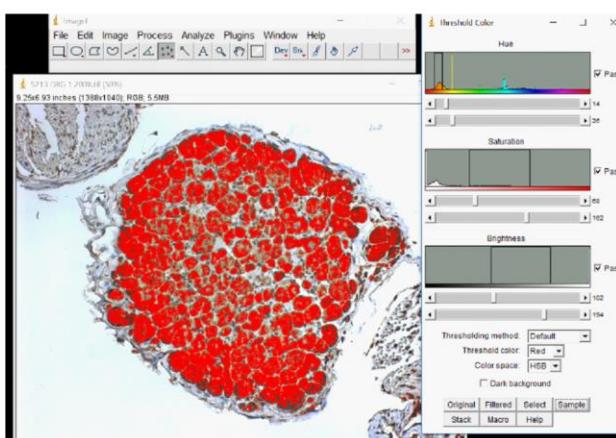
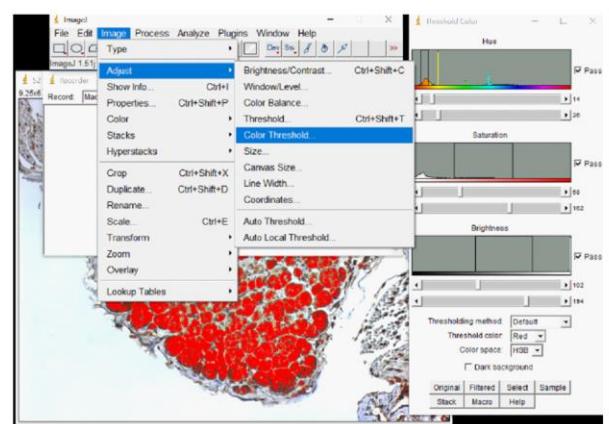
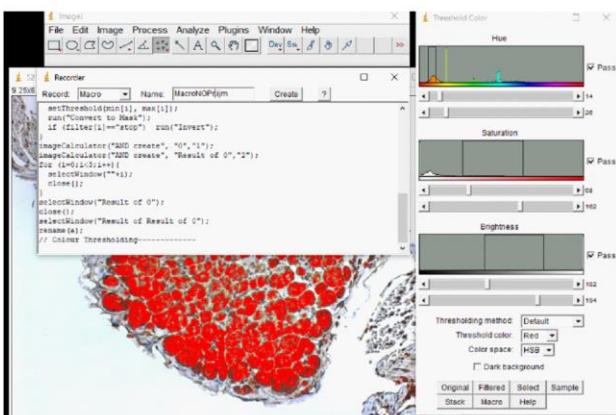
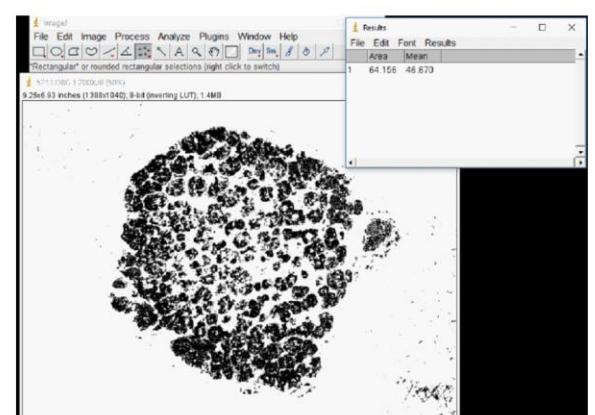
**Supplementary Table 2.** Cytokine levels (TNF, IL-1 $\beta$  and IL-10) in brain, spinal cord and masseter muscle (pg/100 mg tissue), or in serum (pg/ml) in the fibromyalgia model induced by reserpine in mice. The values represent the mean  $\pm$  SEM.

<b>Brain</b>	<b>TNF</b>	<b>IL-10</b>	<b>IL-1<math>\beta</math></b>
Vehicle	1573 $\pm$ 159.7	1430 $\pm$ 195.8	209.4 $\pm$ 43.4
Reserpine	1599 $\pm$ 142.9	1426 $\pm$ 217.6	210.2 $\pm$ 58.5
UFP-101 (1 nmol/kg)	1576 $\pm$ 233.6	1297 $\pm$ 132.8	18 $\pm$ 9
Pregabalin (188 $\mu$ mol/kg)	5707 $\pm$ 299.4*	2522 $\pm$ 155*	692 $\pm$ 103.5 <sup>#</sup>
<b>Spinal cord</b>	<b>TNF</b>	<b>IL-10</b>	<b>IL-1<math>\beta</math></b>
Vehicle	10600 $\pm$ 2115	3958 $\pm$ 138.9	2934 $\pm$ 176
Reserpine	11602 $\pm$ 2077	5193 $\pm$ 609	2636 $\pm$ 381.1
UFP-101 (1 nmol/kg)	13463 $\pm$ 1902	4212 $\pm$ 371.2	2966 $\pm$ 266.1
Pregabalin (188 $\mu$ mol/kg)	13051 $\pm$ 1157	4485 $\pm$ 292.2	2338 $\pm$ 264.7
<b>Masseter</b>	<b>TNF</b>	<b>IL-10</b>	<b>IL-1<math>\beta</math></b>
Vehicle	22769 $\pm$ 2601	4442 $\pm$ 556.2	1090 $\pm$ 144.6
Reserpine	23173 $\pm$ 2745	4207 $\pm$ 387.7	1357 $\pm$ 225.4
UFP-101 (1 nmol/kg)	20900 $\pm$ 2496	4221 $\pm$ 393.2	1118 $\pm$ 146.8
Pregabalin (188 $\mu$ mol/kg)	16922 $\pm$ 2698	3569 $\pm$ 436.8	1189 $\pm$ 110
<b>Serum</b>	<b>TNF</b>	<b>IL-10</b>	<b>IL-1<math>\beta</math></b>
Vehicle	† ND	† ND	2569 $\pm$ 151
Reserpine	† ND	† ND	1856 $\pm$ 460.3
UFP-101 (1 nmol/kg)	† ND	† ND	1767 $\pm$ 444
Pregabalin (188 $\mu$ mol/kg)	† ND	† ND	2615 $\pm$ 65.2

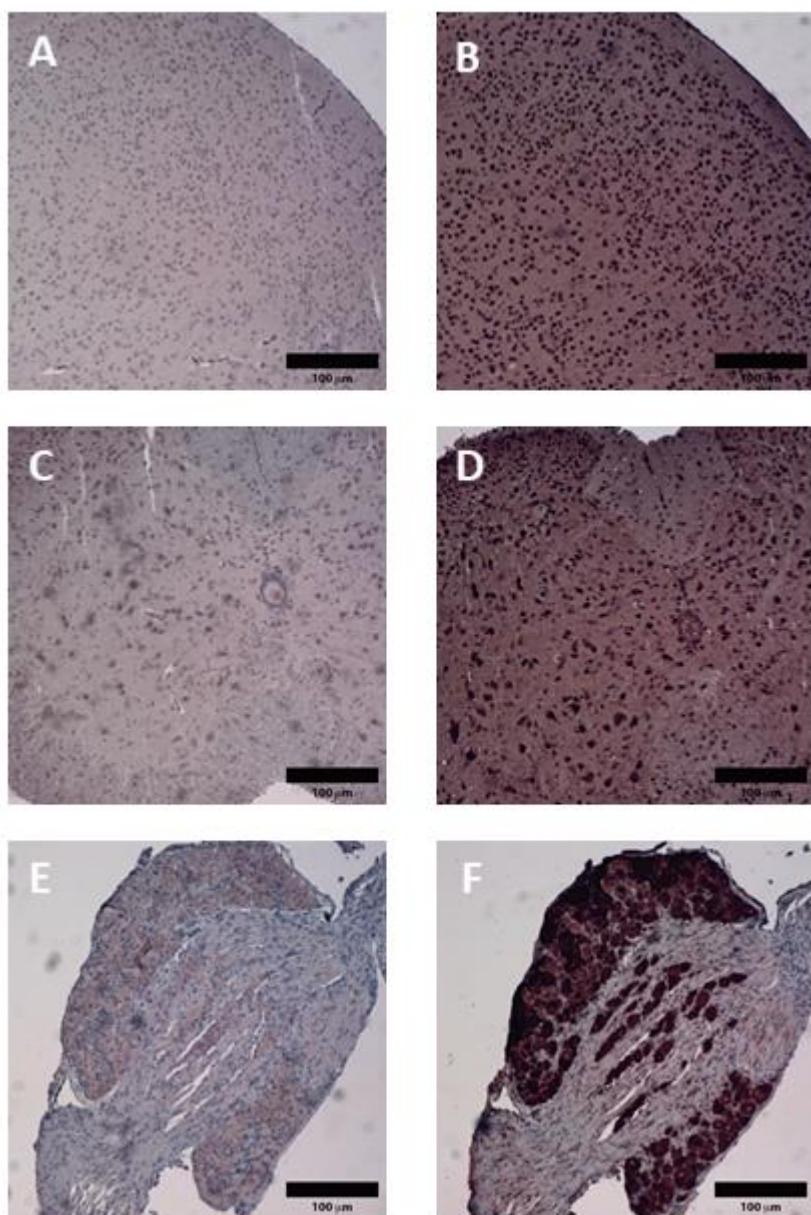
UFP-101 (1 nmol/kg, intraperitoneal) or pregabalin (188  $\mu$ mol/kg, intraperitoneal) were administered for three consecutive days, 30 min after daily reserpine injection. On the fourth day, mice also received UFP-101 or pregabalin, 30 min before of sample collection. \* $p$  < 0.05 when the pregabalin group was compared to the other groups. <sup>#</sup> $p$  < 0.05 when the pregabalin group was compared to the UFP-101-treated group. The statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test or by one-way ANOVA followed by Bonferroni's *post hoc* test.  $n$  = 5-6 mice per group. † ND = not detectable.



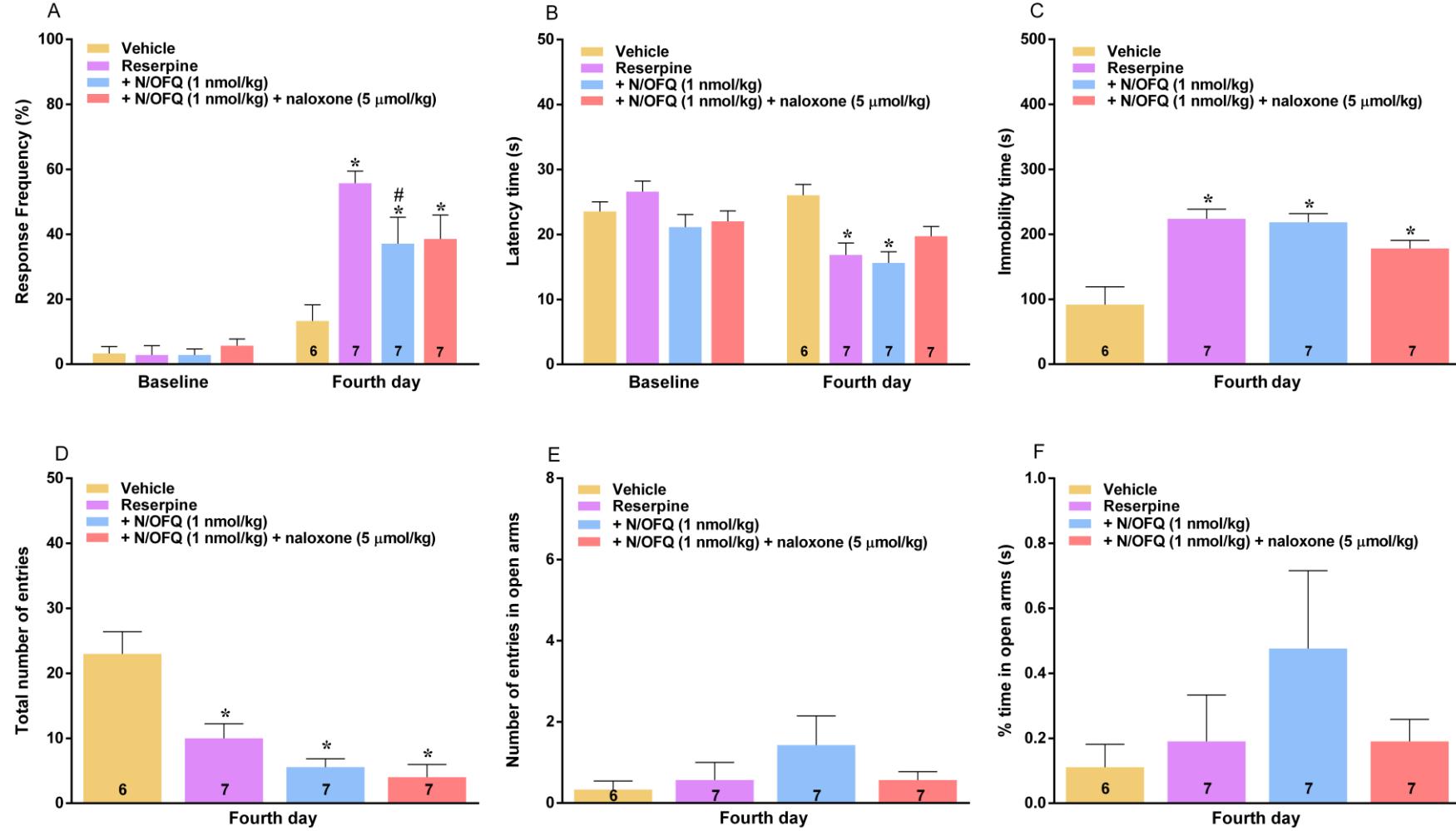
**Supplementary Figure 1:** Schedule of acute treatment with N/OFQ or UFP-101, administered 15 min (intracerebroventricular, i.c.v. and intrathecal, i.t.) or 30 min (intraperitoneal, i.p.) prior behavioural tests, at the fourth day (Panel A). Timeline for the behavioural tests after acute (N/OFQ or UFP-101) or repeated (UFP-101 or SB-612111) treatment (Panel B). Schedule of repeated treatment by i.p. injection of UFP-101 or SB-612111 administered once a day, for 4 consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days) and 30 min prior behavioural tests, at the fourth day (Panel C). Timeline for the fatigue-related tests after the repeated treatment with UFP-101 or SB-612111 (Panel D).

**A****B****C****D****E****F**

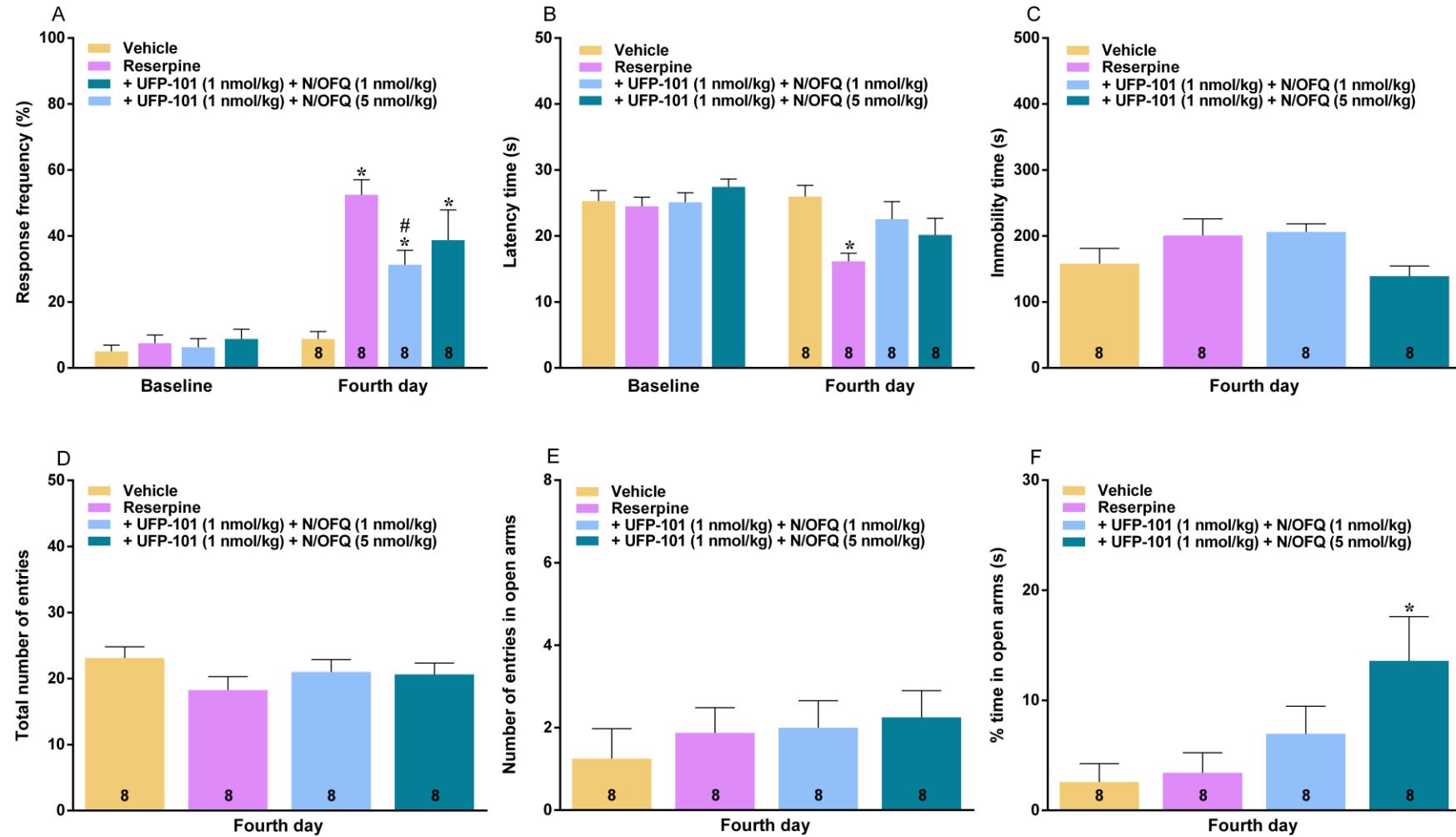
**Supplementary Figure 2:** Representative DRG images showing the creation of the macro used for the analysis of NOPr immunopositivity by ImageJ Software. (A) An image reference of negative control (vehicle-treated mice) was used for the macro creation. (B) The dark-to-medium brown areas (considered as immunopositive neurons for NOPr) were selected (16 small green multi-points) to determine the pixels for the macro creation. (C) Sampling the range of colours from the selected regions. (D) Determination of colour threshold following the next steps: (i) Plugins → macros → record; (ii) Image → adjust→ colour threshold. (E) Creation of macro “MacroNOPr”: Save and install macro in Plugins → macros. (F) Resulting black-and-white image after the analysis of the RGB image. The small window on the right shows the value of the mean grey value of the black regions. The macro “MacroNOPr” was used for analysis of the other images as a reference macro.



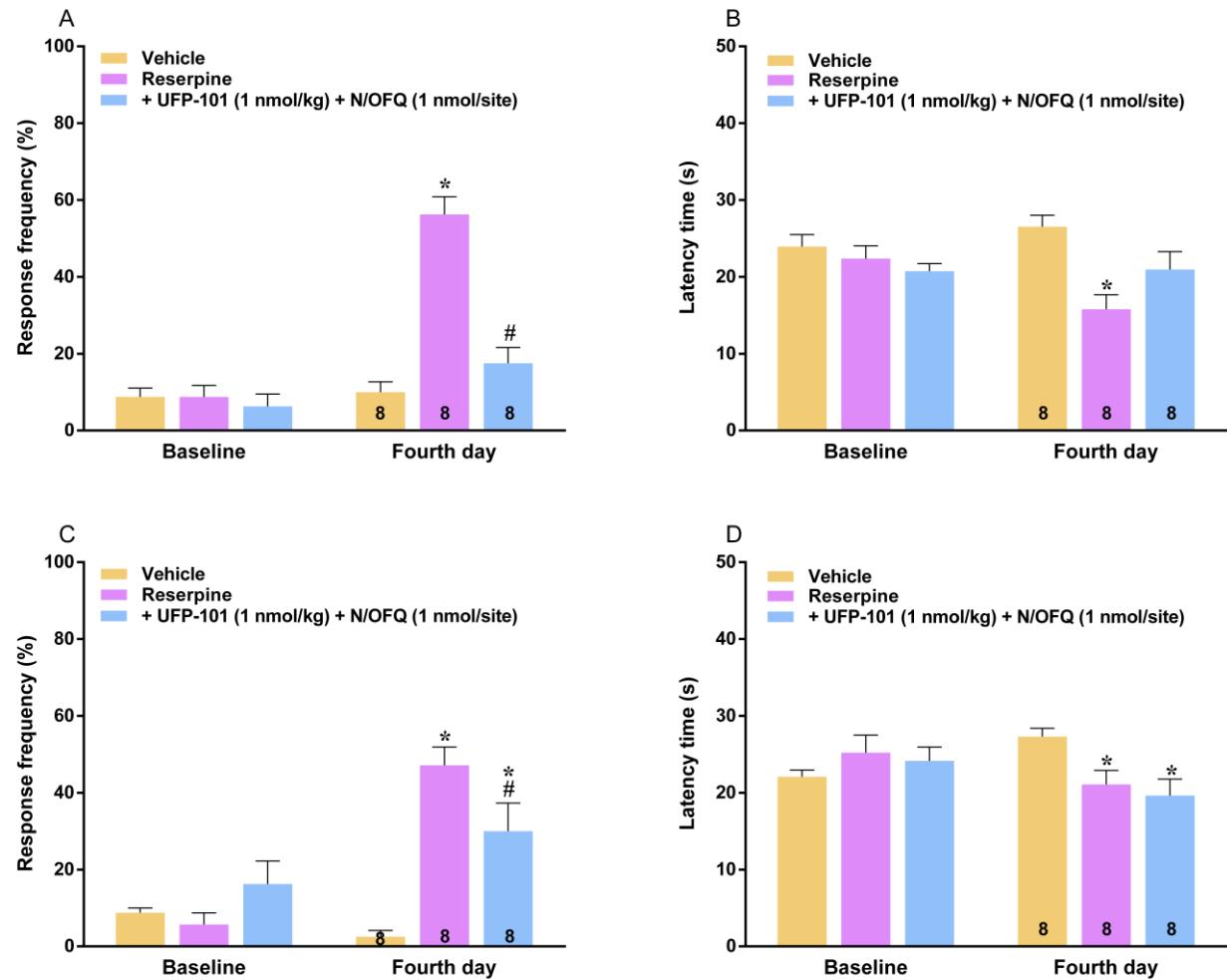
**Supplementary Figure 3:** Immunohistochemistry analysis to confirm the selectivity of the anti-NOPr antibody (A-F). Representative images for NOP receptor immunolabelling in the brain, spinal cord or DRG slides, with (A, C and E) or without (B, D and F) the co-incubation of the internal antigen with the primary antibody. The schematic representations were captured in  $\times 40$  magnification. Scale bar (—) represents 100  $\mu\text{m}$



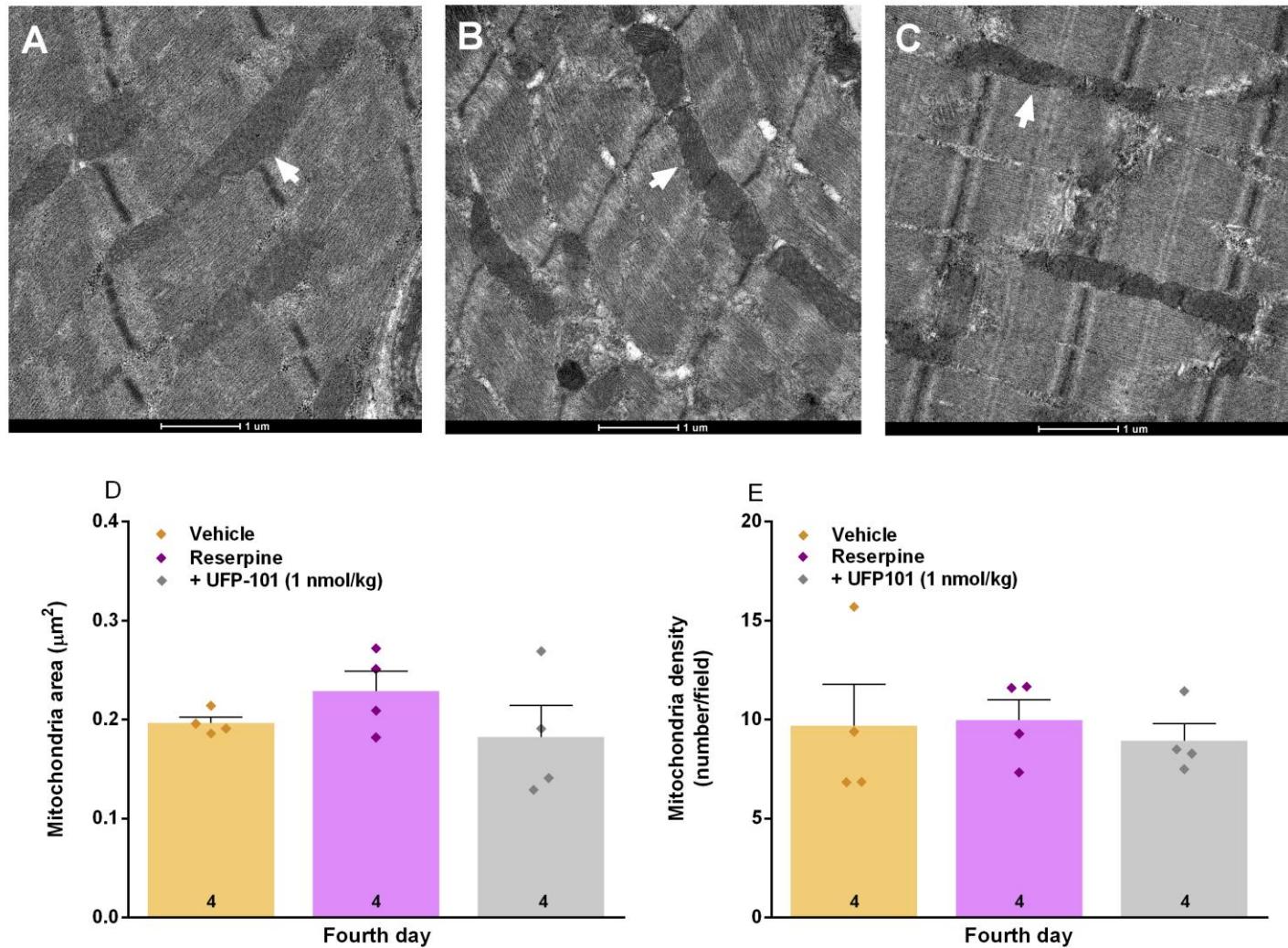
**Supplementary Figure 4:** Effects of N/OFQ alone or after the pre-treatment with naloxone on painful-, depressive- and anxiety-like behaviours in reserpine-treated mice. Effects of N/OFQ (1 nmol/kg) administered acutely, by intraperitoneal (i.p.) route, alone or after the pre-treatment with naloxone (5  $\mu$ mol/kg, dosed i.p.; 5 min before the agonist treatment), on hind paw mechanical allodynia (A), latency time in response to the hot thermal stimulation (B), immobility time (C), and on plus maze parameters (D-F). All of the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). Each bar represents the mean  $\pm$  SEM. \* $p$  < 0.05 when compared to the control vehicle/saline group;  $^{\#}p$  < 0.05 when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis (E and F), one-way (C and D) or two-way (A and B) ANOVA followed by Bonferroni's *post hoc* test. (A-F)  $n$  = 6-7 mice per group



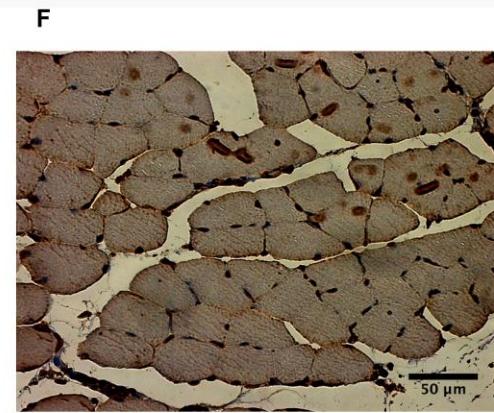
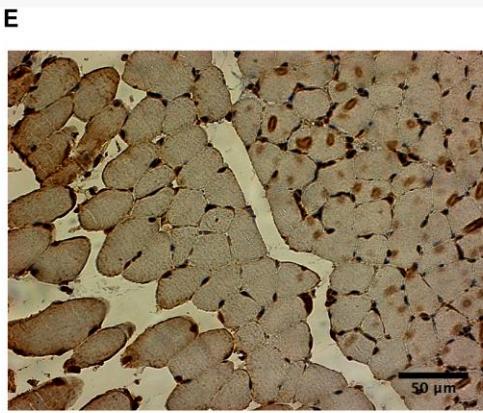
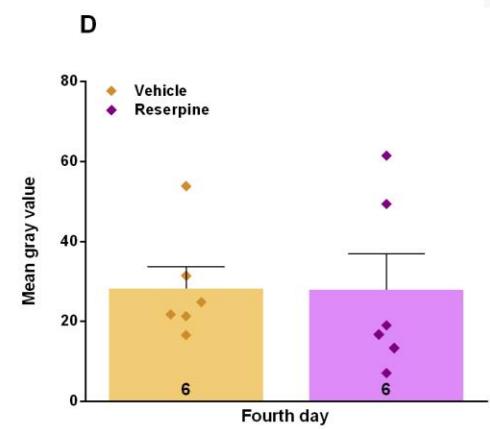
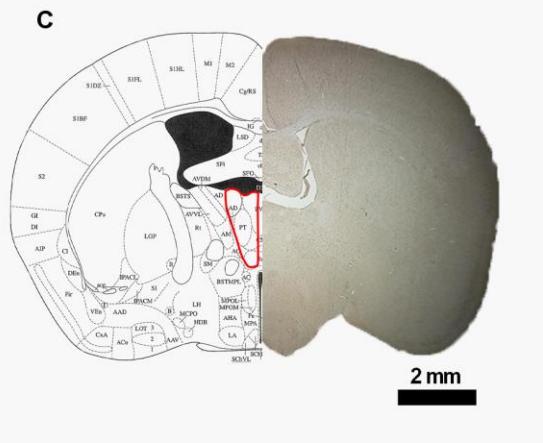
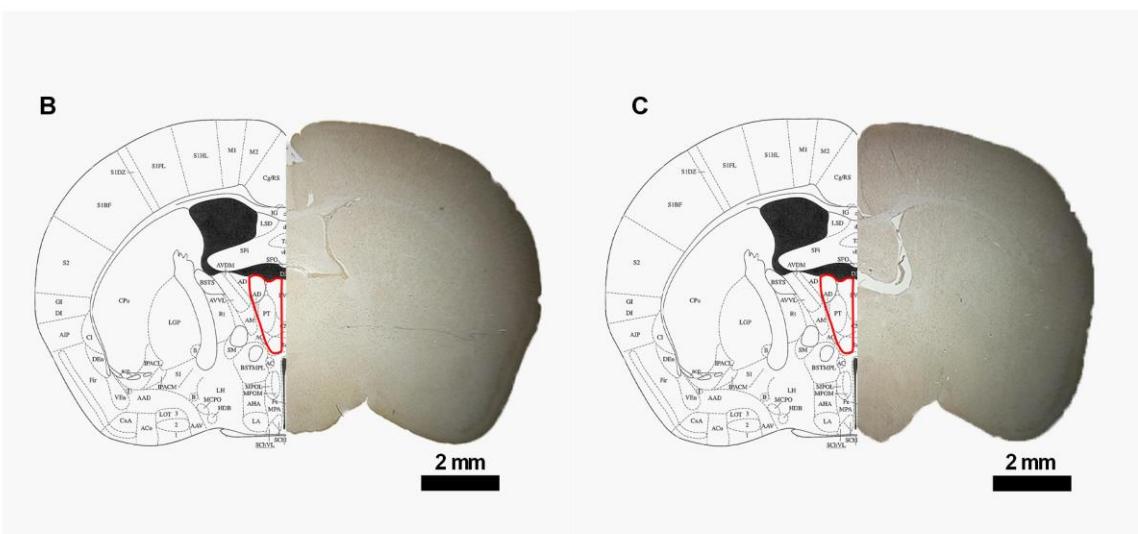
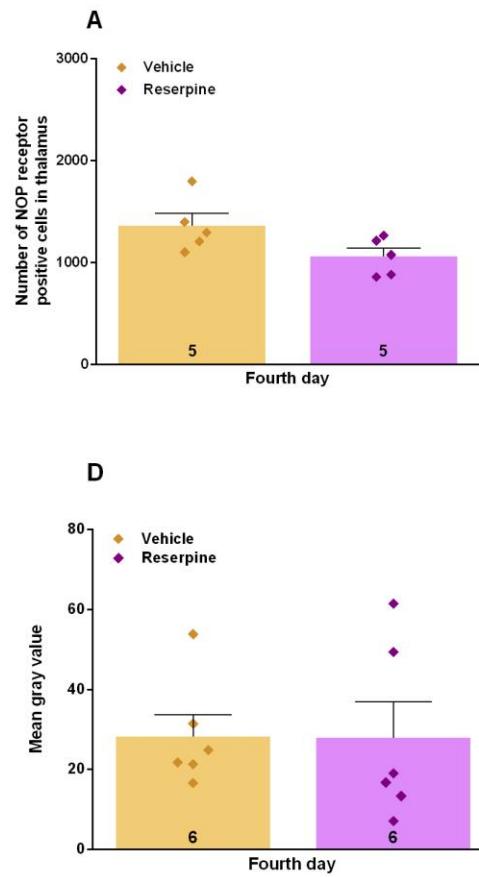
**Supplementary Figure 5:** Effects of UFP-101 combined with N/OFQ, both dosed i.p., on painful-, depressive- and anxiety-like behaviours in reserpine-treated mice. Effects of UFP-101 (1 nmol/kg) administered acutely, by intraperitoneal (i.p.) route, in combination with N/OFQ (1 nmol/kg or 5 nmol/kg, i.p.), on hind paw mechanical allodynia (A), latency time in response to hot thermal stimulation (B), immobility time (C) and on plus maze parameters (D-F). All of the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). Each column represents the mean ± SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by one-way (C-F) or two-way (A and B) ANOVA followed by Bonferroni's *post hoc* test. (A-F)  $n =$  eight mice per group.



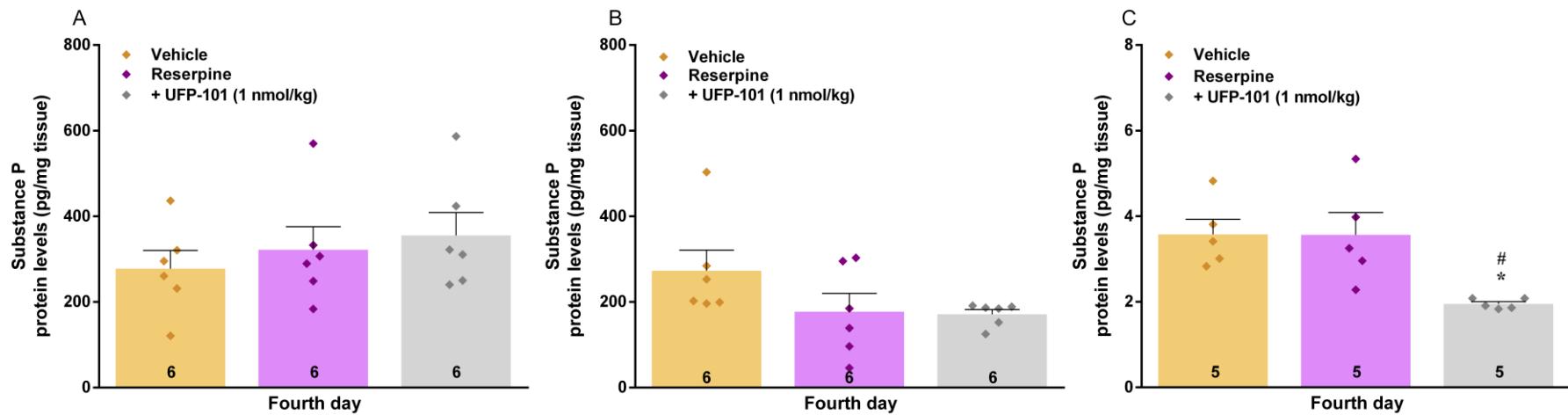
**Supplementary Figure 6:** Effects of systemic treatment with UFP-101 combined with N/OFQ (i.c.v. or i.t.) on mechanical and thermal nociception in reserpine-treated mice. Effects of UFP-101 (1 nmol/kg) administered acutely, by intraperitoneal (i.p.) route, in combination with N/OFQ (1 nmol/kg, i.c.v.; A and B) or N/OFQ (1 nmol/kg, i.t.; C and D), on hind paw mechanical allodynia (A and C) and latency time in response to hot stimulation (B and D). All of the tests were performed at the fourth day after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days). Each column represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group; # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by two-way (A-D) ANOVA followed by Bonferroni's *post hoc* test. (A-F)  $n =$  eight mice per group.



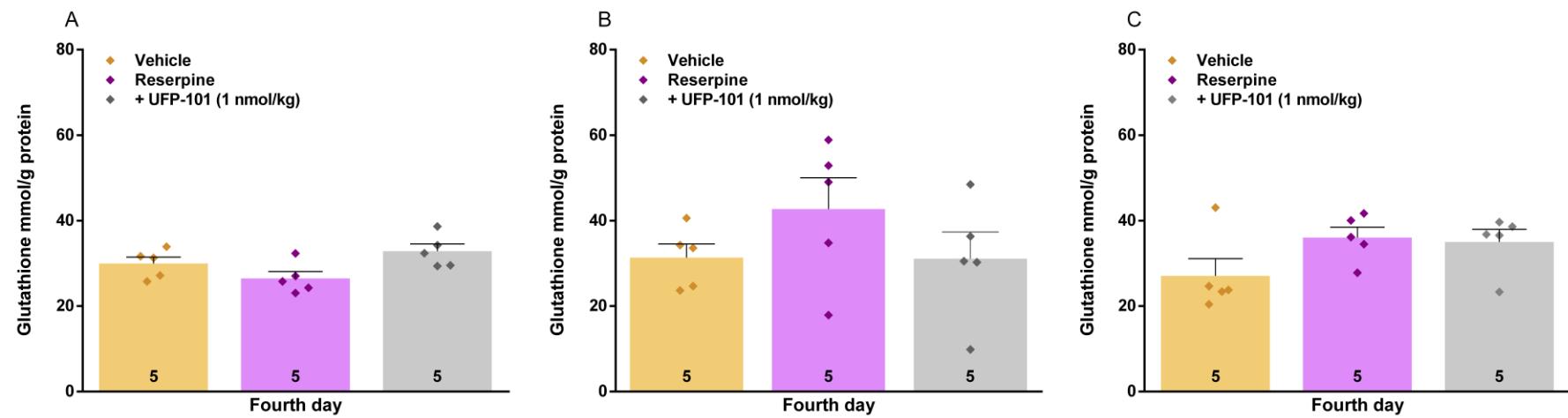
**Supplementary Figure 7:** Ultrastructural analysis of the masseter muscle in reserpine-treated mice. Representative transmission electron microscopic (TEM) images of masseter muscle in vehicle (A), reserpine (B) and UFP-101-treated mice (C). Effects of the repeated administration of UFP-101 (1 nmol/kg, intraperitoneal on mitochondrial area in  $\mu\text{m}^2$  (D) or number of mitochondria/field (E) in masseter muscle. Scale bars represent one  $\mu\text{m}$ . Original magnification x 8,900. White arrows identify the mitochondria. (A, B)  $n =$  four mice per group.



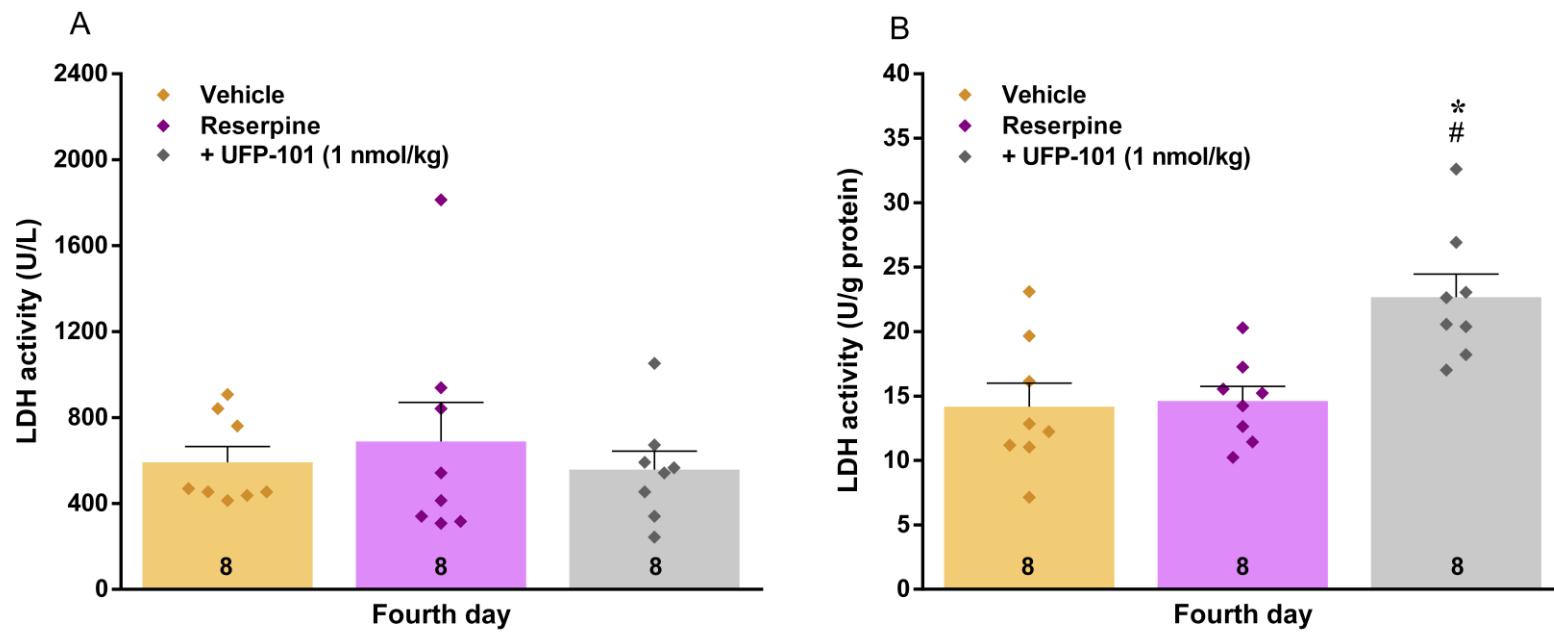
**Supplementary Figure 8:** Quantitative immunohistochemistry analysis for NOP receptor in the thalamus and masseter muscle of vehicle- and reserpine-treated mice (A-F). The samples were collected at the fourth day after the onset of reserpine administration. Representative images for NOP receptor immunolabelling in the thalamus of the vehicle/saline control (B) or reserpine-treated group (C), and in masseter of the vehicle/saline control (E) or reserpine-treated group (F). The schematic representations of brain and masseter were captured in  $\times 8$  and  $\times 400$  magnification, respectively. Red continuous lines delimit the region of interest analysed in the brain. Scale bar (—) represents 2 mm for brain and 50  $\mu\text{m}$  for muscle. Statistical analysis was performed by Student *t* test. (A-C)  $n$  = five mice per group; (D-F)  $n$  = six mice per group.



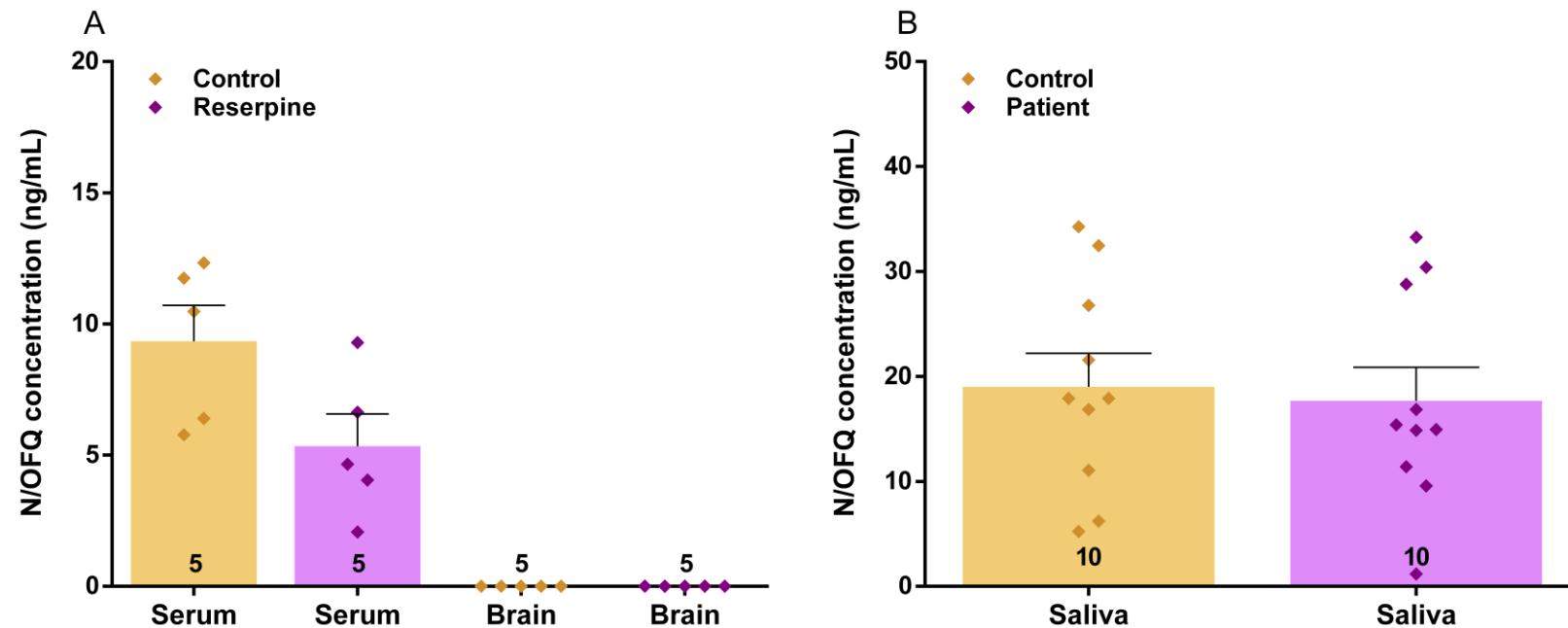
**Supplementary Figure 9:** Substance P levels in brain (A), spinal cord (B) and masseter muscle (C) of vehicle- and reserpine-treated mice. UFP-101 (1 nmol/kg, intraperitoneal) was administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before the tissue collection, at the fourth day. Each column represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group. Statistical analysis was performed by Kruskal-Wallis followed by Dunn's *post hoc* test (B and C) or by one-way ANOVA (A). (A-B)  $n = 6$  mice per group; (C)  $n = 5$  mice per group.



**Supplementary Figure 10:** Tissue concentrations of glutathione in brain (A), spinal cord (B) and masseter muscle (C) of vehicle- and reserpine-treated mice. UFP-101 (1 nmol/kg, intraperitoneal) was administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection, at the fourth day. Each column represents the mean  $\pm$  SEM. Statistical analysis was performed by one-way ANOVA. (A-C)  $n = 5$  mice per group.



**Supplementary Figure 11:** Lactate dehydrogenase (LDH) activity in serum (A) and mitochondrial extracts of masseter (B) of vehicle- and reserpine-treated mice. UFP-101 (1 nmol/kg, intraperitoneal) was administered once a day, for four consecutive days, 30 min after the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days), and 30 min before tissue collection, at the fourth day. Each column represents the mean  $\pm$  SEM. \* $p < 0.05$  when compared to the control vehicle/saline group. # $p < 0.05$  when compared to the reserpine/saline group. Statistical analysis was performed by Kruskal-Wallis (A) or one-way (B) ANOVA followed by Bonferroni's *post hoc* test. (A, B)  $n =$  eight mice per group.



**Supplementary Figure 12:** N/OFQ concentration in serum and brain samples of vehicle- and reserpine-treated mice (A), and in saliva of female control subjects and fibromyalgia patients (B). The levels of endogenous N/OFQ were measured at the fourth day after of the onset of reserpine administration (0.25 mg/kg, subcutaneous, once a day for three days) in mice. Statistical analysis was performed by Student *t* test. (A) *n* = five mice per group; (B) *n* = 10 subjects per group.

## 5 CONSIDERAÇÕES FINAIS

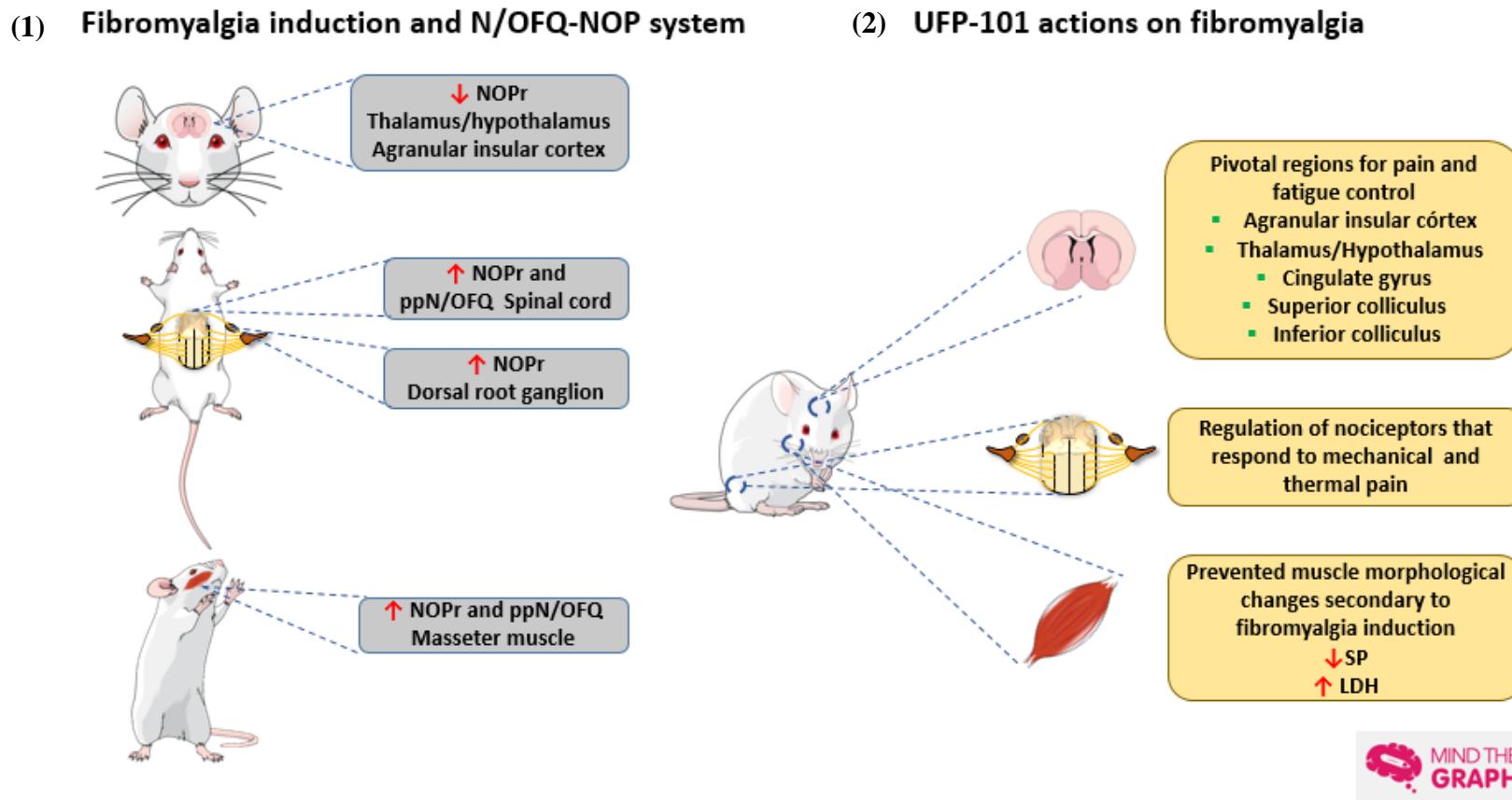
A fibromialgia é uma doença crônica que afeta mais de 2% da população ao redor do mundo. Além disto, as dores musculoesqueléticas crônicas estão no ranking das 10 principais enfermidades globais. É importante ressaltar que a dor crônica gera grandes custos e impacta negativamente na qualidade de vida do paciente, pois é acompanhada por depressão, ansiedade, problemas cognitivos e no sono, além de fadiga e perda de função física. Todas estas co-morbidades associadas, resultam na perda de produtividade e diminuição na habilidade de manter as atividades do dia-dia. Além disto, os fármacos existentes para o tratamento da fibromialgia geram em seus pacientes efeitos adversos e, muitas vezes, perda de eficácia ou tolerabilidade.

Diante deste cenário, a fibromialgia é uma doença que necessita de novos estudos para desvendar os mecanismos da sua patofisiologia e para a descoberta de possíveis tratamentos. O presente trabalho demonstrou a relação entre o sistema N/OFQ-NOPr e o modelo de fibromialgia induzido por reserpina. A modulação da expressão do RNAm da ppN/OFQ e do NOPr, além do padrão de ativação cerebral induzidos pela reserpina ocorreram em regiões importantes para o controle da dor e da fadiga (côrte insular agranular, tálamo/hipotálamo, giro do cingulado, colículos superior e inferior esquerdo e direito). Estas regiões advêm como possíveis pontos chaves para os efeitos anti-alodínico, anti-hipernociceptivo térmico e anti-fadiga encontrados para o antagonista UFP-101, na dose de 1 nmol pela via intraperitoneal. Estes achados visam caracterizar o sistema N/OFQ-NOPr nesta condição patológica. Como a causa da doença ainda é pouco estabelecida, é de extrema importância elucidar os circuitos centrais e periféricos envolvidos nos efeitos do antagonista UFP-101 do NOPr.

O efeito encontrado para o antagonista UFP-101 pode ser devido à sua ação sobre os nociceptores que respondem à dor térmica e mecânica que chegam até a medula espinhal,

regulando, por consequência, as vias ascendentes e descendentes da dor. A partir de sua ação periférica, sobre os músculos esqueléticos, este potencial pode alcançar regiões centrais, que respondem aos estímulos periféricos diminuindo a dor e a fadiga. A melhora proporcionada por UFP-101 nos sintomas de fadiga pode ser em decorrência de uma ação indireta da diminuição da dor ao nível central, levando a maior atividade motora. O antagonista UFP-101 pode estar atuando perifericamente sobre mediadores inflamatórios, neurotransmissores e/ou neuropeptídios, primordiais para a patofisiologia da fibromialgia induzida por reserpina, agindo sobre os seus receptores NOP no músculo. Consequentemente, o antagonista pode, indiretamente, modular os disparos nociceptivos a nível medular, inibindo a via ascendente da dor ou ativando a via descendente inibitória da dor.

Também é válido ressaltar que a prevenção dos sintomas de fadiga pode ser explicado por uma ação direta sobre os NOPr nas fibras musculares, modulando parâmetros bioquímicos, como a LDH e a SP. De maneira interessante, os efeitos anti-fadiga de UFP-101 se sobrepuseram aos efeitos do fármaco pregabalina, amplamente utilizado para o tratamento da fibromialgia. Somando-se a isto, estão os estudos que demonstram a eficácia de ligantes do NOPr em ensaios clínicos para o tratamento da dor neuropática e crônica. Diante deste contexto, onde pela primeira vez descrevemos o efeito de um antagonista peptídico do NOPr para o tratamento de sintomas de dor e fadiga na fibromialgia experimental, se torna claro que este ligante é promissor para futuros ensaios clínicos. Abaixo é demonstrada uma representação esquemática para os principais achados deste estudo.



Representação esquemática da relação entre o modelo da fibromialgia induzido por reserpina em camundongos e o sistema N/OFQ-NOPr e os efeitos do tratamento sistêmico com o antagonista peptídico UFP-101. (1) A injeção subcutânea (s.c.) com reserpina (0.25 mg/kg, por três dias consecutivos) modula a expressão do RNAm da ppN/OFQ e do NOPr ao nível central e periférico. (2) O tratamento repetido intraperitoneal (i.p.) com UFP-101 (1 nmol/kg) produz efeito anti-alodínico, previne a hipernocicepção térmica e reduz a fadiga em camundongos tratados com reserpina. Estes efeitos podem ser em decorrência de sua ação (direta e/ou indireta) em regiões centrais e periféricas fundamentais para o controle da dor e da fadiga. SP = substância P; LDH = lactato desidrogenase.

## 6 PERSPECTIVAS

- Investigar os efeitos do tratamento repetido com o antagonista seletivo do NOPr, UFP-101, sobre a anedonia, no modelo de fibromialgia induzido por reserpina em camundongos, através do teste de ingestão de sacarose;
- Investigar os efeitos do tratamento repetido com o antagonista seletivo do NOPr, UFP-101, sobre parâmetros de ansiedade, no modelo de fibromialgia induzido por reserpina em camundongos, utilizando outros paradigmas experimentais;
- Investigar os efeitos do tratamento repetido com o antagonista seletivo do NOPr, UFP-101, sobre a memória, no modelo de fibromialgia induzido por reserpina em camundongos, no teste de memória espacial;
- Avaliar o peptídeo relacionado ao gene da calcitonina (CGRP) em amostras de cérebro, medula e músculo, após o tratamento repetido com o antagonista seletivo do NOPr, UFP-101;
- Avaliar os níveis cerebrais, espinhais e musculares de dopamina e noradrenalina, após o tratamento repetido com o antagonista seletivo do NOPr, UFP-101;
- Avaliar e diferenciar os tipos de fibras musculares no masseter de camundongos submetidos ao modelo de fibromialgia induzido por reserpina e tratados com o antagonista seletivo do NOPr, UFP-101;
- Avaliar a expressão de MIF, MuRF e Fbxo32 no masseter de camundongos submetidos ao modelo de fibromialgia induzido por reserpina e tratados com o antagonista seletivo do NOPr, UFP-101;
- Quantificar indicadores de estresse oxidativo (catalase e superóxido dismutase) no masseter de camundongos tratados com reserpina e com o antagonista seletivo do NOPr, UFP-101;

- Quantificar os níveis de IL-8, centralmente e perifericamente, em camundongos tratados com reserpina e com o antagonista seletivo do NOPr UFP-101;
- Quantificar os níveis de  $\text{Ca}^+$  em cultivo *in vitro* de neurônios, para elucidar o possível mecanismo de ação do antagonista seletivo do NOPr, UFP-101.

## REFERÊNCIAS BIBLIOGRÁFICAS

1. Lambert DG. The nociceptin/orphanin FQ receptor: a target with broad therapeutic potential. *Nature reviews Drug discovery*. 2008;7(8):694-710.
2. Mustazza C, Pieretti S, Marzoli F. Nociceptin /Orphanin FQ Peptide (NOP) Receptor Modulators: An Update in Structure-Activity Relationships. *Current medicinal chemistry*. 2018;25(20):2353-84.
3. Mollereau C, Simons MJ, Soularue P, Liners F, Vassart G, Meunier JC, et al. Structure, tissue distribution, and chromosomal localization of the prepronociceptin gene. *Proceedings of the National Academy of Sciences of the United States of America*. 1996;93(16):8666-70.
4. Meunier JC, Mollereau C, Toll L, Suaudeau C, Moisand C, Alvinerie P, et al. Isolation and structure of the endogenous agonist of opioid receptor-like ORL1 receptor. *Nature*. 1995;377(6549):532-5.
5. Reinscheid RK, Nothacker HP, Bourson A, Ardati A, Henningsen RA, Bunzow JR, et al. Orphanin FQ: a neuropeptide that activates an opioidlike G protein-coupled receptor. *Science*. 1995;270(5237):792-4.
6. Cox BM, Christie MJ, Devi L, Toll L, Traynor JR. Challenges for opioid receptor nomenclature: IUPHAR Review 9. *British journal of pharmacology*. 2015;172(2):317-23.
7. Schroder W, Lambert DG, Ko MC, Koch T. Functional plasticity of the N/OFQ-NOP receptor system determines analgesic properties of NOP receptor agonists. *British journal of pharmacology*. 2014;171(16):3777-800.
8. Thompson AA, Liu W, Chun E, Katritch V, Wu H, Vardy E, et al. Structure of the nociceptin/orphanin FQ receptor in complex with a peptide mimetic. *Nature*. 2012;485(7398):395-9.
9. Hawes BE, Graziano MP, Lambert DG. Cellular actions of nociceptin: transduction mechanisms. *Peptides*. 2000;21(7):961-7.
10. Knoflach F, Reinscheid RK, Civelli O, Kemp JA. Modulation of voltage-gated calcium channels by orphanin FQ in freshly dissociated hippocampal neurons. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 1996;16(21):6657-64.
11. New DC, Wong YH. The ORL1 receptor: molecular pharmacology and signalling mechanisms. *Neuro-Signals*. 2002;11(4):197-212.
12. Vaughan CW, Christie MJ. Increase by the ORL1 receptor (opioid receptor-like1) ligand, nociceptin, of inwardly rectifying K conductance in dorsal raphe nucleus neurones. *British journal of pharmacology*. 1996;117(8):1609-11.
13. Nicol B, Lambert DG, Rowbotham DJ, Smart D, McKnight AT. Nociceptin induced inhibition of K<sup>+</sup> evoked glutamate release from rat cerebrocortical slices. *British journal of pharmacology*. 1996;119(6):1081-3.
14. Nicol B, Lambert DG, Rowbotham DJ, Okuda-Ashitaka E, Ito S, Smart D, et al. Nocistatin reverses nociceptin inhibition of glutamate release from rat brain slices. *European journal of pharmacology*. 1998;356(2-3):R1-3.
15. Nicol B, Rowbotham DJ, Lambert DG. Nociceptin/orphanin FQ inhibits glutamate release from rat cerebellar and brain stem slices. *Neuroscience letters*. 2002;326(2):85-8.
16. Schlicker E, Morari M. Nociceptin/orphanin FQ and neurotransmitter release in the central nervous system. *Peptides*. 2000;21(7):1023-9.
17. Altier C, Khosravani H, Evans RM, Hameed S, Peloquin JB, Vartian BA, et al. ORL1 receptor-mediated internalization of N-type calcium channels. *Nature neuroscience*. 2006;9(1):31-40.

18. Evans RM, You H, Hameed S, Altier C, Mezghrani A, Bourinet E, et al. Heterodimerization of ORL1 and opioid receptors and its consequences for N-type calcium channel regulation. *The Journal of biological chemistry*. 2010;285(2):1032-40.
19. Murali SS, Napier IA, Rycroft BK, Christie MJ. Opioid-related (ORL1) receptors are enriched in a subpopulation of sensory neurons and prolonged activation produces no functional loss of surface N-type calcium channels. *The Journal of physiology*. 2012;590(Pt 7):1655-67.
20. Lou LG, Ma L, Pei G. Nociceptin/orphanin FQ activates protein kinase C, and this effect is mediated through phospholipase C/Ca<sup>2+</sup> pathway. *Biochemical and biophysical research communications*. 1997;240(2):304-8.
21. Fukuda K, Shoda T, Morikawa H, Kato S, Mima H, Mori K. Activation of phospholipase A2 by the nociceptin receptor expressed in Chinese hamster ovary cells. *Journal of neurochemistry*. 1998;71(5):2186-92.
22. Armstead WM. Differential activation of ERK, p38, and JNK MAPK by nociceptin/orphanin FQ in the potentiation of prostaglandin cerebrovasoconstriction after brain injury. *European journal of pharmacology*. 2006;529(1-3):129-35.
23. Chan AS, Wong YH. Regulation of c-Jun N-terminal kinase by the ORL(1) receptor through multiple G proteins. *The Journal of pharmacology and experimental therapeutics*. 2000;295(3):1094-100.
24. Donica CL, Ramirez VI, Awwad HO, Zaveri NT, Toll L, Standifer KM. Orphanin FQ/nociceptin activates nuclear factor kappa B. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology*. 2011;6(4):617-25.
25. Fukuda K, Shoda T, Morikawa H, Kato S, Mori K. Activation of mitogen-activated protein kinase by the nociceptin receptor expressed in Chinese hamster ovary cells. *FEBS letters*. 1997;412(2):290-4.
26. Zhang J, Salojin KV, Gao JX, Cameron MJ, Bergerot I, Delovitch TL. p38 mitogen-activated protein kinase mediates signal integration of TCR/CD28 costimulation in primary murine T cells. *Journal of immunology*. 1999;162(7):3819-29.
27. Wu EH, Lo RK, Wong YH. Regulation of STAT3 activity by G16-coupled receptors. *Biochemical and biophysical research communications*. 2003;303(3):920-5.
28. Guerrini R, Calo G, Rizzi A, Bigoni R, Bianchi C, Salvadori S, et al. A new selective antagonist of the nociceptin receptor. *British journal of pharmacology*. 1998;123(2):163-5.
29. Candeletti S, Guerrini R, Calo G, Romualdi P, Ferri S. Supraspinal and spinal effects of [Phe<sup>1</sup>psi(CH<sub>2</sub>-NH)<sup>2</sup>]-nociceptin(1-13)-NH<sub>2</sub> on nociception in the rat. *Life sciences*. 2000;66(3):257-64.
30. Okada K, Sujaku T, Chuman Y, Nakashima R, Nose T, Costa T, et al. Highly potent nociceptin analog containing the Arg-Lys triple repeat. *Biochemical and biophysical research communications*. 2000;278(2):493-8.
31. Rizzi D, Rizzi A, Bigoni R, Camarda V, Marzola G, Guerrini R, et al. [Arg(14),Lys(15)]nociceptin, a highly potent agonist of the nociceptin/orphanin FQ receptor: in vitro and in vivo studies. *The Journal of pharmacology and experimental therapeutics*. 2002;300(1):57-63.
32. Calo G, Rizzi A, Rizzi D, Bigoni R, Guerrini R, Marzola G, et al. [Nphe<sup>1</sup>,Arg<sup>14</sup>,Lys<sup>15</sup>]nociceptin-NH<sub>2</sub>, a novel potent and selective antagonist of the nociceptin/orphanin FQ receptor. *British journal of pharmacology*. 2002;136(2):303-11.
33. Calo G, Guerrini R, Rizzi A, Salvadori S, Burmeister M, Kapusta DR, et al. UFP-101, a peptide antagonist selective for the nociceptin/orphanin FQ receptor. *CNS drug reviews*. 2005;11(2):97-112.

34. Brookes ZL, Stedman EN, Brown NJ, Hebbes CP, Guerrini R, Calo G, et al. The nociceptin/orphanin FQ receptor antagonist UFP-101 reduces microvascular inflammation to lipopolysaccharide in vivo. *PloS one.* 2013;8(9):e74943.
35. Han Y, Guo Z, Wang LL, Zhang LZ, Yao TP. Antagonism of endogenous nociceptin/orphanin FQ inhibits infarction-associated ventricular arrhythmias via PKC-dependent mechanism in rats. *British journal of pharmacology.* 2013;170(3):614-23.
36. Ozaki S, Kawamoto H, Itoh Y, Miyaji M, Azuma T, Ichikawa D, et al. In vitro and in vivo pharmacological characterization of J-113397, a potent and selective non-peptidyl ORL1 receptor antagonist. *European journal of pharmacology.* 2000;402(1-2):45-53.
37. Zaratin PF, Petrone G, Sbacchi M, Garnier M, Fossati C, Petrillo P, et al. Modification of nociception and morphine tolerance by the selective opiate receptor-like orphan receptor antagonist (-)-cis-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl]methyl]-6,7,8,9-tetrahydro-5H-benzocyclohepten-5-ol (SB-612111). *The Journal of pharmacology and experimental therapeutics.* 2004;308(2):454-61.
38. Goto Y, Arai-Otsuki S, Tachibana Y, Ichikawa D, Ozaki S, Takahashi H, et al. Identification of a novel spiropiperidine opioid receptor-like 1 antagonist class by a focused library approach featuring 3D-pharmacophore similarity. *Journal of medicinal chemistry.* 2006;49(3):847-9.
39. Fischetti C, Camarda V, Rizzi A, Pela M, Trapella C, Guerrini R, et al. Pharmacological characterization of the nociceptin/orphanin FQ receptor non peptide antagonist Compound 24. *European journal of pharmacology.* 2009;614(1-3):50-7.
40. Mahmoud S, Margas W, Trapella C, Calo G, Ruiz-Velasco V. Modulation of silent and constitutively active nociceptin/orphanin FQ receptors by potent receptor antagonists and Na<sup>+</sup> ions in rat sympathetic neurons. *Molecular pharmacology.* 2010;77(5):804-17.
41. Volta M, Viaro R, Trapella C, Marti M, Morari M. Dopamine-nociceptin/orphanin FQ interactions in the substantia nigra reticulata of hemiparkinsonian rats: involvement of D2/D3 receptors and impact on nigro-thalamic neurons and motor activity. *Experimental neurology.* 2011;228(1):126-37.
42. Ces A, Reiss D, Walter O, Wichmann J, Prinsen EP, Kieffer BL, et al. Activation of nociceptin/orphanin FQ peptide receptors disrupts visual but not auditory sensorimotor gating in BALB/cByJ mice: comparison to dopamine receptor agonists. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology.* 2012;37(2):378-89.
43. Journigan VB, Polgar WE, Khroyan TV, Zaveri NT. Designing bifunctional NOP receptor-mu opioid receptor ligands from NOP-receptor selective scaffolds. Part II. *Bioorganic & medicinal chemistry.* 2014;22(8):2508-16.
44. Zaveri NT, Jiang F, Olsen C, Polgar WE, Toll L. Designing bifunctional NOP receptor-mu opioid receptor ligands from NOP receptor-selective scaffolds. Part I. *Bioorganic & medicinal chemistry letters.* 2013;23(11):3308-13.
45. Johnson RE, Fudala PJ, Payne R. Buprenorphine: considerations for pain management. *Journal of pain and symptom management.* 2005;29(3):297-326.
46. Pergolizzi J, Aloisi AM, Dahan A, Filitz J, Langford R, Likar R, et al. Current knowledge of buprenorphine and its unique pharmacological profile. *Pain practice : the official journal of World Institute of Pain.* 2010;10(5):428-50.
47. Sobczak M, Cami-Kobeci G, Salaga M, Husbands SM, Fichna J. Novel mixed NOP/MOP agonist BU08070 alleviates pain and inhibits gastrointestinal motility in mouse models mimicking diarrhea-predominant irritable bowel syndrome symptoms. *European journal of pharmacology.* 2014;736:63-9.

48. Gavioli EC, de Medeiros IU, Monteiro MC, Calo G, Romao PR. Nociceptin/orphanin FQ-NOP receptor system in inflammatory and immune-mediated diseases. Vitamins and hormones. 2015;97:241-66.
49. Mollereau C, Mouledous L. Tissue distribution of the opioid receptor-like (ORL1) receptor. Peptides. 2000;21(7):907-17.
50. Li L, Dong L, Wang S. Expression of the nociceptin/orphanin FQ receptor in the intestinal mucosa of IBS patients. Experimental and therapeutic medicine. 2013;6(3):679-83.
51. Agostini S, Eutamene H, Broccardo M, Impronta G, Petrella C, Theodorou V, et al. Peripheral anti-nociceptive effect of nociceptin/orphanin FQ in inflammation and stress-induced colonic hyperalgesia in rats. Pain. 2009;141(3):292-9.
52. McDonald J, Leonard AD, Serrano-Gomez A, Young SP, Swanevelder J, Thompson JP, et al. Assessment of nociceptin/orphanin FQ and micro-opioid receptor mRNA in the human right atrium. British journal of anaesthesia. 2010;104(6):698-704.
53. Kummer W, Fischer A. Nociceptin and its receptor in guinea-pig sympathetic ganglia. Neuroscience letters. 1997;234(1):35-8.
54. Xie GX, Meuser T, Pietruck C, Sharma M, Palmer PP. Presence of opioid receptor-like (ORL1) receptor mRNA splice variants in peripheral sensory and sympathetic neuronal ganglia. Life sciences. 1999;64(22):2029-37.
55. Toll L, Bruchas MR, Calo G, Cox BM, Zaveri NT. Nociceptin/Orphanin FQ Receptor Structure, Signaling, Ligands, Functions, and Interactions with Opioid Systems. Pharmacological reviews. 2016;68(2):419-57.
56. Cavanaugh DJ, Lee H, Lo L, Shields SD, Zylka MJ, Basbaum AI, et al. Distinct subsets of unmyelinated primary sensory fibers mediate behavioral responses to noxious thermal and mechanical stimuli. Proceedings of the National Academy of Sciences of the United States of America. 2009;106(22):9075-80.
57. Basbaum AI, Bautista DM, Scherrer G, Julius D. Cellular and molecular mechanisms of pain. Cell. 2009;139(2):267-84.
58. Scherrer G, Imamachi N, Cao YQ, Contet C, Mennicken F, O'Donnell D, et al. Dissociation of the opioid receptor mechanisms that control mechanical and heat pain. Cell. 2009;137(6):1148-59.
59. Vrontou S, Wong AM, Rau KK, Koerber HR, Anderson DJ. Genetic identification of C fibres that detect massage-like stroking of hairy skin in vivo. Nature. 2013;493(7434):669-73.
60. Bardoni R, Tawfik VL, Wang D, Francois A, Solorzano C, Shuster SA, et al. Delta Opioid Receptors Presynaptically Regulate Cutaneous Mechanosensory Neuron Input to the Spinal Cord Dorsal Horn. Neuron. 2014;81(6):1443.
61. Ozawa A, Brunori G, Mercatelli D, Wu J, Cippitelli A, Zou B, et al. Knock-In Mice with NOP-eGFP Receptors Identify Receptor Cellular and Regional Localization. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2015;35(33):11682-93.
62. Palmisano M, Mercatelli D, Caputi FF, Carretta D, Romualdi P, Candeletti S. N/OFQ system in brain areas of nerve-injured mice: its role in different aspects of neuropathic pain. Genes, brain, and behavior. 2017;16(5):537-45.
63. Ozawa A, Brunori G, Cippitelli A, Toll N, Schoch J, Kieffer BL, et al. Analysis of the distribution of spinal NOP receptors in a chronic pain model using NOP-eGFP knock-in mice. British journal of pharmacology. 2018;175(13):2662-75.
64. Ma F, Xie H, Dong ZQ, Wang YQ, Wu GC. Expression of ORL1 mRNA in some brain nuclei in neuropathic pain rats. Brain research. 2005;1043(1-2):214-7.
65. Popolek-Barczyk K, Rojewska E, Jurga AM, Makuch W, Zador F, Borsodi A, et al. Minocycline enhances the effectiveness of nociceptin/orphanin FQ during neuropathic pain. BioMed research international. 2014;2014:762930.

66. Zhang Y, Simpson-Durand CD, Standifer KM. Nociceptin/orphanin FQ peptide receptor antagonist JTC-801 reverses pain and anxiety symptoms in a rat model of post-traumatic stress disorder. *British journal of pharmacology*. 2015;172(2):571-82.
67. Pan B, Schroder W, Jostock R, Schwartz M, Rosson G, Polydefkis M. Nociceptin/orphanin FQ opioid peptide-receptor expression in pachyonychia congenita. *Journal of the peripheral nervous system : JPNS*. 2018;23(4):241-8.
68. Fu X, Wang YQ, Wang J, Yu J, Wu GC. Changes in expression of nociceptin/orphanin FQ and its receptor in spinal dorsal horn during electroacupuncture treatment for peripheral inflammatory pain in rats. *Peptides*. 2007;28(6):1220-8.
69. Anand P, Yianguo Y, Anand U, Mukerji G, Sinisi M, Fox M, et al. Nociceptin/orphanin FQ receptor expression in clinical pain disorders and functional effects in cultured neurons. *Pain*. 2016;157(9):1960-9.
70. Stamer UM, Book M, Comos C, Zhang L, Nauck F, Stuber F. Expression of the nociceptin precursor and nociceptin receptor is modulated in cancer and septic patients. *British journal of anaesthesia*. 2011;106(4):566-72.
71. Sobczak M, Mokrowiecka A, Cygankiewicz AI, Zakrzewski PK, Salaga M, Storr M, et al. Anti-inflammatory and antinociceptive action of an orally available nociceptin receptor agonist SCH 221510 in a mouse model of inflammatory bowel diseases. *The Journal of pharmacology and experimental therapeutics*. 2014;348(3):401-9.
72. Brookes ZL, Stedman EN, Guerrini R, Lawton BK, Calo G, Lambert DG. Proinflammatory and vasodilator effects of nociceptin/orphanin FQ in the rat mesenteric microcirculation are mediated by histamine. *American journal of physiology Heart and circulatory physiology*. 2007;293(5):H2977-85.
73. Kiguchi N, Ding H, Ko MC. Central N/OFQ-NOP Receptor System in Pain Modulation. *Advances in pharmacology*. 2016;75:217-43.
74. Fulford AJ. Endogenous nociceptin system involvement in stress responses and anxiety behavior. *Vitamins and hormones*. 2015;97:267-93.
75. Chiou LC, Liao YY, Fan PC, Kuo PH, Wang CH, Riemer C, et al. Nociceptin/orphanin FQ peptide receptors: pharmacology and clinical implications. *Current drug targets*. 2007;8(1):117-35.
76. Reinscheid RK. The Orphanin FQ / Nociceptin receptor as a novel drug target in psychiatric disorders. *CNS & neurological disorders drug targets*. 2006;5(2):219-24.
77. Witkin JM, Statnick MA, Rorick-Kehn LM, Pintar JE, Ansonoff M, Chen Y, et al. The biology of Nociceptin/Orphanin FQ (N/OFQ) related to obesity, stress, anxiety, mood, and drug dependence. *Pharmacology & therapeutics*. 2014;141(3):283-99.
78. Zaveri NT. Nociceptin Opioid Receptor (NOP) as a Therapeutic Target: Progress in Translation from Preclinical Research to Clinical Utility. *Journal of medicinal chemistry*. 2016;59(15):7011-28.
79. Zaveri N. Peptide and nonpeptide ligands for the nociceptin/orphanin FQ receptor ORL1: research tools and potential therapeutic agents. *Life sciences*. 2003;73(6):663-78.
80. Mogil JS, Pasternak GW. The molecular and behavioral pharmacology of the orphanin FQ/nociceptin peptide and receptor family. *Pharmacological reviews*. 2001;53(3):381-415.
81. Gavioli EC, Holanda VAD, Ruzza C. NOP Ligands for the Treatment of Anxiety and Mood Disorders. *Handbook of experimental pharmacology*. 2018.
82. Hiramatsu M, Miwa M, Hashimoto K, Kawai S, Nomura N. Nociceptin/orphanin FQ reverses mecamylamine-induced learning and memory impairment as well as decrease in hippocampal acetylcholine release in the rat. *Brain research*. 2008;1195:96-103.
83. Reiss D, Prinsen EP, Wichmann J, Kieffer BL, Ouagazzal AM. The nociceptin orphanin FQ peptide receptor agonist, Ro64-6198, impairs recognition memory formation

- through interaction with glutamatergic but not cholinergic receptor antagonists. *Neurobiology of learning and memory.* 2012;98(3):254-60.
84. Halford WP, Gebhardt BM, Carr DJ. Functional role and sequence analysis of a lymphocyte orphan opioid receptor. *Journal of neuroimmunology.* 1995;59(1-2):91-101.
  85. Peluso J, LaForge KS, Matthes HW, Kreek MJ, Kieffer BL, Gaveriaux-Ruff C. Distribution of nociceptin/orphanin FQ receptor transcript in human central nervous system and immune cells. *Journal of neuroimmunology.* 1998;81(1-2):184-92.
  86. Wick MJ, Minnerath SR, Roy S, Ramakrishnan S, Loh HH. Expression of alternate forms of brain opioid 'orphan' receptor mRNA in activated human peripheral blood lymphocytes and lymphocytic cell lines. *Brain research Molecular brain research.* 1995;32(2):342-7.
  87. Pampusch MS, Serie JR, Osinski MA, Seybold VS, Murtaugh MP, Brown DR. Expression of nociceptin/OFQ receptor and prepro-nociceptin/OFQ in lymphoid tissues. *Peptides.* 2000;21(12):1865-70.
  88. Andoh T, Yageta Y, Takeshima H, Kuraishi Y. Intradermal nociceptin elicits itch-associated responses through leukotriene B(4) in mice. *The Journal of investigative dermatology.* 2004;123(1):196-201.
  89. Sobczak M, Salaga M, Storr M, Fichna J. Nociceptin / orphanin FQ (NOP) receptors as novel potential target in the treatment of gastrointestinal diseases. *Current drug targets.* 2013;14(10):1203-9.
  90. Serhan CN, Fierro IM, Chiang N, Pouliot M. Cutting edge: nociceptin stimulates neutrophil chemotaxis and recruitment: inhibition by aspirin-triggered-15-epi-lipoxin A4. *Journal of immunology.* 2001;166(6):3650-4.
  91. Trombella S, Vergura R, Falzarano S, Guerrini R, Calo G, Spisani S. Nociceptin/orphanin FQ stimulates human monocyte chemotaxis via NOP receptor activation. *Peptides.* 2005;26(8):1497-502.
  92. Kimura T, Kitaichi K, Hiramatsu K, Yoshida M, Ito Y, Kume H, et al. Intradermal application of nociceptin increases vascular permeability in rats: the possible involvement of histamine release from mast cells. *European journal of pharmacology.* 2000;407(3):327-32.
  93. Carvalho D, Petronilho F, Vuolo F, Machado RA, Constantino L, Guerrini R, et al. The nociceptin/orphanin FQ-NOP receptor antagonist effects on an animal model of sepsis. *Intensive care medicine.* 2008;34(12):2284-90.
  94. Alt C, Lam JS, Harrison MT, Kershaw KM, Samuelsson S, Toll L, et al. Nociceptin/orphanin FQ inhibition with SB612111 ameliorates dextran sodium sulfate-induced colitis. *European journal of pharmacology.* 2012;683(1-3):285-93.
  95. Kato S, Tsuzuki Y, Hokari R, Okada Y, Miyazaki J, Matsuzaki K, et al. Role of nociceptin/orphanin FQ (Noc/oFQ) in murine experimental colitis. *Journal of neuroimmunology.* 2005;161(1-2):21-8.
  96. Varty GB, Lu SX, Morgan CA, Cohen-Williams ME, Hodgson RA, Smith-Torhan A, et al. The anxiolytic-like effects of the novel, orally active nociceptin opioid receptor agonist 8-[bis(2-methylphenyl)methyl]-3-phenyl-8-azabicyclo[3.2.1]octan-3-ol (SCH 221510). *The Journal of pharmacology and experimental therapeutics.* 2008;326(2):672-82.
  97. Basbaum AI, Jessell T. *The perception of pain.* New York2000.
  98. Littlejohn G. Neuroinflammation in fibromyalgia and CRPS: top-down or bottomup? *Nature reviews Rheumatology.* 2016;12(4):242.
  99. Sakurada T, Komatsu T, Moriyama T, Sasaki M, Sanai K, Orito T, et al. Effects of intraplantar injections of nociceptin and its N-terminal fragments on nociceptive and desensitized responses induced by capsaicin in mice. *Peptides.* 2005;26(12):2505-12.
  100. Zaratin PF, Petrone G, Sbacchi M, Garnier M, Fossati C, Petrillo P, et al. Modification of nociception and morphine tolerance by the selective opiate receptor-like orphan receptor

- antagonist (-)-cis-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl]methyl]-6,7,8,9-tetrahydro-5H-benzocyclohepten-5-ol (SB-612111). *The Journal of pharmacology and experimental therapeutics.* 2004;308(2):454-61.
101. Wang YQ, Zhu CB, Cao XD, Wu GC. Supraspinal hyperalgesia and spinal analgesia by [Phe<sup>1</sup>psi(CH<sub>2</sub>-NH)Gly<sup>2</sup>]nociceptin-(1-13)-NH<sub>2</sub> in rat. *European journal of pharmacology.* 1999;376(3):R1-3.
  102. Tian JH, Xu W, Fang Y, Mogil JS, Grisel JE, Grandy DK, et al. Bidirectional modulatory effect of orphanin FQ on morphine-induced analgesia: antagonism in brain and potentiation in spinal cord of the rat. *British journal of pharmacology.* 1997;120(4):676-80.
  103. Tsai CY, Poon YY, Huang YH, Chan SH. Activation of spinal nociceptin receptors induces cardiovascular depression and antinociception in an independent manner in mice. *Journal of pain research.* 2018;11:2699-708.
  104. Scoto GM, Arico G, Ronsisvalle S, Parenti C. Blockade of the nociceptin/orphanin FQ/NOP receptor system in the rat ventrolateral periaqueductal gray potentiates DAMGO analgesia. *Peptides.* 2007;28(7):1441-6.
  105. Inoue M, Shimohira I, Yoshida A, Zimmer A, Takeshima H, Sakurada T, et al. Dose-related opposite modulation by nociceptin/orphanin FQ of substance P nociception in the nociceptors and spinal cord. *The Journal of pharmacology and experimental therapeutics.* 1999;291(1):308-13.
  106. Sakurada T, Katsuyama S, Sakurada S, Inoue M, Tan-No K, Kisara K, et al. Nociceptin-induced scratching, biting and licking in mice: involvement of spinal NK1 receptors. *British journal of pharmacology.* 1999;127(7):1712-8.
  107. Rizzi A, Gavioli EC, Marzola G, Spagnolo B, Zucchini S, Ciccocioppo R, et al. Pharmacological characterization of the nociceptin/orphanin FQ receptor antagonist SB-612111 [(-)-cis-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl]methyl]-6,7,8,9-tetrahydro-5H-benzocyclohepten-5-ol]: in vivo studies. *The Journal of pharmacology and experimental therapeutics.* 2007;321(3):968-74.
  108. Khroyan TV, Polgar WE, Jiang F, Zaveri NT, Toll L. Nociceptin/orphanin FQ receptor activation attenuates antinociception induced by mixed nociceptin/orphanin FQ/mu-opioid receptor agonists. *The Journal of pharmacology and experimental therapeutics.* 2009;331(3):946-53.
  109. Rizzi A, Cerlesi MC, Ruzza C, Malfacini D, Ferrari F, Bianco S, et al. Pharmacological characterization of cebranopadol a novel analgesic acting as mixed nociceptin/orphanin FQ and opioid receptor agonist. *Pharmacology research & perspectives.* 2016;4(4):e00247.
  110. Rizzi A, Nazzaro C, Marzola GG, Zucchini S, Trapella C, Guerrini R, et al. Endogenous nociceptin/orphanin FQ signalling produces opposite spinal antinociceptive and supraspinal pronociceptive effects in the mouse formalin test: pharmacological and genetic evidences. *Pain.* 2006;124(1-2):100-8.
  111. Khroyan TV, Polgar WE, Orduna J, Montenegro J, Jiang F, Zaveri NT, et al. Differential effects of nociceptin/orphanin FQ (NOP) receptor agonists in acute versus chronic pain: studies with bifunctional NOP/mu receptor agonists in the sciatic nerve ligation chronic pain model in mice. *The Journal of pharmacology and experimental therapeutics.* 2011;339(2):687-93.
  112. Rizzi A, Sukhtankar DD, Ding H, Hayashida K, Ruzza C, Guerrini R, et al. Spinal antinociceptive effects of the novel NOP receptor agonist PWT2-nociceptin/orphanin FQ in mice and monkeys. *British journal of pharmacology.* 2015;172(14):3661-70.
  113. Corradini L, Briscini L, Ongini E, Bertorelli R. The putative OP(4) antagonist, [Nphe<sup>1</sup>]nociceptin(1-13)NH<sup>2</sup>, prevents the effects of nociceptin in neuropathic rats. *Brain research.* 2001;905(1-2):127-33.

114. Scoto GM, Arico G, Iemolo A, Ronsisvalle S, Parenti C. Involvement of the Nociceptin/Orphanin FQ-NOP receptor system in the ventrolateral periaqueductal gray following mechanical allodynia in chronic pain. *Life sciences.* 2009;85(5-6):206-10.
115. Shan D, He Y, Long H, Zhou Y, Liu H, Xu R, et al. The effects of blocking N/OFQ receptors on orofacial pain following experimental tooth movement in rats. *Australian orthodontic journal.* 2016;32(2):206-10.
116. Vang D, Paul JA, Nguyen J, Tran H, Vincent L, Yasuda D, et al. Small-molecule nociceptin receptor agonist ameliorates mast cell activation and pain in sickle mice. *Haematologica.* 2015;100(12):1517-25.
117. Ding H, Hayashida K, Suto T, Sukhtankar DD, Kimura M, Mendenhall V, et al. Supraspinal actions of nociceptin/orphanin FQ, morphine and substance P in regulating pain and itch in non-human primates. *British journal of pharmacology.* 2015;172(13):3302-12.
118. Ko MC, Woods JH, Fantegrossi WE, Galuska CM, Wichmann J, Prinsen EP. Behavioral effects of a synthetic agonist selective for nociceptin/orphanin FQ peptide receptors in monkeys. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology.* 2009;34(9):2088-96.
119. Cremeans CM, Gruley E, Kyle DJ, Ko MC. Roles of mu-opioid receptors and nociceptin/orphanin FQ peptide receptors in buprenorphine-induced physiological responses in primates. *The Journal of pharmacology and experimental therapeutics.* 2012;343(1):72-81.
120. Lin AP, Ko MC. The therapeutic potential of nociceptin/orphanin FQ receptor agonists as analgesics without abuse liability. *ACS chemical neuroscience.* 2013;4(2):214-24.
121. Sukhtankar DD, Lee H, Rice KC, Ko MC. Differential effects of opioid-related ligands and NSAIDs in nonhuman primate models of acute and inflammatory pain. *Psychopharmacology.* 2014;231(7):1377-87.
122. Devine DP, Watson SJ, Akil H. Nociceptin/orphanin FQ regulates neuroendocrine function of the limbic-hypothalamic-pituitary-adrenal axis. *Neuroscience.* 2001;102(3):541-53.
123. Nicholson JR, Akil H, Watson SJ. Orphanin FQ-induced hyperphagia is mediated by corticosterone and central glucocorticoid receptors. *Neuroscience.* 2002;115(2):637-43.
124. Delaney G, Dawe KL, Hogan R, Hunjan T, Roper J, Hazell G, et al. Role of nociceptin/orphanin FQ and NOP receptors in the response to acute and repeated restraint stress in rats. *Journal of neuroendocrinology.* 2012;24(12):1527-41.
125. Leggett JD, Harbuz MS, Jessop DS, Fulford AJ. The nociceptin receptor antagonist [Nphe1,Arg14,Lys15]nociceptin/orphanin FQ-NH<sub>2</sub> blocks the stimulatory effects of nociceptin/orphanin FQ on the HPA axis in rats. *Neuroscience.* 2006;141(4):2051-7.
126. Gavioli EC, Calo G. Nociceptin/orphanin FQ receptor antagonists as innovative antidepressant drugs. *Pharmacology & therapeutics.* 2013;140(1):10-25.
127. Okawa H, Kudo M, Kudo T, Guerrini R, Lambert DG, Kushikata T, et al. Effects of nociceptinNH<sub>2</sub> and [Nphe1]nociceptin(1-13)NH<sub>2</sub> on rat brain noradrenaline release in vivo and in vitro. *Neuroscience letters.* 2001;303(3):173-6.
128. Rizzi A, Molinari S, Marti M, Marzola G, Calo G. Nociceptin/orphanin FQ receptor knockout rats: in vitro and in vivo studies. *Neuropharmacology.* 2011;60(4):572-9.
129. Gavioli EC, Marzola G, Guerrini R, Bertorelli R, Zucchini S, De Lima TC, et al. Blockade of nociceptin/orphanin FQ-NOP receptor signalling produces antidepressant-like effects: pharmacological and genetic evidences from the mouse forced swimming test. *The European journal of neuroscience.* 2003;17(9):1987-90.
130. Gavioli EC, Vaughan CW, Marzola G, Guerrini R, Mitchell VA, Zucchini S, et al. Antidepressant-like effects of the nociceptin/orphanin FQ receptor antagonist UFP-101: new evidence from rats and mice. *Naunyn-Schmiedeberg's archives of pharmacology.* 2004;369(6):547-53.

131. Gavioli EC, Calo G. Antidepressant- and anxiolytic-like effects of nociceptin/orphanin FQ receptor ligands. *Naunyn-Schmiedeberg's archives of pharmacology*. 2006;372(5):319-30.
132. Witkin JM, Rorick-Kehn LM, Benvenga MJ, Adams BL, Gleason SD, Knitowski KM, et al. Preclinical findings predicting efficacy and side-effect profile of LY2940094, an antagonist of nociceptin receptors. *Pharmacology research & perspectives*. 2016;4(6):e00275.
133. Goeldner C, Reiss D, Kieffer BL, Ouagazzal AM. Endogenous nociceptin/orphanin-FQ in the dorsal hippocampus facilitates despair-related behavior. *Hippocampus*. 2010;20(8):911-6.
134. Vitale G, Ruggieri V, Filaferro M, Frigeri C, Alboni S, Tascedda F, et al. Chronic treatment with the selective NOP receptor antagonist [Nphe 1, Arg 14, Lys 15]N/OFQ-NH 2 (UFP-101) reverses the behavioural and biochemical effects of unpredictable chronic mild stress in rats. *Psychopharmacology*. 2009;207(2):173-89.
135. Vitale G, Filaferro M, Micioni Di Bonaventura MV, Ruggieri V, Cifani C, Guerrini R, et al. Effects of [Nphe(1), Arg(14), Lys(15)] N/OFQ-NH2 (UFP-101), a potent NOP receptor antagonist, on molecular, cellular and behavioural alterations associated with chronic mild stress. *Journal of psychopharmacology*. 2017;31(6):691-703.
136. Holanda VA, Medeiros IU, Asth L, Guerrini R, Calo G, Gavioli EC. Antidepressant activity of nociceptin/orphanin FQ receptor antagonists in the mouse learned helplessness. *Psychopharmacology*. 2016;233(13):2525-32.
137. Medeiros IU, Ruzza C, Asth L, Guerrini R, Romao PR, Gavioli EC, et al. Blockade of nociceptin/orphanin FQ receptor signaling reverses LPS-induced depressive-like behavior in mice. *Peptides*. 2015;72:95-103.
138. Redrobe JP, Calo G, Regoli D, Quirion R. Nociceptin receptor antagonists display antidepressant-like properties in the mouse forced swimming test. *Naunyn-Schmiedeberg's archives of pharmacology*. 2002;365(2):164-7.
139. Post A, Smart TS, Krikke-Workel J, Dawson GR, Harmer CJ, Browning M, et al. A Selective Nociceptin Receptor Antagonist to Treat Depression: Evidence from Preclinical and Clinical Studies. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2016;41(7):1803-12.
140. Asth L, Ruzza C, Malfacini D, Medeiros I, Guerrini R, Zaveri NT, et al. Beta-arrestin 2 rather than G protein efficacy determines the anxiolytic-versus antidepressant-like effects of nociceptin/orphanin FQ receptor ligands. *Neuropharmacology*. 2016;105:434-42.
141. Holanda VAD, Santos WB, Asth L, Guerrini R, Calo G, Ruzza C, et al. NOP agonists prevent the antidepressant-like effects of nortriptyline and fluoxetine but not R-ketamine. *Psychopharmacology*. 2018;235(11):3093-102.
142. Fernandez F, Misilmeri MA, Felger JC, Devine DP. Nociceptin/orphanin FQ increases anxiety-related behavior and circulating levels of corticosterone during neophobic tests of anxiety. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2004;29(1):59-71.
143. Green MK, Barbieri EV, Brown BD, Chen KW, Devine DP. Roles of the bed nucleus of stria terminalis and of the amygdala in N/OFQ-mediated anxiety and HPA axis activation. *Neuropeptides*. 2007;41(6):399-410.
144. Duzzioni M, Duarte FS, Leme LR, Gavioli EC, De Lima TC. Anxiolytic-like effect of central administration of NOP receptor antagonist UFP-101 in rats submitted to the elevated T-maze. *Behavioural brain research*. 2011;222(1):206-11.
145. Aujla H, Cannarsa R, Romualdi P, Cicocioppo R, Martin-Fardon R, Weiss F. Modification of anxiety-like behaviors by nociceptin/orphanin FQ (N/OFQ) and time-dependent changes in N/OFQ-NOP gene expression following ethanol withdrawal. *Addiction biology*. 2013;18(3):467-79.

146. Kamei J, Matsunawa Y, Miyata S, Tanaka S, Saitoh A. Effects of nociceptin on the exploratory behavior of mice in the hole-board test. European journal of pharmacology. 2004;489(1-2):77-87.
147. Asth L, Correia N, Lobao-Soares B, De Lima TC, Guerrini R, Calo G, et al. Nociceptin/orphanin FQ induces simultaneously anxiolytic and amnesic effects in the mouse elevated T-maze task. Naunyn-Schmiedeberg's archives of pharmacology. 2015;388(1):33-41.
148. Filaferro M, Ruggieri V, Novi C, Calo G, Cifani C, Micioni Di Bonaventura MV, et al. Functional antagonism between nociceptin/orphanin FQ and corticotropin-releasing factor in rat anxiety-related behaviors: involvement of the serotonergic system. Neuropeptides. 2014;48(4):189-97.
149. Gavioli EC, Rae GA, Calo G, Guerrini R, De Lima TC. Central injections of nocistatin or its C-terminal hexapeptide exert anxiogenic-like effect on behaviour of mice in the plus-maze test. British journal of pharmacology. 2002;136(5):764-72.
150. Griebel G, Perrault G, Sanger DJ. Orphanin FQ, a novel neuropeptide with anti-stress-like activity. Brain research. 1999;836(1-2):221-4.
151. Jenck F, Moreau JL, Martin JR, Kilpatrick GJ, Reinscheid RK, Monsma FJ, Jr., et al. Orphanin FQ acts as an anxiolytic to attenuate behavioral responses to stress. Proceedings of the National Academy of Sciences of the United States of America. 1997;94(26):14854-8.
152. Vitale G, Arletti R, Ruggieri V, Cifani C, Massi M. Anxiolytic-like effects of nociceptin/orphanin FQ in the elevated plus maze and in the conditioned defensive burying test in rats. Peptides. 2006;27(9):2193-200.
153. Dautzenberg FM, Wichmann J, Higelin J, Py-Lang G, Kratzeisen C, Malherbe P, et al. Pharmacological characterization of the novel nonpeptide orphanin FQ/nociceptin receptor agonist Ro 64-6198: rapid and reversible desensitization of the ORL1 receptor in vitro and lack of tolerance in vivo. The Journal of pharmacology and experimental therapeutics. 2001;298(2):812-9.
154. Jenck F, Wichmann J, Dautzenberg FM, Moreau JL, Ouagazzal AM, Martin JR, et al. A synthetic agonist at the orphanin FQ/nociceptin receptor ORL1: anxiolytic profile in the rat. Proceedings of the National Academy of Sciences of the United States of America. 2000;97(9):4938-43.
155. Ross TM, Battista K, Bignan GC, Brenneman DE, Connolly PJ, Liu J, et al. A selective small molecule NOP (ORL-1 receptor) partial agonist for the treatment of anxiety. Bioorganic & medicinal chemistry letters. 2015;25(3):602-6.
156. Aziz AM, Brothers S, Sartor G, Holm L, Heilig M, Wahlestedt C, et al. The nociceptin/orphanin FQ receptor agonist SR-8993 as a candidate therapeutic for alcohol use disorders: validation in rat models. Psychopharmacology. 2016;233(19-20):3553-63.
157. Wichmann J, Adam G, Rover S, Hennig M, Scalzone M, Cesura AM, et al. Synthesis of (1S,3aS)-8-(2,3,3a,4,5,6-hexahydro-1H-phenalen-1-yl)-1-phenyl-1,3,8-triaza-spiro[4.5]decan-4-one, a potent and selective orphanin FQ (OFQ) receptor agonist with anxiolytic-like properties. European journal of medicinal chemistry. 2000;35(9):839-51.
158. Wichmann J, Adam G, Rover S, Cesura AM, Dautzenberg FM, Jenck F. 8-acenaphthen-1-yl-1-phenyl-1,3,8-triaza-spiro[4.5]decan-4-one derivatives as orphanin FQ receptor agonists. Bioorganic & medicinal chemistry letters. 1999;9(16):2343-8.
159. Post A, Smart TS, Jackson K, Mann J, Mohs R, Rorick-Kehn L, et al. Proof-of-Concept Study to Assess the Nociceptin Receptor Antagonist LY2940094 as a New Treatment for Alcohol Dependence. Alcoholism, clinical and experimental research. 2016;40(9):1935-44.
160. Christoph A, Eerdekkens MH, Kok M, Volkers G, Freyngagen R. Cebranopadol, a novel first-in-class analgesic drug candidate: first experience in patients with chronic low back pain in a randomized clinical trial. Pain. 2017;158(9):1813-24.

161. Kantola I, Scheinin M, Gulbrandsen T, Meland N, Smerud KT. Safety, Tolerability, and Antihypertensive Effect of SER100, an Opiate Receptor-Like 1 (ORL-1) Partial Agonist, in Patients With Isolated Systolic Hypertension. *Clinical pharmacology in drug development.* 2017;6(6):584-91.
162. Woodcock A, McLeod RL, Sadeh J, Smith JA. The efficacy of a NOP1 agonist (SCH486757) in subacute cough. *Lung.* 2010;188 Suppl 1:S47-52.
163. Staud R. Is it all central sensitization? Role of peripheral tissue nociception in chronic musculoskeletal pain. *Current rheumatology reports.* 2010;12(6):448-54.
164. Costigan M, Scholz J, Woolf CJ. Neuropathic pain: a maladaptive response of the nervous system to damage. *Annual review of neuroscience.* 2009;32:1-32.
165. Ceko M, Bushnell MC, Gracely RH. Neurobiology underlying fibromyalgia symptoms. *Pain research and treatment.* 2012;2012:585419.
166. Wolfe F, Clauw DJ, Fitzcharles MA, Goldenberg DL, Katz RS, Mease P, et al. The American College of Rheumatology preliminary diagnostic criteria for fibromyalgia and measurement of symptom severity. *Arthritis care & research.* 2010;62(5):600-10.
167. Wolfe F, Clauw DJ, Fitzcharles MA, Goldenberg DL, Hauser W, Katz RL, et al. 2016 Revisions to the 2010/2011 fibromyalgia diagnostic criteria. *Seminars in arthritis and rheumatism.* 2016;46(3):319-29.
168. Aydede M, Shriver A. Recently introduced definition of "nociplastic pain" by the International Association for the Study of Pain needs better formulation. *Pain.* 2018;159(6):1176-7.
169. Lawrence RC, Felson DT, Helmick CG, Arnold LM, Choi H, Deyo RA, et al. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part II. *Arthritis and rheumatism.* 2008;58(1):26-35.
170. Wolfe F, Smythe HA, Yunus MB, Bennett RM, Bombardier C, Goldenberg DL, et al. The American College of Rheumatology 1990 Criteria for the Classification of Fibromyalgia. Report of the Multicenter Criteria Committee. *Arthritis and rheumatism.* 1990;33(2):160-72.
171. Wolfe F, Clauw DJ, Fitzcharles MA, Goldenberg DL, Hauser W, Katz RS, et al. Fibromyalgia criteria and severity scales for clinical and epidemiological studies: a modification of the ACR Preliminary Diagnostic Criteria for Fibromyalgia. *The Journal of rheumatology.* 2011;38(6):1113-22.
172. Cabo-Meseguer A, Cerdá-Olmedo G, Trillo-Mata JL. Fibromyalgia: Prevalence, epidemiologic profiles and economic costs. *Medicina clinica.* 2017;149(10):441-8.
173. Galek A, Erbsloh-Moller B, Kollner V, Kuhn-Becker H, Langhorst J, Petermann F, et al. [Mental disorders in patients with fibromyalgia syndrome: screening in centres of different medical specialties]. *Schmerz.* 2013;27(3):296-304.
174. Hauser W, Jung E, Erbsloh-Moller B, Gesmann M, Kuhn-Becker H, Petermann F, et al. The German fibromyalgia consumer reports - a cross-sectional survey. *BMC musculoskeletal disorders.* 2012;13:74.
175. Bennett RM, Jones J, Turk DC, Russell IJ, Matallana L. An internet survey of 2,596 people with fibromyalgia. *BMC musculoskeletal disorders.* 2007;8:27.
176. Kim H, Kim J, Loggia ML, Cahalan C, Garcia RG, Vangel MG, et al. Fibromyalgia is characterized by altered frontal and cerebellar structural covariance brain networks. *NeuroImage Clinical.* 2015;7:667-77.
177. Jensen KB, Srinivasan P, Spaeth R, Tan Y, Kosek E, Petzke F, et al. Overlapping structural and functional brain changes in patients with long-term exposure to fibromyalgia pain. *Arthritis and rheumatism.* 2013;65(12):3293-303.
178. Usui C, Soma T, Hatta K, Aratani S, Fujita H, Nishioka K, et al. A study of brain metabolism in fibromyalgia by positron emission tomography. *Progress in neuropsychopharmacology & biological psychiatry.* 2017;75:120-7.

179. Pujol J, Macia D, Garcia-Fontanals A, Blanco-Hinojo L, Lopez-Sola M, Garcia-Blanco S, et al. The contribution of sensory system functional connectivity reduction to clinical pain in fibromyalgia. *Pain*. 2014;155(8):1492-503.
180. Pujol J, Lopez-Sola M, Ortiz H, Vilanova JC, Harrison BJ, Yucel M, et al. Mapping brain response to pain in fibromyalgia patients using temporal analysis of fMRI. *PloS one*. 2009;4(4):e5224.
181. Gracely RH, Petzke F, Wolf JM, Clauw DJ. Functional magnetic resonance imaging evidence of augmented pain processing in fibromyalgia. *Arthritis and rheumatism*. 2002;46(5):1333-43.
182. Cook DB, Lange G, Ciccone DS, Liu WC, Steffener J, Natelson BH. Functional imaging of pain in patients with primary fibromyalgia. *The Journal of rheumatology*. 2004;31(2):364-78.
183. Lopez-Sola M, Woo CW, Pujol J, Deus J, Harrison BJ, Monfort J, et al. Towards a neurophysiological signature for fibromyalgia. *Pain*. 2017;158(1):34-47.
184. Clauw DJ. Fibromyalgia: a clinical review. *Jama*. 2014;311(15):1547-55.
185. Chinn S, Caldwell W, Gritsenko K. Fibromyalgia Pathogenesis and Treatment Options Update. *Current pain and headache reports*. 2016;20(4):25.
186. Besson JM. The neurobiology of pain. *Lancet*. 1999;353(9164):1610-5.
187. Woolf CJ. Central sensitization: implications for the diagnosis and treatment of pain. *Pain*. 2011;152(3 Suppl):S2-15.
188. Häuser W, Ablin J, Fitzcharles M, Littlejohn G, Luciano JV, Usui C, et al. Fibromyalgia. *Nat Rev Disease Primers*. 2015;1.
189. Choy EH. The role of sleep in pain and fibromyalgia. *Nature reviews Rheumatology*. 2015;11(9):513-20.
190. Giovengo SL, Russell IJ, Larson AA. Increased concentrations of nerve growth factor in cerebrospinal fluid of patients with fibromyalgia. *The Journal of rheumatology*. 1999;26(7):1564-9.
191. Russell IJ, Orr MD, Littman B, Vipraio GA, Albourek D, Michalek JE, et al. Elevated cerebrospinal fluid levels of substance P in patients with the fibromyalgia syndrome. *Arthritis and rheumatism*. 1994;37(11):1593-601.
192. Sarchielli P, Di Filippo M, Nardi K, Calabresi P. Sensitization, glutamate, and the link between migraine and fibromyalgia. *Current pain and headache reports*. 2007;11(5):343-51.
193. Harris RE, Sundgren PC, Craig AD, Kirshenbaum E, Sen A, Napadow V, et al. Elevated insular glutamate in fibromyalgia is associated with experimental pain. *Arthritis and rheumatism*. 2009;60(10):3146-52.
194. Russell IJ, Vaeroy H, Javors M, Nyberg F. Cerebrospinal fluid biogenic amine metabolites in fibromyalgia/fibrositis syndrome and rheumatoid arthritis. *Arthritis and rheumatism*. 1992;35(5):550-6.
195. Schmidt-Wilcke T, Clauw DJ. Fibromyalgia: from pathophysiology to therapy. *Nature reviews Rheumatology*. 2011;7(9):518-27.
196. Baraniuk JN, Whalen G, Cunningham J, Clauw DJ. Cerebrospinal fluid levels of opioid peptides in fibromyalgia and chronic low back pain. *BMC musculoskeletal disorders*. 2004;5:48.
197. Harris RE, Clauw DJ, Scott DJ, McLean SA, Gracely RH, Zubieta JK. Decreased central mu-opioid receptor availability in fibromyalgia. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2007;27(37):10000-6.
198. Lucas HJ, Brauch CM, Settas L, Theoharides TC. Fibromyalgia--new concepts of pathogenesis and treatment. *International journal of immunopathology and pharmacology*. 2006;19(1):5-10.

199. Blanco I, Beritze N, Arguelles M, Carcaba V, Fernandez F, Janciauskiene S, et al. Abnormal overexpression of mastocytes in skin biopsies of fibromyalgia patients. *Clinical rheumatology*. 2010;29(12):1403-12.
200. Torresani C, Bellafiore S, De Panfilis G. Chronic urticaria is usually associated with fibromyalgia syndrome. *Acta dermato-venereologica*. 2009;89(4):389-92.
201. Theoharides TC, Tsilioni I, Arbetman L, Panagiotidou S, Stewart JM, Gleason RM, et al. Fibromyalgia, a syndrome in search of pathogenesis and therapy. *The Journal of pharmacology and experimental therapeutics*. 2015.
202. McLean SA, Williams DA, Stein PK, Harris RE, Lyden AK, Whalen G, et al. Cerebrospinal fluid corticotropin-releasing factor concentration is associated with pain but not fatigue symptoms in patients with fibromyalgia. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology*. 2006;31(12):2776-82.
203. Aad G, Abbott B, Abdallah J, Abdelalim AA, Abdesselam A, Abdinov O, et al. Search for dilepton resonances in pp collisions at radicals=7 TeV with the ATLAS detector. *Physical review letters*. 2011;107(27):272002.
204. Littlejohn G. Neurogenic neuroinflammation in fibromyalgia and complex regional pain syndrome. *Nature reviews Rheumatology*. 2015.
205. Milligan ED, Watkins LR. Pathological and protective roles of glia in chronic pain. *Nature reviews Neuroscience*. 2009;10(1):23-36.
206. Uceyler N, Hauser W, Sommer C. Systematic review with meta-analysis: cytokines in fibromyalgia syndrome. *BMC musculoskeletal disorders*. 2011;12:245.
207. Rodriguez-Pinto I, Agmon-Levin N, Howard A, Shoenfeld Y. Fibromyalgia and cytokines. *Immunology letters*. 2014;161(2):200-3.
208. Wang H, Moser M, Schiltenwolf M, Buchner M. Circulating cytokine levels compared to pain in patients with fibromyalgia -- a prospective longitudinal study over 6 months. *The Journal of rheumatology*. 2008;35(7):1366-70.
209. Ernberg M, Christidis N, Ghafouri B, Bileviciute-Ljungar I, Lofgren M, Bjersing J, et al. Plasma Cytokine Levels in Fibromyalgia and Their Response to 15 Weeks of Progressive Resistance Exercise or Relaxation Therapy. *Mediators of inflammation*. 2018;2018:3985154.
210. Mendieta D, De la Cruz-Aguilera DL, Barrera-Villalpando MI, Becerril-Villanueva E, Arreola R, Hernandez-Ferreira E, et al. IL-8 and IL-6 primarily mediate the inflammatory response in fibromyalgia patients. *Journal of neuroimmunology*. 2016;290:22-5.
211. Gur A, Karakoc M, Nas K, Remzi, Cevik, Denli A, et al. Cytokines and depression in cases with fibromyalgia. *The Journal of rheumatology*. 2002;29(2):358-61.
212. Wallace DJ, Linker-Israeli M, Hallegua D, Silverman S, Silver D, Weisman MH. Cytokines play an aetiopathogenetic role in fibromyalgia: a hypothesis and pilot study. *Rheumatology*. 2001;40(7):743-9.
213. Pay S, Calguneri M, Caliskaner Z, Dinc A, Apras S, Ertenli I, et al. Evaluation of vascular injury with proinflammatory cytokines, thrombomodulin and fibronectin in patients with primary fibromyalgia. *Nagoya journal of medical science*. 2000;63(3-4):115-22.
214. Bazzichi L, Rossi A, Massimetti G, Giannaccini G, Giuliano T, De Feo F, et al. Cytokine patterns in fibromyalgia and their correlation with clinical manifestations. *Clinical and experimental rheumatology*. 2007;25(2):225-30.
215. Kadetoff D, Lampa J, Westman M, Andersson M, Kosek E. Evidence of central inflammation in fibromyalgia-increased cerebrospinal fluid interleukin-8 levels. *Journal of neuroimmunology*. 2012;242(1-2):33-8.
216. Geiss A, Rohleider N, Anton F. Evidence for an association between an enhanced reactivity of interleukin-6 levels and reduced glucocorticoid sensitivity in patients with fibromyalgia. *Psychoneuroendocrinology*. 2012;37(5):671-84.

217. Pernambuco AP, Schetino LP, Alvim CC, Murad CM, Viana RS, Carvalho LS, et al. Increased levels of IL-17A in patients with fibromyalgia. *Clinical and experimental rheumatology*. 2013;31(6 Suppl 79):S60-3.
218. Uceyler N, Kewenig S, Kafke W, Kittel-Schneider S, Sommer C. Skin cytokine expression in patients with fibromyalgia syndrome is not different from controls. *BMC neurology*. 2014;14(1):185.
219. Uceyler N, Valenza R, Stock M, Schedel R, Sprotte G, Sommer C. Reduced levels of antiinflammatory cytokines in patients with chronic widespread pain. *Arthritis and rheumatism*. 2006;54(8):2656-64.
220. Sturgill J, McGee E, Menzies V. Unique cytokine signature in the plasma of patients with fibromyalgia. *Journal of immunology research*. 2014;2014:938576.
221. Yigit S, Inanir A, Tekcan A, Inanir S, Tural S, Ates O. Association between fibromyalgia syndrome and polymorphism of the IL-4 gene in a Turkish population. *Gene*. 2013;527(1):62-4.
222. Zhang Z, Cherryholmes G, Mao A, Marek C, Longmate J, Kalos M, et al. High plasma levels of MCP-1 and eotaxin provide evidence for an immunological basis of fibromyalgia. *Experimental biology and medicine*. 2008;233(9):1171-80.
223. Garcia JJ, Cidoncha A, Bote ME, Hinchado MD, Ortega E. Altered profile of chemokines in fibromyalgia patients. *Annals of clinical biochemistry*. 2014;51(Pt 5):576-81.
224. Garcia JJ, Carvajal-Gil J, Guerrero-Bonmatty R. Altered release of chemokines by phagocytes from fibromyalgia patients: a pilot study. *Innate immunity*. 2016;22(1):3-8.
225. Nuesch E, Hauser W, Bernardy K, Barth J, Juni P. Comparative efficacy of pharmacological and non-pharmacological interventions in fibromyalgia syndrome: network meta-analysis. *Annals of the rheumatic diseases*. 2013;72(6):955-62.
226. Thorpe J, Shum B, Moore RA, Wiffen PJ, Gilron I. Combination pharmacotherapy for the treatment of fibromyalgia in adults. *The Cochrane database of systematic reviews*. 2018;2:CD010585.
227. Bernardy K, Klose P, Busch AJ, Choy EH, Hauser W. Cognitive behavioural therapies for fibromyalgia. *The Cochrane database of systematic reviews*. 2013;9:CD009796.
228. Fitzcharles MA, Shir Y, Ablin JN, Buskila D, Amital H, Henningsen P, et al. Classification and clinical diagnosis of fibromyalgia syndrome: recommendations of recent evidence-based interdisciplinary guidelines. *Evidence-based complementary and alternative medicine : eCAM*. 2013;2013:528952.
229. Perrot S, Russell IJ. More ubiquitous effects from non-pharmacologic than from pharmacologic treatments for fibromyalgia syndrome: a meta-analysis examining six core symptoms. *European journal of pain*. 2014;18(8):1067-80.
230. Ang DC, Jensen MP, Steiner JL, Hilligoss J, Gracely RH, Saha C. Combining cognitive-behavioral therapy and milnacipran for fibromyalgia: a feasibility randomized-controlled trial. *The Clinical journal of pain*. 2013;29(9):747-54.
231. Okifuji A, Hare BD. Management of fibromyalgia syndrome: review of evidence. *Pain and therapy*. 2013;2(2):87-104.
232. Rico-Villademoros F, Slim M, Calandre EP. Amitriptyline for the treatment of fibromyalgia: a comprehensive review. *Expert review of neurotherapeutics*. 2015;15(10):1123-50.
233. Arnold LM, Keck PE, Jr., Welge JA. Antidepressant treatment of fibromyalgia. A meta-analysis and review. *Psychosomatics*. 2000;41(2):104-13.
234. Arnold LM. Duloxetine and other antidepressants in the treatment of patients with fibromyalgia. *Pain medicine*. 2007;8 Suppl 2:S63-74.

235. Hauser W, Wolfe F, Tolle T, Uceyler N, Sommer C. The role of antidepressants in the management of fibromyalgia syndrome: a systematic review and meta-analysis. *CNS drugs.* 2012;26(4):297-307.
236. Diaz-Marsa M, Palomares N, Moron MD, Tajima K, Fuentes ME, Lopez-Ibor JJ, et al. Psychological factors affecting response to antidepressant drugs in fibromyalgia. *Psychosomatics.* 2011;52(3):237-44.
237. Dwight MM, Arnold LM, O'Brien H, Metzger R, Morris-Park E, Keck PE, Jr. An open clinical trial of venlafaxine treatment of fibromyalgia. *Psychosomatics.* 1998;39(1):14-7.
238. Sayar K, Aksu G, Ak I, Tosun M. Venlafaxine treatment of fibromyalgia. *The Annals of pharmacotherapy.* 2003;37(11):1561-5.
239. VanderWeide LA, Smith SM, Trinkley KE. A systematic review of the efficacy of venlafaxine for the treatment of fibromyalgia. *Journal of clinical pharmacy and therapeutics.* 2015;40(1):1-6.
240. Keller MB, Trivedi MH, Thase ME, Shelton RC, Kornstein SG, Nemerooff CB, et al. The Prevention of Recurrent Episodes of Depression with Venlafaxine for Two Years (PREVENT) Study: Outcomes from the 2-year and combined maintenance phases. *The Journal of clinical psychiatry.* 2007;68(8):1246-56.
241. Lenox-Smith AJ, Jiang Q. Venlafaxine extended release versus citalopram in patients with depression unresponsive to a selective serotonin reuptake inhibitor. *International clinical psychopharmacology.* 2008;23(3):113-9.
242. Perahia DG, Pritchett YL, Kajdasz DK, Bauer M, Jain R, Russell JM, et al. A randomized, double-blind comparison of duloxetine and venlafaxine in the treatment of patients with major depressive disorder. *Journal of psychiatric research.* 2008;42(1):22-34.
243. MacLean AJ, Schwartz TL. Tramadol for the treatment of fibromyalgia. *Expert review of neurotherapeutics.* 2015;15(5):469-75.
244. Peng X, Robinson RL, Mease P, Kroenke K, Williams DA, Chen Y, et al. Long-term evaluation of opioid treatment in fibromyalgia. *The Clinical journal of pain.* 2015;31(1):7-13.
245. Sommer C, Hauser W, Alten R, Petzke F, Spath M, Tolle T, et al. [Drug therapy of fibromyalgia syndrome. Systematic review, meta-analysis and guideline]. *Schmerz.* 2012;26(3):297-310.
246. Russell IJ, Fletcher EM, Michalek JE, McBroom PC, Hester GG. Treatment of primary fibrositis/fibromyalgia syndrome with ibuprofen and alprazolam. A double-blind, placebo-controlled study. *Arthritis and rheumatism.* 1991;34(5):552-60.
247. Wolfe F, Zhao S, Lane N. Preference for nonsteroidal antiinflammatory drugs over acetaminophen by rheumatic disease patients: a survey of 1,799 patients with osteoarthritis, rheumatoid arthritis, and fibromyalgia. *Arthritis and rheumatism.* 2000;43(2):378-85.
248. Corrigan R, Derry S, Wiffen PJ, Moore RA. Clonazepam for neuropathic pain and fibromyalgia in adults. *The Cochrane database of systematic reviews.* 2012(5):CD009486.
249. Macfarlane GJ, Kronisch C, Dean LE, Atzeni F, Hauser W, Fluss E, et al. EULAR revised recommendations for the management of fibromyalgia. *Annals of the rheumatic diseases.* 2017;76(2):318-28.
250. Clauw DJ. Pharmacotherapy for patients with fibromyalgia. *The Journal of clinical psychiatry.* 2008;69 Suppl 2:25-9.
251. Moldofsky H, Lue FA, Mously C, Roth-Schechter B, Reynolds WJ. The effect of zolpidem in patients with fibromyalgia: a dose ranging, double blind, placebo controlled, modified crossover study. *The Journal of rheumatology.* 1996;23(3):529-33.
252. Corrigan R, Derry S, Wiffen PJ, Moore RA. Clonazepam for neuropathic pain and fibromyalgia in adults. *The Cochrane database of systematic reviews.* 2012;5:CD009486.

253. Holman AJ, Myers RR. A randomized, double-blind, placebo-controlled trial of pramipexole, a dopamine agonist, in patients with fibromyalgia receiving concomitant medications. *Arthritis and rheumatism*. 2005;52(8):2495-505.
254. Staud R. Sodium oxybate for the treatment of fibromyalgia. *Expert opinion on pharmacotherapy*. 2011;12(11):1789-98.
255. Russell IJ, Holman AJ, Swick TJ, Alvarez-Horine S, Wang YG, Guinta D, et al. Sodium oxybate reduces pain, fatigue, and sleep disturbance and improves functionality in fibromyalgia: results from a 14-week, randomized, double-blind, placebo-controlled study. *Pain*. 2011;152(5):1007-17.
256. Clauw DJ. Fibromyalgia and related conditions. *Mayo Clinic proceedings*. 2015;90(5):680-92.
257. Uceyler N, Sommer C, Walitt B, Hauser W. Anticonvulsants for fibromyalgia. *The Cochrane database of systematic reviews*. 2013;10:CD010782.
258. Goldenberg DL, Clauw DJ, Fitzcharles MA. New concepts in pain research and pain management of the rheumatic diseases. *Seminars in arthritis and rheumatism*. 2011;41(3):319-34.
259. Skrabek RQ, Galimova L, Ethans K, Perry D. Nabilone for the treatment of pain in fibromyalgia. *The journal of pain : official journal of the American Pain Society*. 2008;9(2):164-73.
260. Walitt B, Klose P, Fitzcharles MA, Phillips T, Hauser W. Cannabinoids for fibromyalgia. *The Cochrane database of systematic reviews*. 2016;7:CD011694.
261. Ware MA, Fitzcharles MA, Joseph L, Shir Y. The effects of nabilone on sleep in fibromyalgia: results of a randomized controlled trial. *Anesthesia and analgesia*. 2010;110(2):604-10.
262. Lynch ME, Campbell F. Cannabinoids for treatment of chronic non-cancer pain; a systematic review of randomized trials. *British journal of clinical pharmacology*. 2011;72(5):735-44.
263. Holtermann A, Gronlund C, Roeleveld K, Gerdle B. The relation between neuromuscular control and pain intensity in fibromyalgia. *Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology*. 2011;21(3):519-24.
264. Srikuea R, Symons TB, Long DE, Lee JD, Shang Y, Chomentowski PJ, et al. Association of fibromyalgia with altered skeletal muscle characteristics which may contribute to postexertional fatigue in postmenopausal women. *Arthritis and rheumatism*. 2013;65(2):519-28.
265. Bonaterra GA, Then H, Oezel L, Schwarzbach H, Ocker M, Thieme K, et al. Morphological Alterations in Gastrocnemius and Soleus Muscles in Male and Female Mice in a Fibromyalgia Model. *PloS one*. 2016;11(3):e0151116.
266. Oezel L, Then H, Jung AL, Jabari S, Bonaterra GA, Wissniowski TT, et al. Fibromyalgia syndrome: metabolic and autophagic processes in intermittent cold stress mice. *Pharmacology research & perspectives*. 2016;4(5):e00248.
267. Anderberg UM, Liu Z, Berglund L, Nyberg F. Plasma levels on nociceptin in female fibromyalgia syndrome patients. *Zeitschrift fur Rheumatologie*. 1998;57 Suppl 2:77-80.

## ANEXO A – Aprovação da CEUA



Pontifícia Universidade Católica do Rio Grande do Sul  
PRÓ-REITORIA DE PESQUISA, INovação e DESENVOLVIMENTO  
COMISSÃO DE ÉTICA NO USO DE ANIMAIS

Ofício 98/2015 - CEUA

Porto Alegre, 03 de dezembro de 2015.

Prezado Sr(a). Pesquisador(a),

A Comissão de Ética no Uso de Animais da PUCRS apreciou e aprovou seu Protocolo de Pesquisa, registro CEUA 15/00487 intitulado **"Caracterização do sistema nociceptina/orfanina FG-receptor NOP na modulação da fibromialgia experimental"**.

Sua investigação, respeitando com detalhe as descrições contidas no projeto e formulários avalidados pela CEUA, está autorizada a partir da presente data.

Informamos que é necessário o encaminhamento de relatório final quando finalizar esta investigação. Adicionalmente, ressaltamos que conforme previsto na Lei no. 11.794, de 08 de outubro de 2008 (Lei Arouca), que regulamenta os procedimentos para o uso científico de animais, é função da CEUA zelar pelo cumprimento dos procedimentos informados, realizando inspeções periódicas nos locais de pesquisa.

Nº de Animais	Espécie	Duração do Projeto
548	Mus musculus	03/2015 – 03/2019

Atenciosamente,

Prof. Dr. João Batista Blessmann Weber  
 Coordenador da CEUA/PUCRS

Ilma. Sra.

Profa. Dra. Maria Martha Campos

FABIO

## ANEXO B – Aceite do manuscrito – artigo em produção

<https://www.editorialmanager.com/pain/default.aspx>

Action	Manuscript Number	Article Title	Initial Date Submitted	Final Decision Date
	PAIN-D-18-00973R2	Nociceptin/orphanin FQ receptor modulates painful and fatigue symptoms in a mouse model of fibromyalgia	Oct 02, 2018	Jan 25, 2019

De: [em.pain.0.60d518.7fa7f1b9@editorialmanager.com](mailto:em.pain.0.60d518.7fa7f1b9@editorialmanager.com) <[em.pain.0.60d518.7fa7f1b9@editorialmanager.com](mailto:em.pain.0.60d518.7fa7f1b9@editorialmanager.com)> em nome de Pain

<[em@editorialmanager.com](mailto:em@editorialmanager.com)>

Enviado: sexta-feira, 25 de janeiro de 2019 11:17:47

Para: Maria Martha Campos

Assunto: Decision on Your Submission: PAIN-D-18-00973R2

Journal: PAIN

Title: Nociceptin/orphanin FQ receptor modulates painful and fatigue symptoms in a mouse model of fibromyalgia

ID: PAIN-D-18-00973R2

Format: Research Paper

Authors: Ana Paula Aquistapace Dagnino, Master; Rodrigo Braccini Madeira da Silva, PhD; Pedro Cesar Chagastelles, PhD; Talita Carneiro Brandão Pereira, PhD; Gianina Venturin Teribebe, PhD; Samuel Greggio, PhD; Jaderson Costa da Costa, PhD; Maurício Reis Bogo, PhD; Maria Martha Campos, Ph.D.

Dear Dr Campos,

I am pleased to inform you that your manuscript has been accepted for publication in PAIN®.

Your submission is being forwarded to our publishing partner, Wolters Kluwer, who will contact you shortly with information regarding proofs, images, and any other questions you might have.

**Please Note:** All accepted articles will be posted online within 7 business days of release to Wolters Kluwer (unless your article is held pending receipt of a commentary or an Editor's Choice video). This posted version of the article will be a PDF of your accepted files, so it will not be typeset at this juncture. Therefore, the text may contain typos or minor inaccuracies present in the accepted manuscript. (Corrections will be made at the proofing stage.) Importantly, the PDF of your accepted article will be submitted to PubMed, and will be fully citable. Supplementary material, such as raw data, videos, etc., will not be included at this stage. Supplementary materials will be included when the article is typeset, and will be present in the final corrected article when it is published in an issue. The issue publication will replace the accepted manuscript online upon publication.

<https://outlook.live.com/mail/senditemsmail/AMQkADAwATYwMAI0GQxOS00NABmMy0wMAIMDAKALEYAAN%2FZRJViIAjToJc%2FMdyuJzBwChWdL%2FBC58Tp6D9lmZKkUAAAACQkAAACHWdL%2FBC58Tp...> 1/3

08/02/2019

Email – ana paula dagnino – Outlook

For authors who have PATENT CONCERNS or PRESS RELEASE PLANS, please advise the editorial office ([painj@asp-pain.org](mailto:painj@asp-pain.org)) immediately if either of these issues would have implications for immediate online posting of your manuscript.

#### OPEN ACCESS

If you indicated in the revision stage that you would like your submission, if accepted, to be made open access, please go directly to step 2. If you have not yet indicated that you would like your accepted article to be open access, please follow the steps below to complete the process:

1. Notify the journal office via email that you would like this article to be available open access. Please send your Email to [painjournal@asp-pain.org](mailto:painjournal@asp-pain.org). Please include your article title and manuscript number.
2. A License to Publish (LTP) form must be completed for your submission to be made available open access. Please download the form from <http://links.lww.com/LWW-FS/A49>, sign it, and Email the completed form to the journal office.
3. Within 48 hours of receiving this e-mail: Go to <http://wolterskluwer.qconnect.com> to pay for open access. You will be asked for the following information. Please enter exactly as shown:
  - a. Article Title - Nociceptin/orphanin FQ receptor modulates painful and fatigue symptoms in a mouse model of fibromyalgia
  - b. Manuscript Number - PAIN-D-18-00973R2

If you have any questions for the publisher, please contact:

Jennie Kiniry  
 Journal Production Editor  
 Medical Research  
 Wolters Kluwer  
 410-528-4465 tel  
[jennie.kiniry@wolterskluwer.com](mailto:jennie.kiniry@wolterskluwer.com)

Also, we are seeking suitable cover images for PAIN. If you are interested in submitting an image for our consideration, please let us know by contacting our editorial office at [painj@asp-pain.org](mailto:painj@asp-pain.org). Please note that for the cover we are seeking images that either reflect the science in your article or are more art than science. While we cannot assure that one of your suggestions will be accepted for the cover, we do appreciate contributions.

#### Article Level Metrics

PAIN is excited to announce the addition of individual article-level metrics to the PAIN website, allowing authors new tools to help evaluate readers' interest in content. Simply find an article on the site and look below the right-hand tool bar where it reads "Article Level Metrics." By clicking the badge located there, you will find metrics including mentions in social media (Tweets, Facebook posts, etc.), news stories, and more. We hope readers and authors enjoy this new feature which provides a more immediate view of the impact of an article in the wider community.

<https://outlook.live.com/mail/sentitems/fd/AQMkADAwATYwMAiOGQxO600NABmMy0wMAiIMDAKAEYAAAN%2FZRJVIAlToJc%2FMDyiuJzBwChWdL%2FBC58Tp6D9ImLZkUAAAACAcIAAAACHWdL%2FBC58Tp...> 2/3

08/02/2019

Email – ana paula dagnino – Outlook

Thank you for sending us your paper. I hope you will consider sending us future studies as well.

Sincerely yours,

Francis J. Keefe, PhD  
 Editor-in-Chief, PAIN

---

*In compliance with data protection regulations, please contact the publication office if you would like to have your personal information removed from the database.*

<https://outlook.live.com/mail/sentitems/fd/AQMkADAwATYwMAiOGQxO600NABmMy0wMAiIMDAKAEYAAAN%2FZRJVIAlToJc%2FMDyiuJzBwChWdL%2FBC58Tp6D9ImLZkUAAAACAcIAAAACHWdL%2FBC58Tp...> 3/3



Pontifícia Universidade Católica do Rio Grande do Sul  
Pró-Reitoria de Graduação  
Av. Ipiranga, 6681 - Prédio 1 - 3º. andar  
Porto Alegre - RS - Brasil  
Fone: (51) 3320-3500 - Fax: (51) 3339-1564  
E-mail: [prograd@pucrs.br](mailto:prograd@pucrs.br)  
Site: [www.pucrs.br](http://www.pucrs.br)